



**CHEM-E1150**  
**BIOMASS PRETREATMENT AND FRACTIONATION**  
Theme 1: Kraft pulping, Module 1  
Raw Materials, Mechanical pretreatment

*Herbert Sixta, 2019*

**Outline Theme 1**

**Module 1: Raw materials and mechanical pretreatment**

**Module 2: Prehydrolysis**

**Module 3: Kraft cooking**

**Module 4. Screening, washing, bleaching and drying**

**Module 5: Pulp properties and uses**



## Module 1

1. Why pretreatment?
2. Lignocellulosic raw materials
3. Wood chipping

## Pretreatment vs Fractionation?

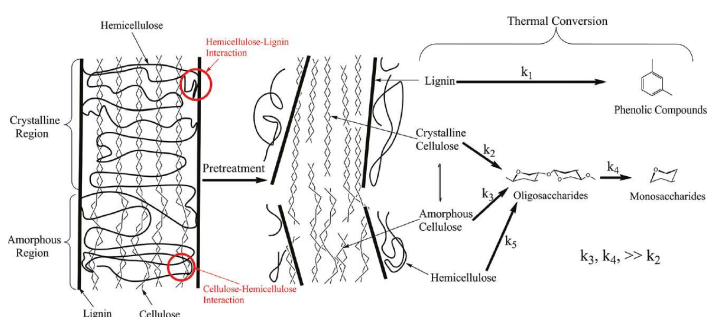
- **Pretreatment** focuses on the creation to **access** carbohydrates
- **Fractionation** focuses on the **separation** of lignocellulosic components

## Why pretreatment?

- **Hydrophobic lignin** forms a physical barrier
- **Recalcitrance of cellulose** owing to its crystalline structure
- **Hemicelluloses** form **sheets around fibrils**
- Low surface area and small capillaries inaccessible to enzymes

5

## Schematics of Pretreatment



- Prerequisite for **chemical** or **enzymatic** treatments of biomass.
- **Size reduction prior to** thermochemical processes

Zhang, Xiaolei et al. Energy&amp;Fuels, 2011, 25, 4786-4795

6

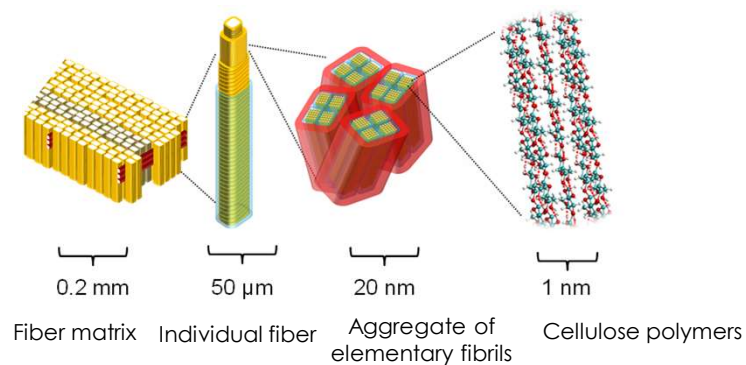
## Pretreatment Requirements

- High recovery of all carbohydrates
- High cellulose digestibility in subsequent enzymatic hydrolysis
- No or little sugar and lignin degradation products
- High sugar concentration in solution
- Low energy demand
- Low capital and operational costs

Galbe and Zacchi, Adv. Biochem. Eng./Biotechnol. (2007), 108, 41-65

7

## Hierarchical Structure of Lignocellulose



Lasse Tolonen: PhD thesis 2016

8



## Recalcitrance of lignocellulose

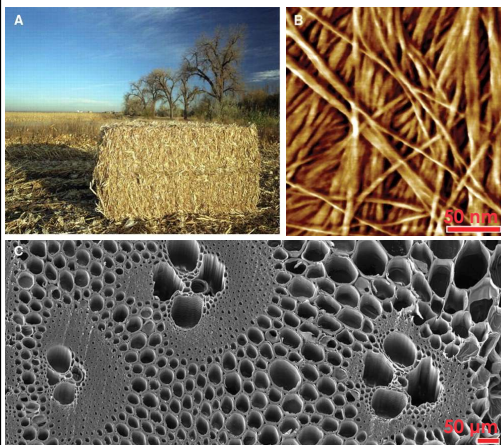
- Hydrophobic lignin forms a physical barrier
- **Recalcitrance of cellulose** owing to its crystalline structure
- **Hemicelluloses** form sheets around fibrils
- Low surface area and small capillaries inaccessible to enzymes:
  - Cellulose diameter ~5.1 nm (Arantes and Saddler, Biotechnol. Biofuels, 2011, 4:3, 1-16)
  - Lamellar space between spruce microfibril aggregates: ~10 nm (Fahlen and Salmen, Holzforschung, 2005, 59, 589-597)

Adriaan van Heiningen, lecture notes autumn 2009

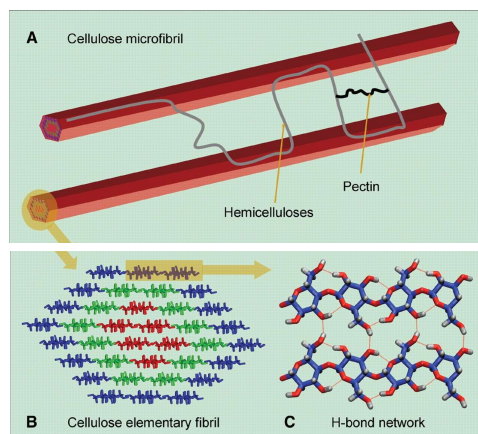
9

## Plant cell wall

### Fiber level



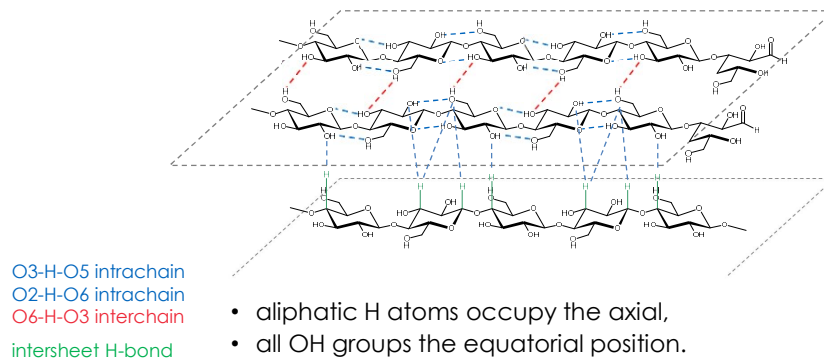
### Molecular level



Science, 2007: Vol. 315, Issue 5813, pp. 804-807. DOI: 10.1126/science.1137016

10

## Supramolecular structure of Cellulose



H. Sixto, M. Hummel, EPJCE 2011

11

## Crystal Systems

**Unit cell:** smallest repetitive volume which contains the complete lattice pattern of a crystal.

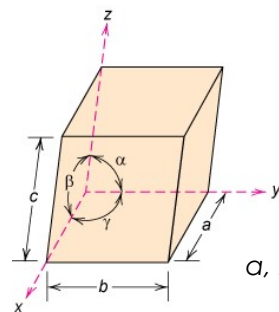


Fig. 3.4, Callister &amp; Rethwisch 8e.

7 crystal systems

14 crystal lattices

a, b, and c are the lattice constants

12

## Crystallographic Planes

- **Miller Indices:** Reciprocals of the (three) axial intercepts for a plane, cleared of fractions & common multiples. All parallel planes have same Miller indices.

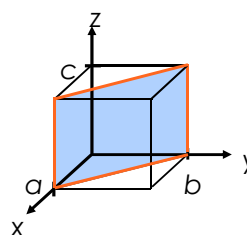
- **Algorithm**

1. Read off intercepts of plane with axes in terms of  $a, b, c$
2. Take reciprocals of intercepts
3. Reduce to smallest integer values
4. Enclose in parentheses, no commas i.e.  $(hkl)$

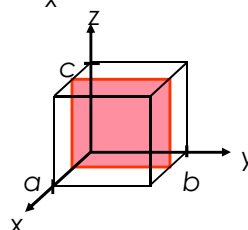
13

## Crystallographic Planes

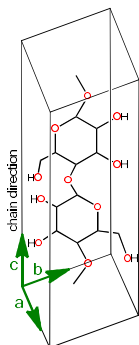
<u>example</u>	$a$	$b$	$c$
1. Intercepts	1	1	$\infty$
2. Reciprocals	$1/1$	$1/1$	$1/\infty$
	1	1	0
3. Reduction	1	1	0
4. Miller Indices	(110)		



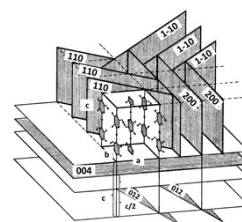
<u>example</u>	$a$	$b$	$c$
1. Intercepts	$1/2$	$\infty$	$\infty$
2. Reciprocals	$1/(1/2)$	$1/\infty$	$1/\infty$
	2	0	0
3. Reduction	2	0	0
4. Miller Indices	(200)		



## Unit Cell - Coordinate System



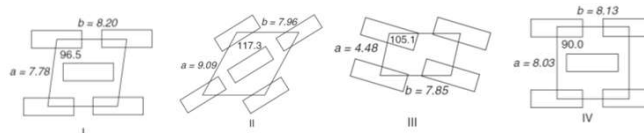
- **Simplest repeating unit in a crystal**, characterized by
- Vectors: a, b and c form the edges of a parallelepiped
- Angles:  $\alpha$  (b/c),  $\beta$  (a/c),  $\gamma$  (a/b)
- **Convention:**
  - c** cellulose chain axis;
  - b** forms the plane with c;
  - a** forms the distance between the sheets
- **Monoclinic:**  $a \neq b \neq c$ ,  $\alpha = \beta = 90^\circ \neq \gamma$
- **Triclinic:**  $a \neq b \neq c$ ,  $\alpha \neq \beta \neq \gamma$



Terminology (1973), p. 161 revised  
Cellulose [2013], 20: 20703-2718, DOI:10.1007/978-94-007-18

## Cellulose polymorphs

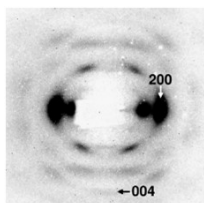
Poly-morph	Space group	Chains / unit cell	Chain direction	a [Å]	b [Å]	c [Å]	$\alpha$ [°]	$\beta$ [°]	$\gamma$ [°]
Ia	triclinic, P1	1	parallel	6.72	5.96	10.40	118.1	114.8	80.4
I $\beta$	Monoclinic, 2P1	2	parallel	7.78	8.20	10.38	90.0	90.0	96.5
II	Monoclinic, 2P1	2	antiparallel	8.10	9.08	10.36	90.0	90.0	117.3
III <sub>1</sub>	Monoclinic, 2P1	1	parallel	4.48	7.85	10.31	90.0	90.0	105.1
IV <sub>1</sub>	Triclinic, P1	2	parallel	8.03	8.13	10.4	90.0	90.0	90.0



Seong H. Kim et al. Korean J. Chem. Eng., [2013], 30(12), 2127-2141  
Nishiyama, Y., et al., J. Am. Chem. Soc., 2003, 125 p. 14300-14306.  
Nishiyama, Y., P. Langan, and H. Chanzy, J. Am. Chem. Soc., 2002, 124 p. 9074-9082.

Kobayashi, K., et al., Carbohydr. Polym., 2011, 86 (p. 975-981)  
Bellezza, G., et al., J. Phys. Chem. B, 2012, 116 p. 8031-8037

## Wide angle X-ray diffraction (WAXS)



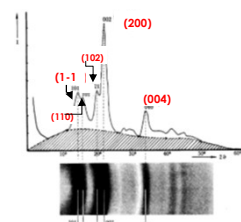
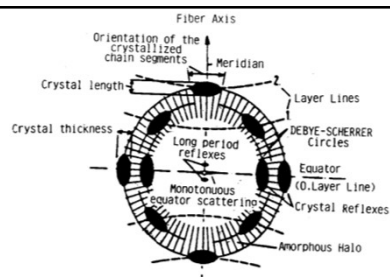
Cellulose I (flax)

Source:

CuKα X-ray

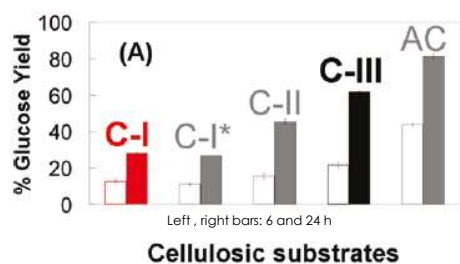
- Transmission geometry
- 2D detector
- Air scattering subtraction

Sawada Daisuke



Cotton linters, Temming, p160

## Effect of supramolecular structure on enzymatic hydrolysis?



AC > C-III > C-II > C-I

- C-I native cellulose Iβ
- C-II mercerized cellulose
- C-III liquid ammonia treated
- AC treated in concentrated phosphoric acid: amorphous cellulose

Chundawat, Shishir P. S. et al. JACS, 2011, 133, 11163–11174

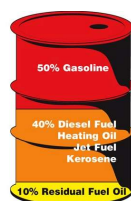
18

## Module 1

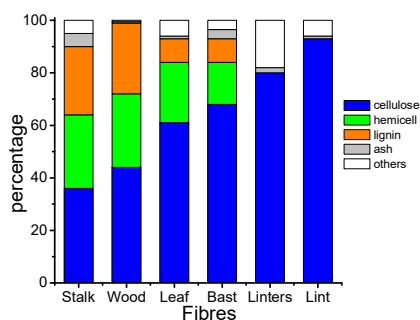
1. Why pretreatment?
2. Lignocellulosic raw materials
3. Wood chipping

## Elemental Composition of Feedstock

	Crude oil	Oil / Fat	Lignocellulose
C	85-90%	76 %	50 %
H	10-14%	13 %	6 %
O	0-1,5%	11 %	43 %

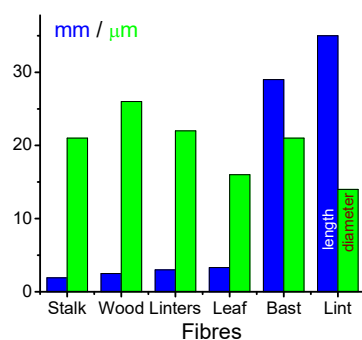


## Lignocellulosic Substrates



Stalk fibers: cereal straw, bamboo, sugar cane bagasse  
 Bast fibres: flax, hemp, jute, kenaf  
 Leaf fibres: abaca, sisal  
 Seed and fruit fibres: cotton lint, cotton linters  
 Wood fibres: soft-, hardwood

## Lignocellulosic Substrates



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## Wood, a Composite Material

Three Polymers:

### Cellulose

40 – 47 % **Iron**

Reinforcement Fibre, Microfibrils

### Hemicelluloses

matrix polymer between microfibrils

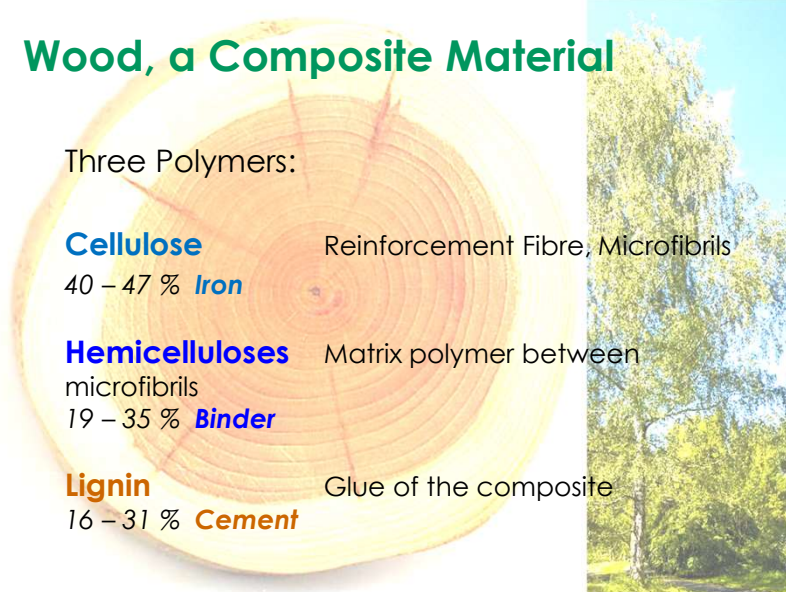
19 – 35 % **Binder**

Matrix polymer between microfibrils

### Lignin

16 – 31 % **Cement**

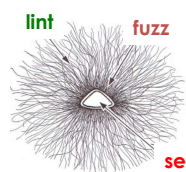
Glue of the composite



## Cotton Linters (seed fibers)

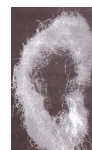


Open, ripe  
cotton ball  
*Gossypium\**



Cotton hair with  
lint and fuzz

Delinting



seed with  
linters

seed with fuzz

Temming Linters, 1973



## Bast fibers

Flax



Hemp



Jute



Temming Linters, 1973

## Stalk fibers



Bamboo fibers

## Leaf Fibers

Sisal



Abaca



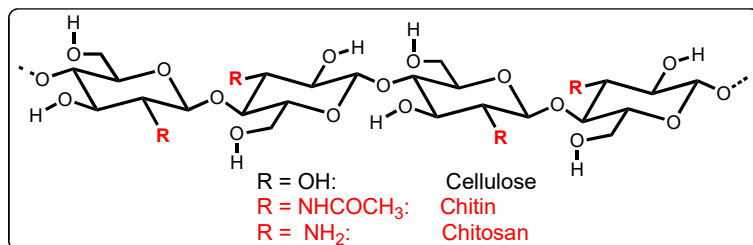
## Other polysaccharides

- Chitin / Chitosan
- Starch
- Pullulan
- Xanthan
- Dextrins, cyclodextrins
- Alginate

27

## Chitin, Chitosan

- Derived from shells of prawns and crabs
- **Chemistry:**
  - white to light-red solid powder, insoluble in water, soluble in organic acid



[http://www.ensymm.com/pdf/pharmaceutical/ensymm\\_chitosan\\_production\\_abstract.pdf](http://www.ensymm.com/pdf/pharmaceutical/ensymm_chitosan_production_abstract.pdf)

28

## Products derived of Chitosan

- **Products:**

- Cosmetic products: shampoos, moisturizer for the skin
- Additives to textile, pharmaceutical ingredients,
- Antibacterial, anti-fungal and anti-viral: wound dressing
- Food additive: dietary cookies, potato chips

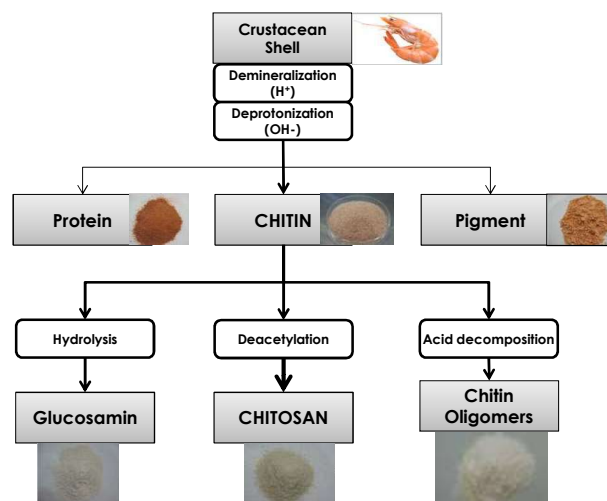
- **Price:**

- 20 – 1000 US\$/kg

[http://www.ensymm.com/pdf/pharmaceutical/ensymm\\_chitosan\\_production\\_abstract.pdf](http://www.ensymm.com/pdf/pharmaceutical/ensymm_chitosan_production_abstract.pdf)

29

## Chitin, Chitosan Production



30

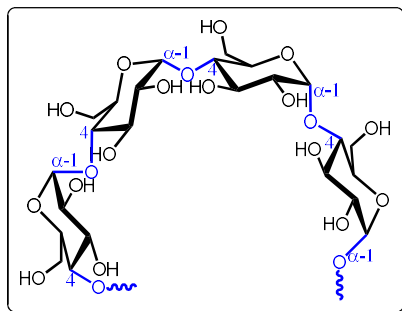
## Starch

- In cereal grains, potatoes, legumes (beans): amylose (linear) and amylopectin (branched).
- **1812**: First starch sugar plant in Weimar, Germany
- **1835**: **J.J. Berzelius**, Sweden, introduced enzymatic hydrolysis of starch into sugar and introduced the term „catalysis“.
- Consumption in Europe (2007):
  - 5.6 Mio t for food;
  - 3.7 Mio t for non-food (**paper**),
  - 1.9 Mio t for fuel ethanol.

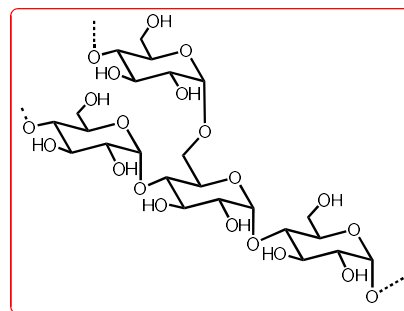
## Starch

Endproduct of photosynthesis in plant; available in wheat, rice, corn and potato.

**Amyloses** (20-30%) : helical polymer made of  $\alpha$ -D-glucose units, bound to each other through  $\alpha(1 \rightarrow 4)$  glycosidic bonds



**Amylopectin** (70-80%): branching with  $\alpha(1 \rightarrow 6)$  every 24 to 30 units; linear segments as double helix



## Thermoplastic starch (TPS)

- Native starch is not thermoplastic
- The H-bond interaction between starch molecules have to be reduced to melt-process native starch.
- TPS formed through the de-structuring of native starch granules by heating at relatively high temperatures, in high shear conditions, and with limited amounts of water→individual chains move freely.

33

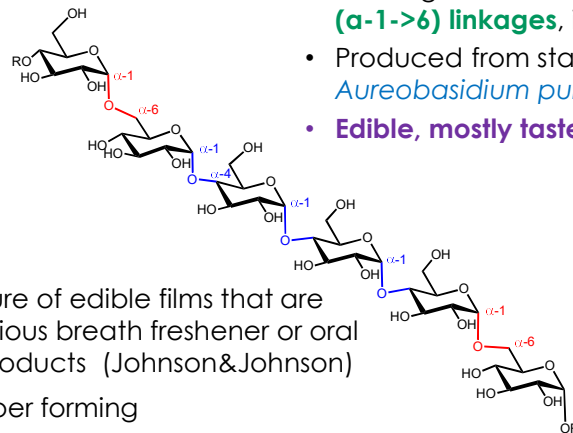
## Thermoplastic starch (TPS)

- **High amount of amylose preferred**, however
- Problem of retrogradation, a natural crystallization over time, leading to increased brittleness. Tendency of macro-molecules to form H-bonds during the expulsion of water.
- Can be avoided by the addition of plasticizer (water, glycerol,...)



34

## Pullulan

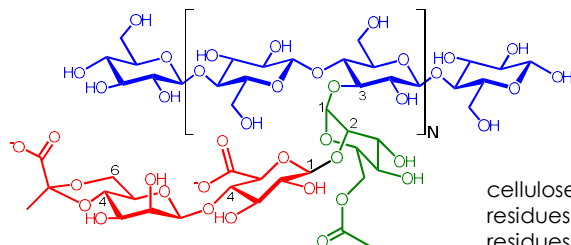


- Linear glucan containing  **$\alpha$ -(1 $\rightarrow$ 4)** and  **$\alpha$ -(1 $\rightarrow$ 6) linkages**, in 2:1 ratio
- Produced from starch by the fungus *Aureobasidium pullulans*.
- **Edible, mostly tasteless polymer.**
- Manufacture of edible films that are used in various breath freshener or oral hygiene products (Johnson&Johnson)
- Film and fiber forming

35

## Xanthan

- Microbial desiccation-resistant polymer, **>30,000 tons per year** by aerobic submerged fermentation from *Xanthomonas campestris* (E415)
- **In food** (gluten-free baking, emulsifier, ice cream), drilling mud; concrete additive



cellulose-like backbone of (1,4)-linked  $\beta$ -D-Glcp residues substituted at O-3 of alternate glucose residues, with a trisaccharide side chain:

$\alpha$ -(1 $\rightarrow$ 3)-glycosidic:  $\beta$ -D-mannopyranosyl-(1 $\rightarrow$ 4)- $\beta$ -D-glucuronopyranosyl-(1 $\rightarrow$ 2)-6-O-acetyl- $\alpha$ -D-manno-pyranosyl- side chain

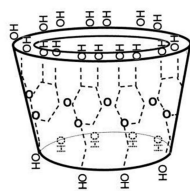
36

## Cyclodextrine

### Cyclic Oligosaccharides

Available from starch using cyclodextrin glucotransferase (CGTases), amylolytic enzymes produced by *Bacillus macerans*

The most common and commercially available cyclodextrins consist of **six**, **seven** and **eight** glucose units, **α**-, **β**- und **γ**-cyclodextrines.

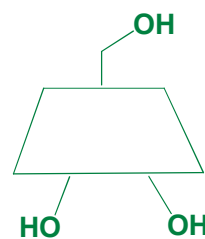
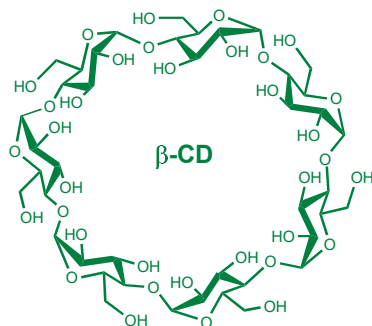


37

## Cyclodextrine

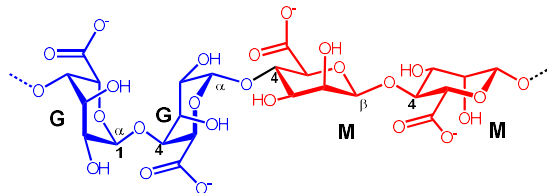
Conical molecules, with well defined cavities, **hydrophilic exteriors**, and more **hydrophobic interiors**.

Hydrophobic compounds entrapped in the cavities → **drug delivery system**



## Alginate

- Alginic acid is a linear copolymer with blocks of (1-4)-linked  **$\beta$ -D-mannuronate (M)** and its C-5 epimer  **$\alpha$ -L-guluronate (G)** residues.



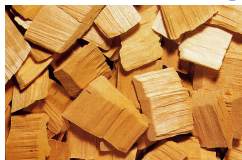
- In cell walls of brown algae (**seaweed**).
- Binding of water  $\rightarrow$  viscous gum. In extracted form it absorbs water quickly up to 200–300 times its weight. Thickening drinks and food

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39

## Module 1

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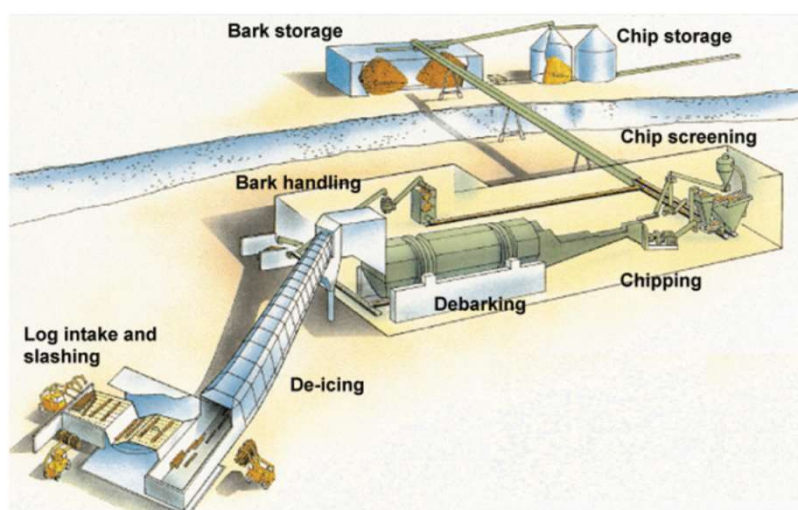




## Chip Preparation

1. Wood yard
2. Wood intake
3. Debarking
4. Chipping
5. Chip screening
6. Chip storage

### Wood yard



42

## Wood handling



Metso, 2003

Herbert Sixta

43

## Log Storage



PhD Thesis, A. Promberger, 2004

## Fungal Infestation

### (1) Heart Rot: Infestation along the Roots



5 cm distance



100 cm distance

### (2) Heart Rot: Formation during Storage



Deterioration zones along the rays

PhD Thesis, A. Promberger, 2004

## Dominating Fungi

### (1) *Schizophyllum commune*



### (2) *Chondrostereum pupureum*



PhD Thesis, A. Promberger, 2004

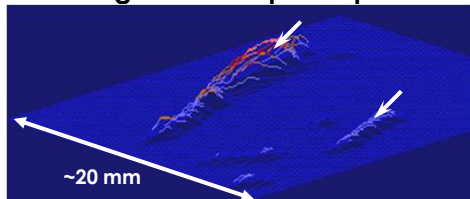
## Objective, Experimental

### Storage of Beech Logs



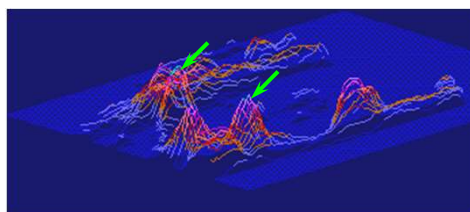
## Topochemical Characterization of Unbleached Sulfite Pulp

Scanning UV microspectrophotometry at  $\lambda_{278\text{nm}}$



**WET**

Fragments of a Middle Lamella.



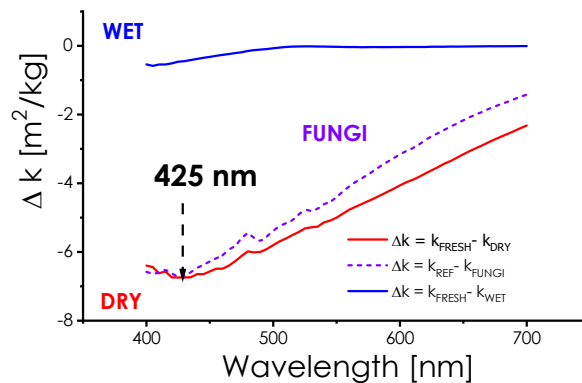
**DRY**

Polyphenolic compounds, attached to the cell wall and deposited in the lumina of parenchyma cells.

PhD Thesis, A. Promberger, 2004

## Chromophores in Unbleached Pulps

Absorption Difference Spectra



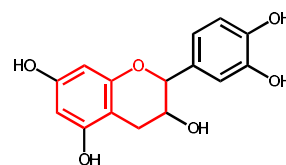
A. Promberger, PhD Thesis, 2004

## Chromophores in Unbleached Pulps

Pulps from **DRY** and **INFECTED** confirm the presence of strongly absorbing chromophores.

Minimum at 425 nm and pronounced tailing towards longer wavelengths indicates occurrence of highly conjugated compounds (**e.g. polyphenolic structures\***).

**WET** almost comparable to **FRESH**



A. Promberger, PhD Thesis, 2004

## Chip Preparation

1. Wood yard
- 2. Wood intake**
3. Debarking
4. Chipping
5. Chip screening
6. Chip storage

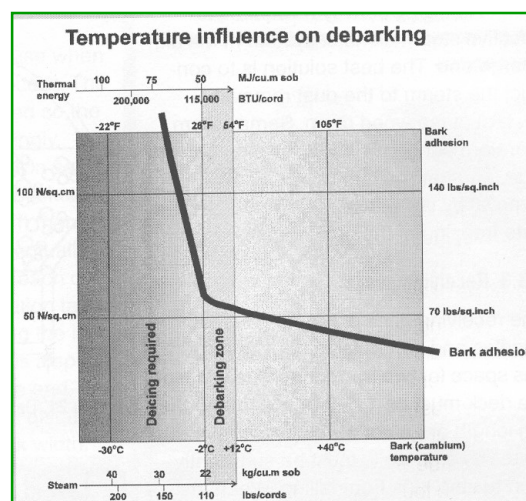
## Wood Intake

- Logs are transported from the forest by truck, train, ship,...
- The length of the logs varies between 3 - 5 m
- Part of wood supply maybe saw mill chips
- Logs are stored in the woodyard, or, in certain cases also in the forest.

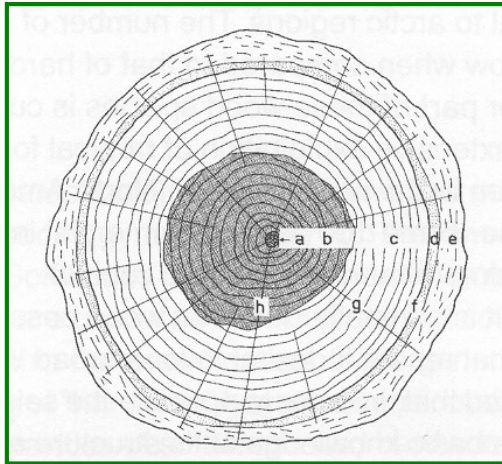
## De-icing

- Necessary to ensure debarking
- Sufficient to de-ice the surface
- Usually de-icing with hot water:
  - **50 – 100 MJ/m<sup>3</sup> softwood**
  - **80 – 140 MJ/m<sup>3</sup> birch**

## De-icing



## Cross section of wood stem



- a) Pith
- b) Heart wood
- c) Sap wood
- d) Inner Bark (Phloem)
- e) Outer Bark
- f) Cambium
- g) Secondary rays
- h) Primary rays

55

## Chip Preparation

1. Wood yard
2. Wood intake
- 3. Debarking**
4. Chipping
5. Chip screening
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## Debarking

- Efficient removal of bark since its **fiber yield is only 20%**
- **Shear forces** primarily act on the **radial layers** of the **cambial cells**.
- Depending on the species, moisture content, age, location, temperature, etc. the **bond strength** of the bark ~ **20 and 500 N/cm<sup>2</sup>**.
- Production of **chips with uniform dimensions** regarding length, width and thickness.

57

## Debarking efficiency

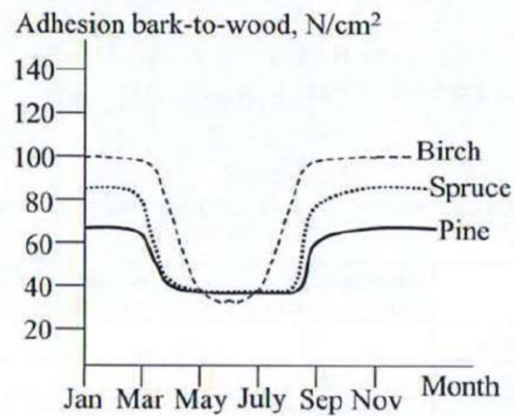
Bark thickness, bark structure affect the ease of debarking.

However, not the species, but the season has the most prominent influence on debarking efficiency.

Adhesion N/cm <sup>2</sup>	Debarking Efficiency	Wood Species
30	very easy	<b>Southern Pine, maple</b>
50	easy	hemlock, spruce, beech
90	difficult	elm, birch
130	very difficult	poplar
>200	almost impossible	<b>Hickory, limewood</b>

58

## Effect of Cutting Season on Debarking



E. Brännvall, *Pulping Chemistry and technology*, Vol 2, p. 16

## Factors affecting debarking degree

- Wood species and density
- Wood temperature
- Log length and diameter
- Drum filling degree
- Drum rotation speed
- Drum diameter and length

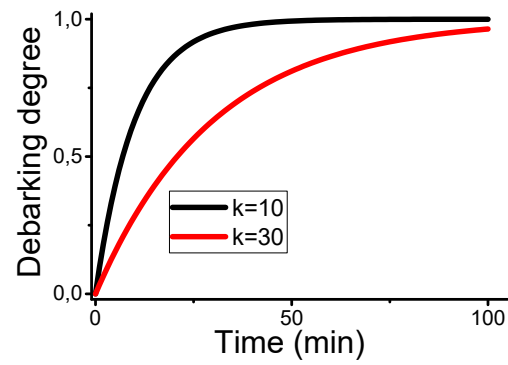
## Debarking Methods

- **Drum Debarker**
- Rotary or Cradle Debarker
- Ring Debarker

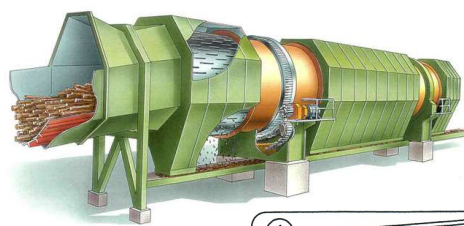
## Drum Debarking Parameters

- Rotation speed almost linearly affects its efficiency.
- 4 – 7 rpm at 5.5 m of diameter
- Degree of barking proportional to  $(1 - \text{Exp}(-t/k))$  with  
t = retention time, k = f(rpm, filling degree, etc.)
- **Wood loss linearly dependent on retention time and influenced by rotation speed.**

## Drum Debarking Parameters



## Rotary Parallel Drum Barker

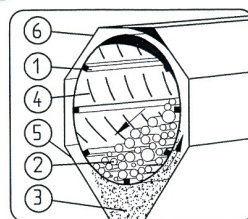


**Parallel drum for long logs:**

$D = 3 - 4.5 \text{ m}$ ,  $L = 40 - 60 \text{ m}$

**Tumble drum for short logs:**

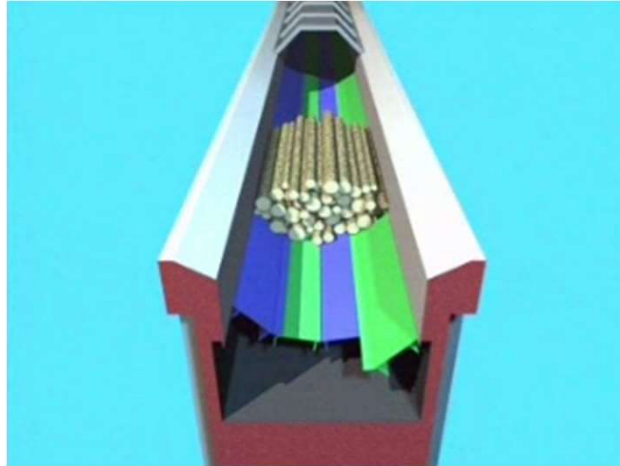
$D = 4 - 6 \text{ m}$ ,  $L = 20 - 40 \text{ m}$ ;



- |                       |                |
|-----------------------|----------------|
| 1. Log lifters        | 4. Bark slots  |
| 2. Logs               | 5. Bark chutes |
| 3. Bark, sand, stones | 6. Hood        |

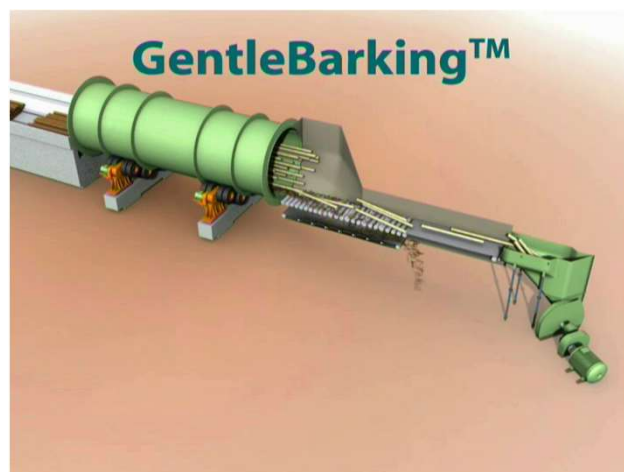
64

## Gentle Log Feed



65

## Rotary Parallel Drum Barker



66

## Rotary Debarker

- Assembly containing pair-wise openings in the bottom.
- Debarking plates are fixed on the surface of the rotors.
- Spinning plates hit the logs and bark removal occurs by breaking the fiber bonds at the cambium layer.



67

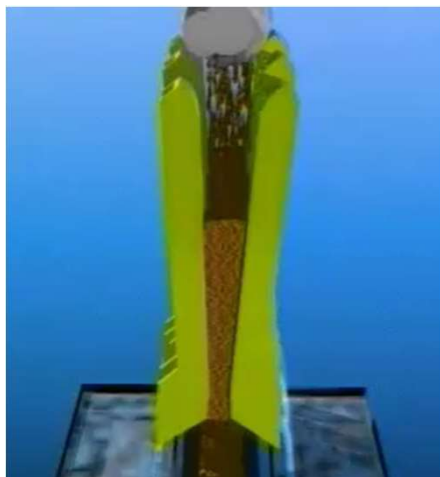
## Ring Debarker



Variable pressure air seal debarking ring (Ø 31...183 cm)

- Single stem process
- Counter rotating ring for stringy bark
- Cannot accept logs < 1.8 m
- Higher wood loss than rotary

## Bark Handling



- Long contact between water and bark impairs water removal
- Pressing improves water removal
- Uniform pressure over the whole area of the bark layer

## Utilization of Bark

Proportion of bark between 10 and 20 % (vol). Important energy source.

<u>Species</u>	<u>volume fraction of bark, %</u>
Pine	11.8
Spruce	12.8
Birch	14.1

## Utilization of Bark

### High Heating Value, HHV:

- 12.2 MJ/kg at 40% moisture content
- 8.1 MJ/kg at 60% moisture content
- 6.1 MJ/kg at 70% moisture content

$$HV_{wet} = HV_{dry} \cdot (1 - m_{water}) - H_{25} \cdot m_{water}$$

HV	heating value in MJ/kg fuel
$m_{water}$	mass of water in kg water/kg biomass
$H_{25}$	evaporation heat for water at 25°C = 2.44 MJ/kg water

**Bark is pressed and burnt in a fluidized bed incinerator.**

## Chip Preparation

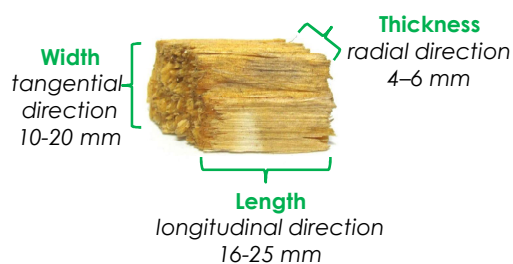
1. Wood yard
2. Wood intake
3. Debarking
- 4. Chipping**
5. Chip screening
6. Chip storage



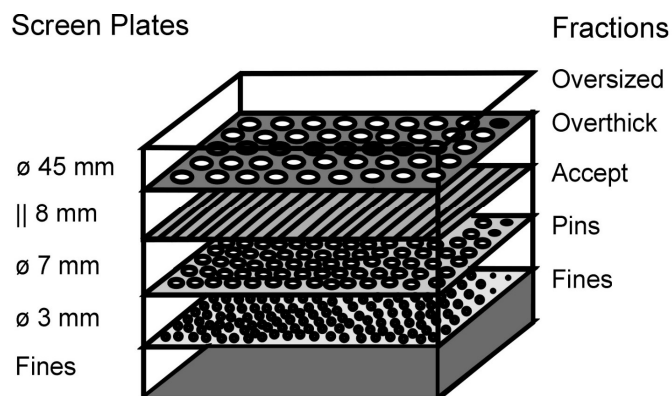
## Size Reduction Energy

	GJ/t
• <b>Wood chipping</b>	<b>0.1</b>
• Chips to saw dust by hammer mill	1.0
• Stone grinding	5.0
• Disc refining	7.0
• Wood to ethanol (0.24 t/t w)	7.2

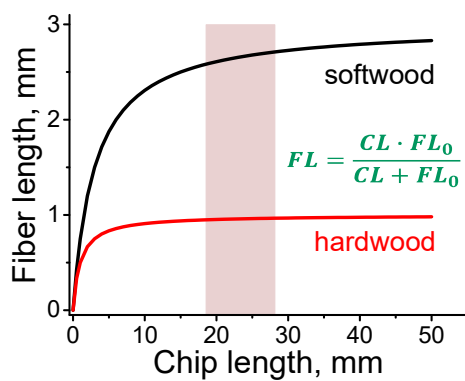
## (Target) Chip dimensions



## Measurement of Chip Size Distribution



## Chip length vs Fiber length

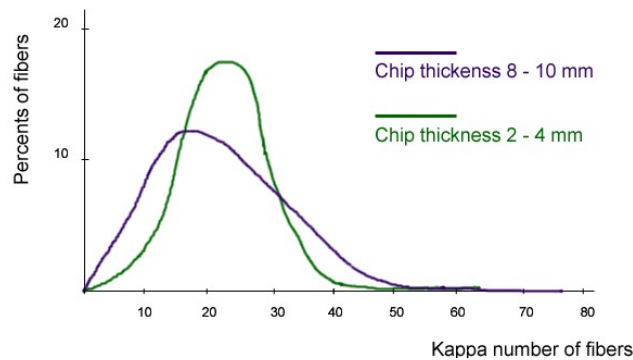


**Numbers of 3.5 mm long fibres in the longitudinal direction of a 25 mm long chip** are  $\sim 25/3.5 = \sim 7$ .

**Number of fibres in the radial direction of a 3 mm thick chip** would be  $\sim 3.0/0.03 = \sim 100$ .

## Effect of chip thickness on delignification

Kappa number (=lignin content) for both pulps is 20 (3%)



### CHIP DENSITY

$$\rho_{dc} = \frac{m_c}{V_c}$$

$$\rho_{dc} = \rho_s(1 - \varepsilon_i)$$

$$V_v = \frac{1}{\rho_{dc}} - \frac{1}{\rho_s}$$

$$\rho_s = 1.53 \text{ kg} \cdot \text{L}^{-1}$$

$\rho_i$  ... internal chip

$\varepsilon_i$  ... void volume fraction

### Wood densities

Wood species	Dry density, $\rho_{dc}$ , kg/L	Void volume, $V_v$ , L/kg
Aspen	0,37	2,05
Spruce	0,43	1,67
Pine	0,47	1,47
Beech	0,68	0,82

**EXAMPLE**

$$\rho_{dc} = 0.40 \text{ kg} \cdot \text{L}^{-1}$$

$$\varepsilon_i = 1 - \frac{\rho_{dc}}{1.53} = 1 - \frac{0.40}{1.53} = 0.74$$

**Total void volume:**

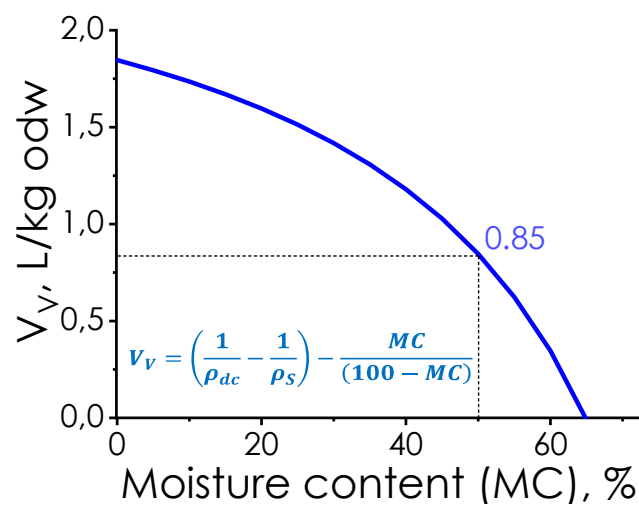
$$V_V = \frac{1}{0.40} - \frac{1}{1.53} = 1.85 \text{ L} \cdot \text{kg}^{-1}$$

**Volume of air inside chip at MC<sub>w</sub>=50%:**

$$MC_w = 50\% = \frac{m_{wc} - m_{dc}}{m_{wc}} = \frac{2 - 1}{2} = 0.5$$

$$V_V = 1.85 - 1.00 = 0.85 \text{ L} \cdot \text{kg}^{-1}$$

79



80

**PENETRATION :**  $P_0 = \frac{V_{i,H_2O}}{V_V}$

**Aspen chip:**

$$\rho_{dc} = 0.37 \text{ kg} \cdot \text{L}^{-1}$$

$$DS = \frac{m_{dc}}{m_{wc}} = 0.55$$

$$m_{wc} = \frac{1}{0.55} = 1.82 \text{ kg} \cdot \text{kg}^{-1} DS$$

$$m_{moisture} = 1.82 - 1.00 = 0.82 \text{ kg} \cdot \text{kg}^{-1} DS$$

$$V_V = \frac{1}{\rho_{dc}} - \frac{1}{\rho_S} = \frac{1}{0.37} - \frac{1}{1.53} = 2.05 \text{ L} \cdot \text{kg}^{-1} DS$$

$$P_0 = \frac{0.82}{2.05} \cdot 100 = 40\%$$

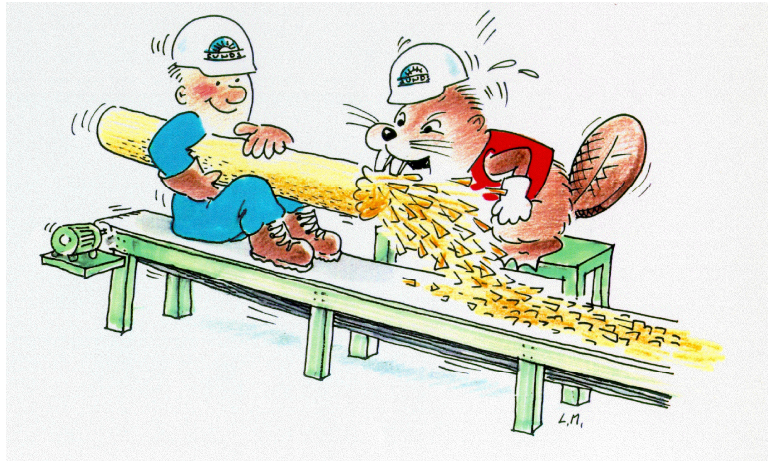
## Physical Properties of Chips

Comparison of chip dimensions and the spherical equivalent diameter (=diameter of a sphere of the same volume as a chip particle):

L	W	T	ED
	mm		mm
25	15	5	15,3
25	15	10	19,3
35	20	10	23,7

$$ED = \left( \frac{6 \cdot L \cdot W \cdot T}{\pi} \right)^{\frac{1}{3}}$$

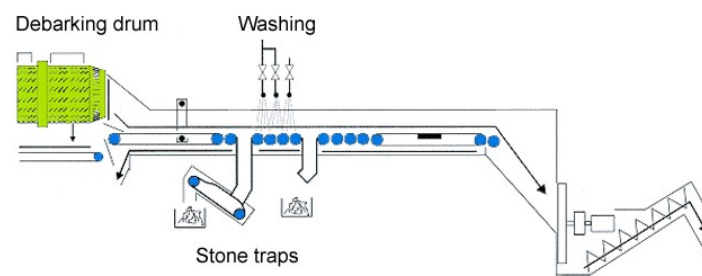
## Chip Production



## Transport To Chipper

- Alignment of the logs
- Removal of stones and other particulate impurities
- Removal of loose bark
- Log washing
- Sorting too short and too long logs
- Metal detection and removal

## Chipper Infeed Line



## Chippers

- **Disc Chipper**
- Drum Chipper

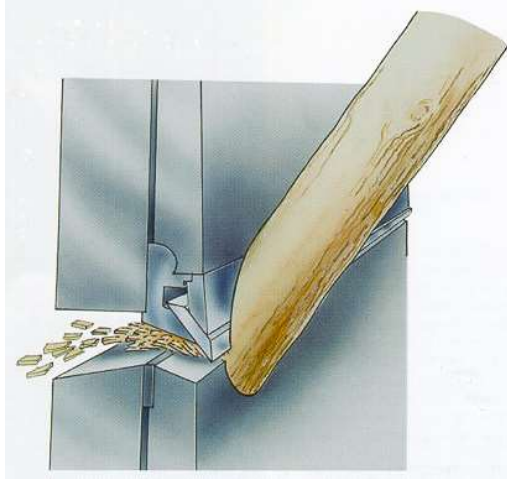
### Disk Chipper

- Most frequently used in pulp mills.
- Several straight knives (6 - 16) are mounted on a heavy, rigid disc in radial arrangement.
- Disc revolves in either a vertical or in a slanted plane.
- Generated chips pass through slots in the disc and may be discharged from the top, bottom or sides of the disc housing.

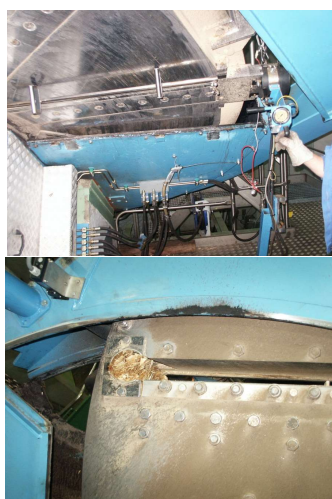
### Disk Chipper

- Supply of logs via an **infeed spout** at an **angle** between the face of the disc and the spout axis of about **30 - 40°**.
- When the knife edge cuts the wood, the fibres are more or less compressed in a longitudinal direction.





## Chippers



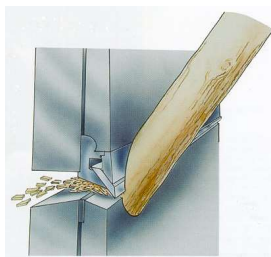
- Diameter of the disc 3.4 m
- Number of knives 10 – 14
- Capacity 450 m<sup>3</sup>/h (av)
- Velocity 3.8 m/s
- Moment of inertia 23 000 kg\*m<sup>2</sup>
- Total weight 33.5 t
- Twin-motor drive 2\*1000 kW

90

## Chipper



91

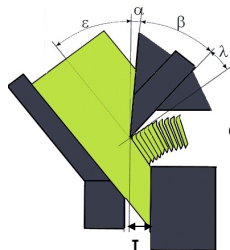


## Disk Chipper

- Generated chips pass through slots in the disc

- $L = \frac{T}{\sin \epsilon}$

- With increasing  $\epsilon$ , chip length,  $L$ , decreases.  $\epsilon$ -angle is typically 32-38°
- Chip thickness decreases with increasing  $\lambda$ .



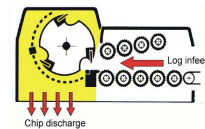
$L$  chip length  
 $T$  distance chipper/disc/knife  
 $\epsilon$  infeed angle (30 - 40°)  
 $\alpha$  pull-in angle = clearance angle (1-7°)  
 $\beta$  sharpness or knife angle (30 - 40°)  
 $\lambda$  complementary angle =  $90^\circ - (\alpha + \beta + \epsilon)$

92

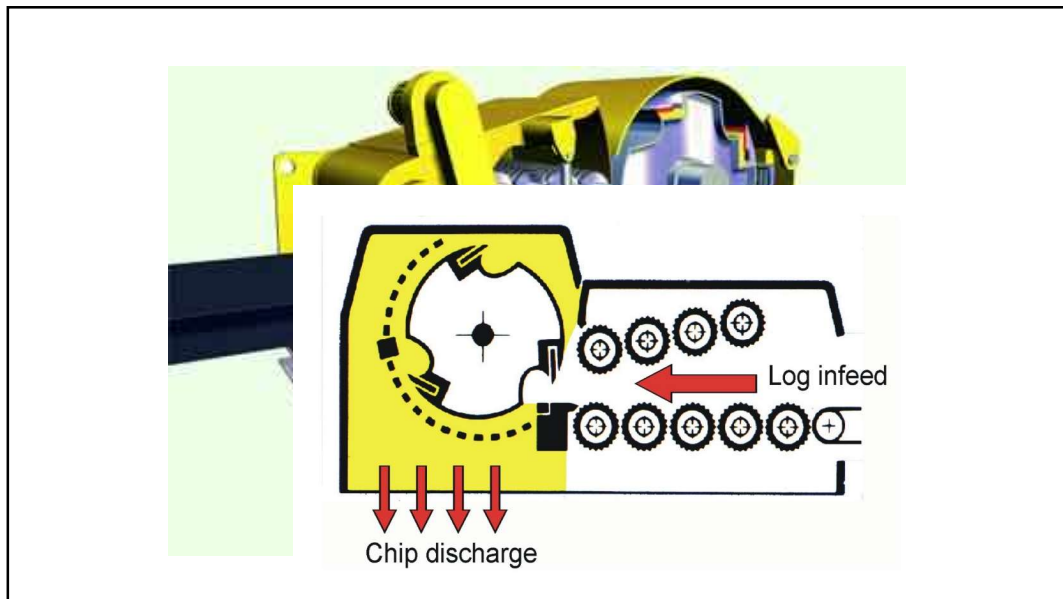
## Chippers

- Disc Chipper
- **Drum Chipper**

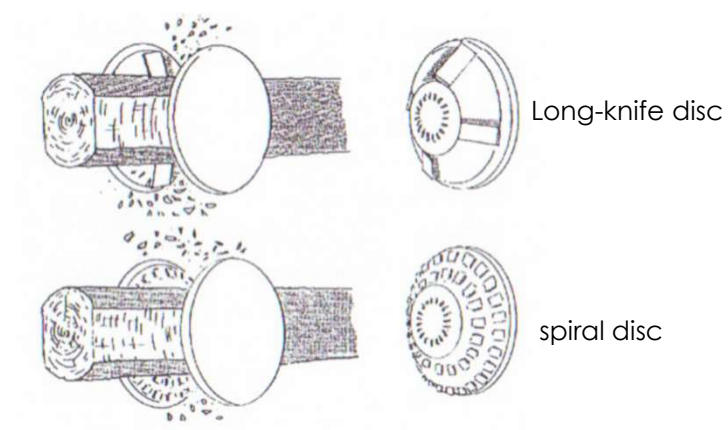
### Drum Chipper



- Preferred in sawmills (wood residues, etc.)
- Wood is **cut between the knife edge** and **an anvil** which is fixed at the bottom end of the infeed spout.
- Cut chips are caught in pockets, while larger pieces are further broken down by specially designed bridges.



## Chips from Sawmills



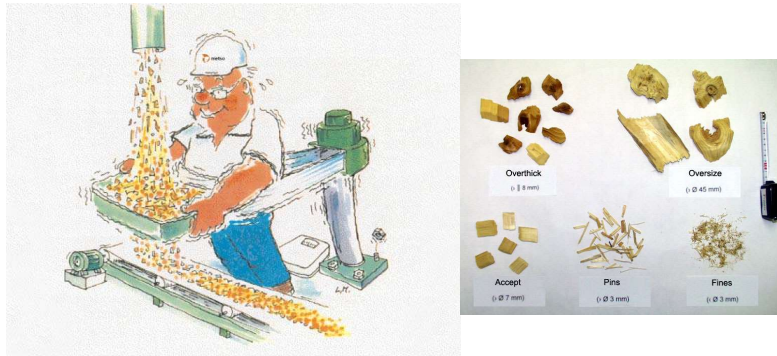
## Chips from Sawmills

- About 30% of the wood chips in Nordic countries derive from saw mills.
- In sawmills chipping is achieved while timber is reduced to blocks → **Reducers**.
- **Chip length is controlled by the log feed speed** and the **rotation speed** of the **reducer**.

## Chip Preparation

1. Wood yard
2. Wood intake
3. Debarking
4. Chipping
- 5. Chip screening**
6. Chip storage

## Chip Screening



99

## Chip Screening

Removal of oversize, overthicks, pins and fines:

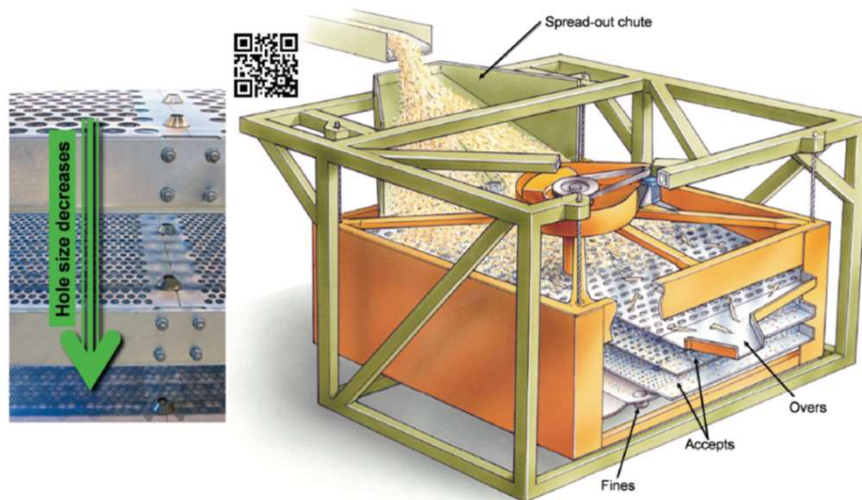
**Gyratory screening**

Perforated plates placed one above the other.

Chips are spread out on top plate and a vibrating motion shakes the chips.

100

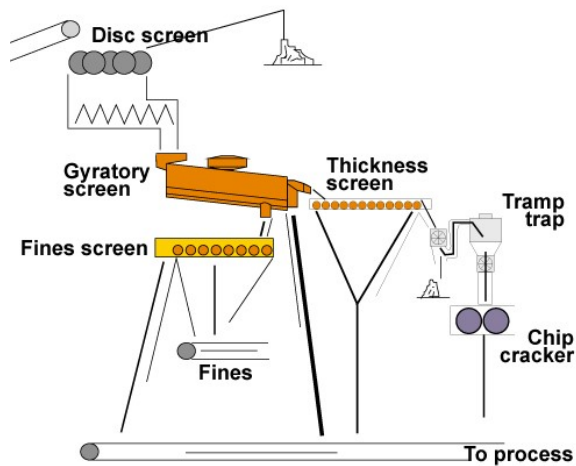
## Chip Screening (EasyScreen)



## Disc Screens

- Rotating shafts with discs mounted on them.
- The spacing between the shafts are the slots through which thin chips thin enough can escape.
- Used for removing overthick chips.

## Disc Screens



## Roll Screens

Pre-adjusted **space between adjoining rolls**.

**Fines** and **acceptable** chips pass these gaps at

- different travelling distances and are
- discharged from below the roll carpet by a conveyor



## Roll Screens

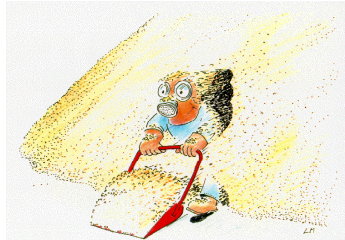


105

## Chip Preparation

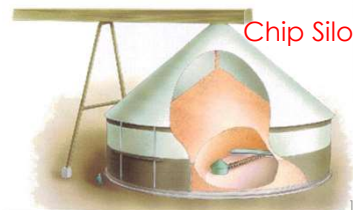
1. Wood yard
2. Wood intake
3. Debarking
4. Chipping
5. Chip screening
- 6. Chip storage**

## Chip storage



Continuous supply of high-quality chips.

Chip storage time  $\leq 3$  d



107

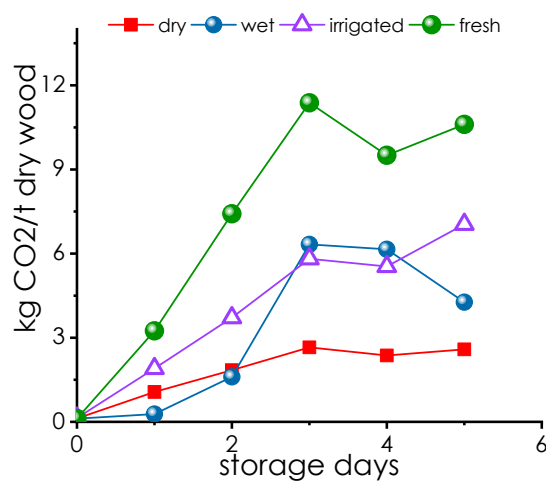
## Chip Storage

- Continuous supply of high-quality chips.
- **Chip storage time should not exceed 3 days:**
  - because of economic reason and
  - because of quality reason: hardwood chips are easily affected by **bacteria** and **fungi**.

## Chip Storage

- First-in first-out principle.
- Two forms of chip storage:
  - PILES
  - SILOS

## CO<sub>2</sub> formation upon Chip Storage



**Beech wood 30°C**

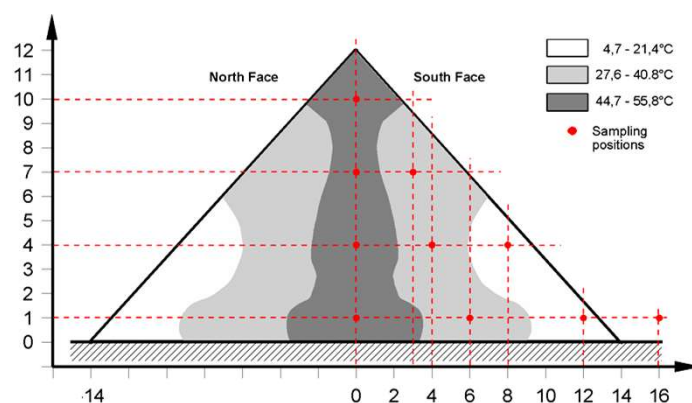
**Parenchyma cells**

in fresh cut chips are still alive and consume oxygen to oxidize the residual reserve polysaccharides (e.g. starch).

A. Promberger, PhD Thesis, 2004

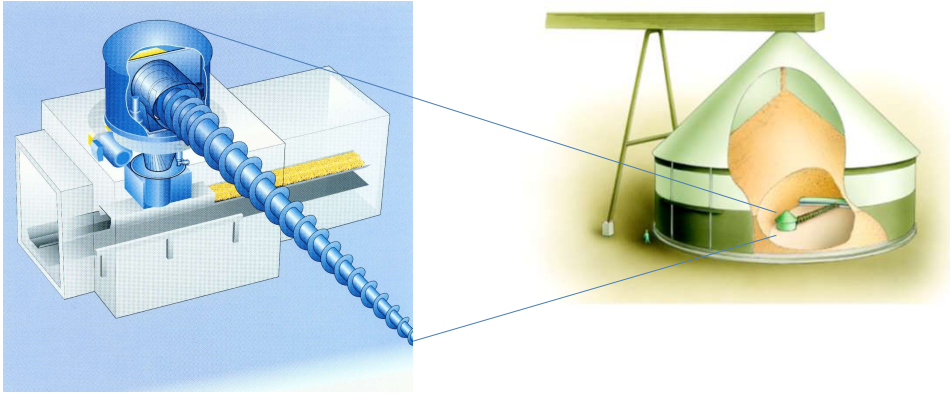
## Effect of Chip Storage

- At  $\sim 40^{\circ}\text{C}$ , autoxidation and hydrolysis of carbohydrates starts.
- **Wood substance losses during chip storage are at least 1 % per month**
- Losses are significantly higher with hardwood than for softwood chips. The same applies to sapwood.



(according to Wolfhaardt, Rabie 2003)

## Screw Reclaimer



Slewing screw reclaimer - CenterScrew™ (Andritz 1999)

## Wood Yard Losses

