

# Biological treatment processes of water and waste Lecture 6

# WAT - E2180

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#### **Lecture outline**

#### **Biological growth**

**Kinetics** 

Advanced process design

#### **BACTERIA GAME**

Dynamic nitrogen removal process

#### Storage processes

Storage polymers Applications

#### **Biological phosphorus removal**

Removal mechanism Existing process configurations **Anaerobic processes** Anaerobic digestion Fermentation

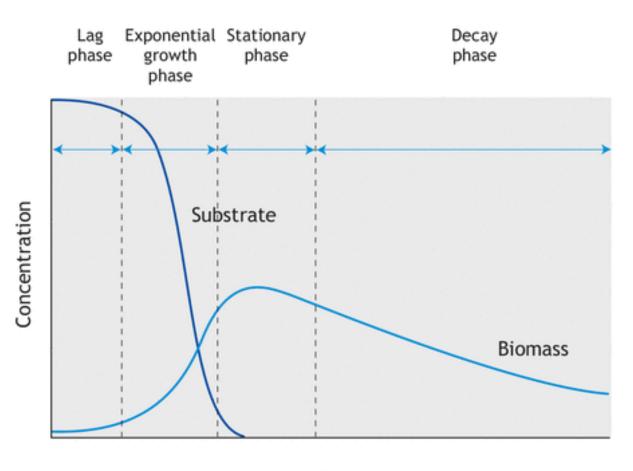


# Biological processes - growth



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# **Cell growth**



Time

Figure 2.16 Biomass growth in batch mode (adapted from Metcalf & Eddy, 2003)

# **Microbial growth**

Growth can be described with the equation:

$$r_{V,XB} = \mu_{max} f(S) X_B$$

Where

r<sub>V,XB</sub> = growth per unit of volume and time (e.g. kgCOD/m<sup>3</sup>d)

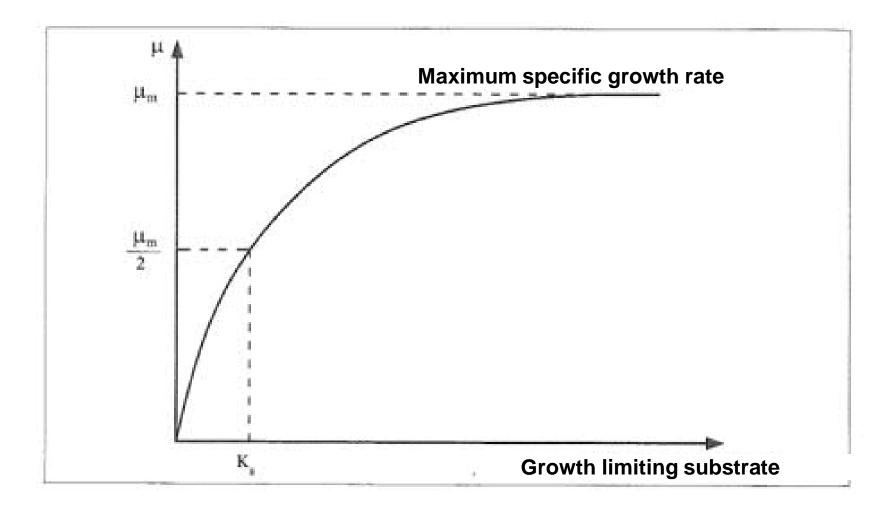
μ<sub>max</sub> = max specific growth rate (1/h or 1/d)

f(S) = growth kinetic function (depending on substrate), typically Monod

X<sub>B</sub> = biomass concentration (kgCOD/m<sup>3</sup> or kgVSS/m<sup>3</sup>)

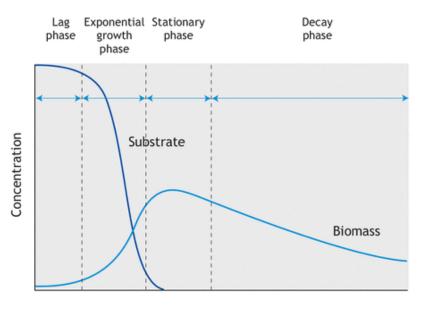


#### **Monod's kinetics**





## **Bacterial growth**



Time

Figure 2.16 Biomass growth in batch mode (adapted from Metcalf & Eddy, 2003)

$$r_{V,XB} = \mu_{\max} \cdot \frac{S}{S + K_S} \cdot X_B$$
$$r_{V,S} = \frac{\mu_{\max}}{Y_{\max}} \cdot \frac{S}{S + K_S} \cdot X_B$$

The growth of biomass depends on the substrate consumption (with a yield) and on the decay rate b

$$r_g = Y r_s - b X$$



## **Substrate consumption**

 $r_{V,S} = r_{V,B} / Y_{max}$ 

 $Y_{max} = maximum yield$ (kgCOD(B)/kgCOD(S) or kgVSS(B)/kgCOD(S) Yield shows how much of the consumed substrate is transformed into new biomass in the reaction. Note also  $Y_{obs}$  which is smaller than  $Y_{max}$ 



## **Monod kinetics**

# Monod kinetics are typically used for microbial growth

$$r_{V,XB} = \mu_{\max} \cdot \frac{S}{S + K_S} \cdot X_B$$
$$r_{V,S} = \frac{\mu_{\max}}{Y_{\max}} \cdot \frac{S}{S + K_S} \cdot X_B$$

For biomass growth (g/m<sup>3</sup>d)

For substrate consumption (g/m<sup>3</sup>d)

 $\mu_{obs} = \mu_{max} S / (S + K_s) [1/d]$ 

Observed specific growth rate



# Taking into account the growth conditions

Oxygen:

$$\mu_{obs} = \mu_{\max} \cdot \frac{S_{O2,2}}{S_{O2,2} + K_{S,O2}}$$

$$\mu_{obs} = \mu_{\max} \cdot \frac{S_2}{S_2 + K_s} \cdot \frac{S_{O2,2}}{S_{O2,2} + K_{S,O2}}$$

**Temperature:** 

$$\mu_{\max(\mathcal{T})} = \mu_{\max(20^\circ C)} \cdot e^{K(\mathcal{T}-20)}$$



# Typical values for stoichiometric and kinetic parameters

**Table 2.9** Typical values of stoichiometric ( $f_{s}^{\circ}$ , Y) and kinetic ( $q_{max}$ ,  $\mu_{max}$ ) parameters for various bacterial groups, (adapted from Rittmann and McCarty 2001)

| Electron donor                       |                              | Electron<br>acceptor  | ${f_S}^0$ | Y                             | $\mu_{max}$ | K                                |  |
|--------------------------------------|------------------------------|---|-----------|-------------------------------|-------------|----------------------------------|--|
| Microbial group e <sup>-</sup> donor |                              |   |           |                               |             |                                  |  |
| Chemotrophic organotroph             | ıs                           |   |           |                               |             |                                  |  |
| Aerobic heterotrophs                 | Sugar                        | O <sub>2</sub>  | 0.70      | 0.49 gVSS/gbCOD               | 13.2        | 27.0 g bCOD/gVSS.d               |  |
| Aerobic heterotrophs                 | No sugar                     | O <sub>2</sub>  | 0.60      | 0.42 gVSS/gbCOD               | 8.4         | 17.0 g bCOD/gVSS.d               |  |
| Denitrifiers                         | Organic                      | NO <sub>3</sub> <sup>-</sup> , NO <sub>2</sub> <sup>-</sup> | 0.50      | 0.25 gVSS/gbCOD               | 4.0         | 16.0 g bCOD/gVSS.d               |  |
| Fermenting organisms                 | Sugar                        | Organic   | 0.18      | 0.18 gVSS/gbCOD               | 1.2         | 10.0 g bCOD/gVSS.d               |  |
| Sulphate reducers                    | Acetate                      | SO4 <sup>2-</sup>   | 0.08      | 0.057 gVSS/gbCOD              | 0.5         | 8.7 g bCOD/gVSS.d                |  |
| Methanogens<br>(acetoclastic)        | Acetate                      | Acetate   | 0.05      | 0.035 gVSS/gbCOD              | 0.3         | 8.4 g bCOD/gVSS.d                |  |
| Chemotrophic lithotrophs             |                              |   |           |                               |             |                                  |  |
| Nitrifiers : AOB                     | NH4                          | O <sub>2</sub>  | 0.14      | 0.34 gVSS/gNH <sub>4</sub> -N | 0.9         | 2.7 g NH <sub>4</sub> -N /gVSS.d |  |
| Nitrifiers :NOB                      | NO <sub>2</sub> <sup>-</sup> | O <sub>2</sub>  | 0.10      | 0.08 gVSS/gNO2-N              | 0.5         | 1.1 g NO2-N/gVSS.d               |  |
| Methanogens<br>(hydrogenotrophic)    | H <sub>2</sub>               | $CO_2$  | 0.08      | $0.45 \text{ gVSS/gH}_2$      | 0.3         | 1.1 g H <sub>2</sub> /gVSS.d     |  |

 $k = \mu_{max}/Y = \text{specific } r_{max}$  (per unit biomass)

#### **Denitrification rate**

$$r_{\rm V,NO} = \left(\frac{1 - Y_{\rm H}}{2.86Y_{\rm H}}\right) \mu_{\rm max,H} \left(\frac{S_{\rm BOD}}{K_{\rm COD} + S_{\rm COD}}\right) \left(\frac{S_{\rm NO}}{K_{\rm NO} + S_{\rm NO}}\right) \eta_{\rm g} X_{\rm b,h}$$

where

 $r_{\rm V, NO}$  = reaction rate per unit volume nitrate- and nitrite-nitrogen,

 $Y_{\rm H}$  = biomass yield coefficient,

 $\mu_{max, H}$  = maximum specific growth rate of heterotrophs,

 $S_{COD}$  = soluble material concentration organic substrate,

 $K_{COD}$  = half-saturation coefficient organic substrate,

 $S_{\rm NO}$  = soluble material concentration nitrate- and nitrite-nitrogen,

$$K_{\rm NO}$$
 = half-saturation coefficient nitrate-nitrite,

 $\eta_g$  = correction factor for  $\mu_H$  under anoxic conditions, and

 $X_{b,h}$  = particulate material concentrations.



# Design of biological processes



# **DEMO 1**

| Average daily flow rate  | 37 850 m3/d          |  |
|--------------------------|----------------------|--|
| Influent water:          |                      |  |
| BOD7                     | 140 mg/l             |  |
| Ammonium-N               | 35 mg/l              |  |
| Suspended solids         | 90 mg/l              |  |
| Of which unbiodegradable | 30 mg/l              |  |
| Effluent water:          |                      |  |
| BOD7                     | 10 mg/l              |  |
| Ammonium-N               | 0,5 mg/l             |  |
| Total N                  | 10 mg/l              |  |
| Suspended solids         | 15 mg/l              |  |
| Temperature              | 12 C                 |  |
| MLVSS/MLSS               | 0,8                  |  |
| MLVSS                    | 2,4 g/l              |  |
| Y (heterotrophs)         | 0,6 kg VSS/kg BOD    |  |
| b (12 C)                 | 0,044 d-1            |  |
| Y (nitrifiers)           | 0,12 kg VSS/kg NH4-N |  |
| bN (12 C)                | 0,06 d-1             |  |

Dimension an activated sludge process where full nitrification and 70% denitrification is achieved.

Use the denitrification rate (12°C, raw WW) of 1,5 gN/kgVSSh

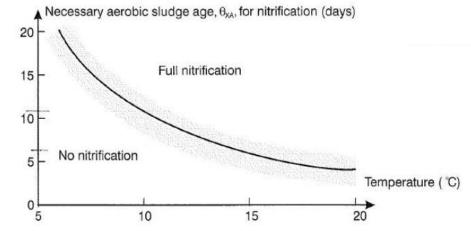
Assumption 1: No nitrate in the influent water

Assumption 2: No denitrification in the secondary clarifiers.



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# DEMO 1



- Dimensioning of nitrifying process:
- Choose SRT→ nitrification
- 12 ° C  $\rightarrow$  10 d
- Calculate the needed biomass per day:

- Biomass XV = 
$$\frac{YQ(So-Se)}{1+b\theta c}$$
 +  $\frac{Y_nQ(S_{NH4}-Se_{NH4})}{1+b_n\theta c}$  =

2031,2 + 97,0 = 2128 kgVSS/d

- Biomass (VSS  $\rightarrow$  SS) = 2660 kgSS/d
- Inert particulate influent 1136 kgSS/d → Total sludge amount SS = 3796 kgSS/d, sludge concentration 3 g/l
- Reactor volume needed= (sludge amount x SRT) / X = 12653 m3

# NOTE! This calculation is simplified → inert SS produced not considered



#### **DEMO 1**

The volume needed for denitrification is calculated based on the denitrification rate.

Total N 70 %  $\rightarrow$  to be denitrified 927 kg/d = 38,6 kgN/h = 38 600 gN/h

Denitrification rate (12C, raw WW) $\rightarrow$  1,5 gN/kgVSS/h MLVSS 2,4 g/l  $\rightarrow$  needed volume 10 722 m<sup>3</sup>



#### **Removal in biological processes**

| Removal   | Conditions   | When   |  |
|---|--|--|--|
| Organic<br>matter   | Aerobic, short SRT                                       | Focus on removal                                   |  |
| Organic<br>matter   | Anaerobic, long HRT&SRT                                  | for high strenght waters, focus on energy recovery |  |
| Ammonium<br>(nitrification)                                 | Aerobic, long SRT  | Well known process                                 |  |
| <b>Total nitrogen</b><br>Nitrification +<br>denitrification | Aerobic + anoxic zones,<br>Short/long SRT                | Well known process                                 |  |
| Total nitrogen<br>Short-cuts                                | Low DO (nitritation + denitritation)<br>Deammonification | Focus on energy savings<br>High strength waters    |  |
| Phosphorus  | Aerobic + anaerobic zones                                | P reuse, sludge production, carbon source          |  |
| NOM, iron,<br>manganese,<br>organic<br>micropollutants      | Aerobic, mainly long SRT                                 | Often cost effective solution                      |  |

# **Design approaches for biological processes**

Sludge age (days)  $\rightarrow$  Sludge age selected on the basis of process goals (e.g. nitrification)  $\rightarrow$  Basin volume

Sludge loading (kgBOD/kgMLSSd)  $\rightarrow$  As above but does not take into account differences in sludge yield (e.g. process design when water is very typical)

Volumetric loading (kgBOD/m<sup>3</sup>d) e.g. rough estimations for bugdeting

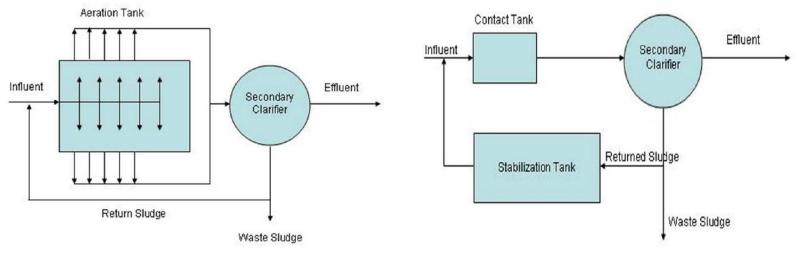
Surface loading (kgBOD/m<sup>2</sup>d or kgN/m<sup>2</sup>d) MBBR and biological filters

Reaction rate (gN/gMLSSh) e.g. anoxic zones

Hydraulic retention time (hours) e.g. anaerobic and anoxic zones



## Important design aspects

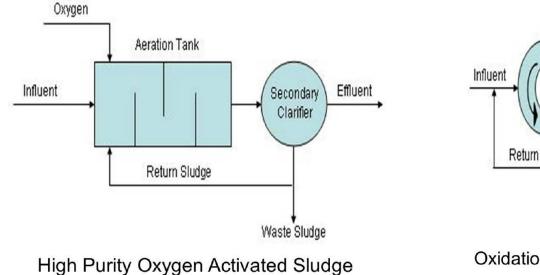


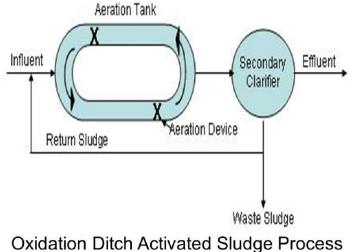
Complete Mix Activated Sludge Process

Contact Stabilization Activated Sludge



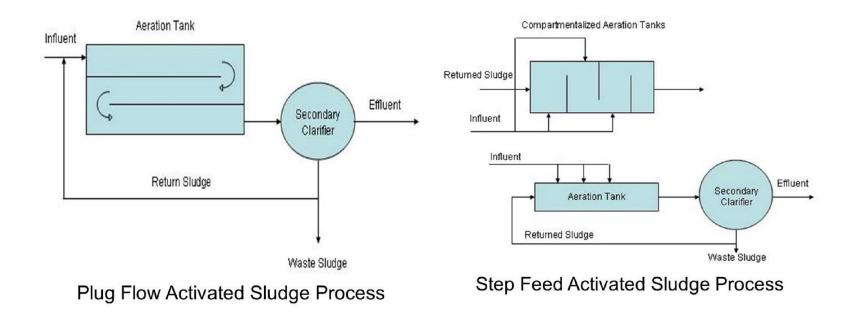
## Intensive or extended aeration





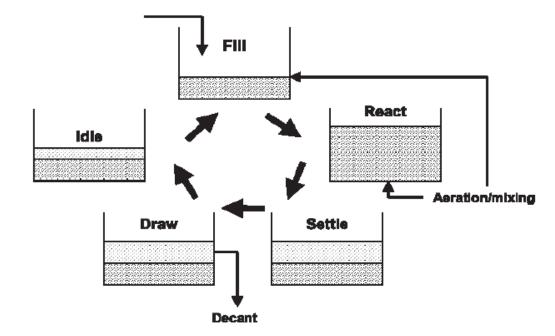


## **Process configurations**





# **Process configuration**



Sequencing batch reactor All reactions happen in the same volume Typically 2 or 3 parallel SBRs



#### **Process type**

Activated sludge Moving bed bioreactor (MBBR) Membrane bioreactor (MBR) Biological filters Aerobic granular sludge (AGS)

Selection depends on wastewater characteristics, climate conditions, energy aspects, effluent requirements, size etc.



# Nitrogen and phosphorus removal

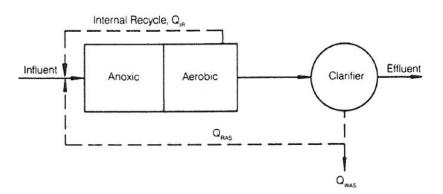


Figure 8.9 Modified Ludzack–Ettinger process for nitrogen removal (WAS = waste activated sludge).

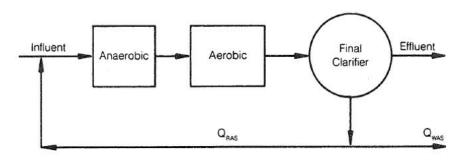


Figure 8.3 The A/O process (RAS = return activated sludge, WAS = waste activated sludge).



# **Alternative configurations**

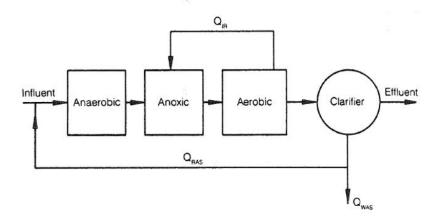


Figure 8.14 A<sup>2</sup>/O process for phosphorus removal (WAS = waste activated sludge).

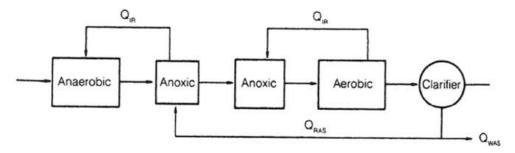


Figure 8.15 Modified University of Cape Town process for phosphorus and nitrogen removal (WAS = waste activated sludge).



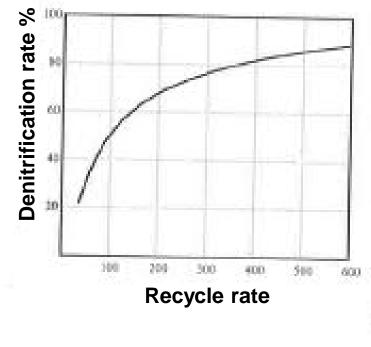
# How to calculate the recycle ratio of RAS?

Recycle ratio = 100X / ((1200/SVI) - X ) Where X = MLSS (g/l)

And SVI = sludge volume index (ml/g)

Recycle must be sufficient for denitrification

Typical recycle ratio 100 - 200 %







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# BACTERIA GAME: Advanced version 2 Dynamic nitrogen removal process ③

- First test the process with ND configuration
- Each player picks a type of microbe and places it in suitable conditions in the process

#### ZONE 1:

- Each player picks two wastewater constituent. This is the influent wastewater that enters the first zone.
- Try to form suitable sets for your microbe's reaction using influent wastewater and zone conditions.
- When a reaction occurs, select the correct end-products
- Check if possible reactions exist with



#### **ZONE 2**:

- Move the wastewater constituents and the endproducts that have not yet reacted to zone 2.
- Repeat the steps from zone 1.

#### RAS flow

- Move the wastewater constituents and the endproducts that have not yet reacted to zone 1 via RAS flow.
- Repeat the steps from above.
- GAME ENDS WHEN ALL THE REACTIONS HAVE OCCURRED. Laitoksen nimi 03/24/2021

# BACTERIA GAME: Advanced version 2 Dynamic nitrogen removal process ©

- Now test the process with DN configuration
- Each player picks a type of microbe and places it in suitable conditions in the process

#### ZONE 1:

- Each player picks two wastewater constituent.
- Try to form suitable sets for your microbe's reaction using influent wastewater and zone conditions.
- When a reaction occurs, select the correct end-products
- Check if possible reactions exist with the end-products
  - Aalto Un School

- ZONE 2:
  - Move the wastewater constituents and the end-products that have not yet reacted to zone 2.
  - Repeat the steps from zone 1.

#### - RAS flow

- Move the wastewater constituents and the end-products that have not yet reacted to zone 1 via RAS flow.
- Repeat the steps from above.

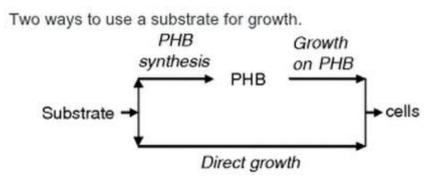
#### - GAME ENDS WHEN ALL THE REACTIONS HAVE OCCURRED.

# Storage processes



# Role of storage processes in growth

- Substrate can be converted and stored within bacterial cells as energy storage.
- Bacterial growth can be based on direct growth on the substrate or on growth on these storage polymers.
- Growth on storage has a bit lower yield (energetically less efficient)
  4 - 10% less sludge production.
- Common storage polymers Polyhydroxyalkaonate PHA and polyhydroxybutyrate PHB
- Storage polymers are a benefit in bacterial competition.



# PHA & PHB

#### PHA

- Up to 90% cell dry weight
- Similar characteristics to plastics
- Biodegradable
- Example: Mars, Attero Venlo (PHA from biowaste)

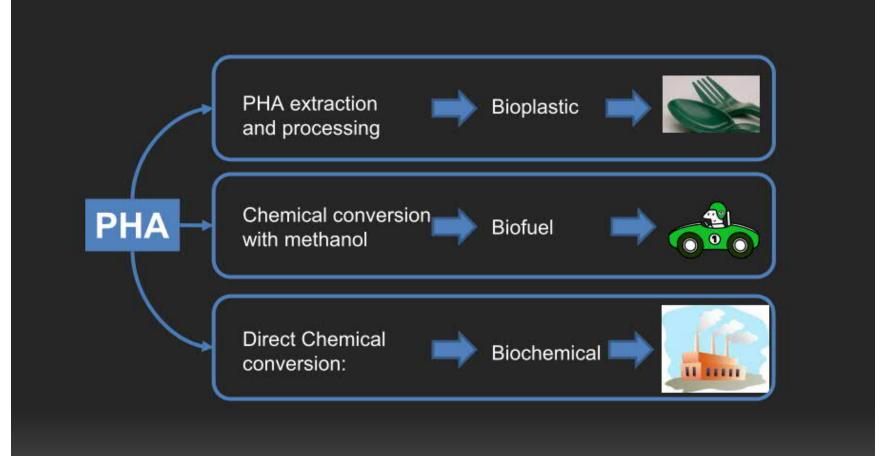
#### **PHA&PHB** production

Important things to consider

- Production yield
- Volumetric productivity
- PHA&PHB concentration
- PHA&PHB composition

#### PHB: example Mirel (USA) Caproates: animal feed,





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| Company  | Products   |  |
|--|--|--|
| Berlin Packaging Corp. (U.S.)                              | Zeneca/ICI Biopol  |  |
| Bioscience Ltd. (Finland)                                  | Medical applications of PHAs   |  |
| Bioventures Alberta, Inc. (Canada)                         | PHA produced by recombinant Escherichia coli   |  |
| Metabolix, Inc. (U.S.)                                     | PHB, P(HB : HV) (Mirel)  |  |
| Metabolix/ADM  | Transgenic plant PHAs  |  |
| Monsanto (U.S.)  | Transgenic plant PHAs  |  |
| Polyferm, Inc. (Canada)                                    | PHAs from hemicellulose; use of Burkholderia cepacia on<br>xylose                            |  |
| Monsanto-Metabolix (U.S.)                                  | Biopol from Cupriavidus necator  |  |
| Nodax Procter and Gamble (U.S.)                            | PHBHx, PHBO, PHBOd (Nodax)   |  |
| Tianan Biologic Material Co (China)                        | PHB and P(HB : HV) (Enmat)   |  |
| Tianjin GreenBio Materials Co., Ltd.<br>(GreenBio) (China) | Sogreen  |  |
| Biocycle Copersucar (Brasil)                               | PHB and P(HB : HV) (Biocycle)  |  |
| Biomer (Germany)   | PHB and P(HB : HV) (Biomer L)  |  |
| BIO-ON (Italy)   | Minerv-PHA (from sugar beets)  |  |
| NatureWorks LLC (U.S.)                                     | Ingeo biopolymer   |  |
| Micromidas   | Constructed microbial population able to adapt to a variety of<br>materials, including waste |  |

#### Table 15.1 Some Companies Involved in PHA Production.

#### How Mirel is Made

#### **Biodegradable\***

Mirel is biodegradable in natural soil and water environments, home and industrial composting facilities, where available.

Applications

Mirel can be processed on conventional equipment and used in everyday products. Biobased Starting with corn.

#### Corn Sugar

One of many products made from each kernel of corn, used as feedstock for Mirel.

#### Fermentation A patented process,

transforms the sugar into Mirel biopolymers.

#### Formulation

Mirel is compounded into resin pellets.

23 Metabolix

Proprietary

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# Biological phosphorus removal



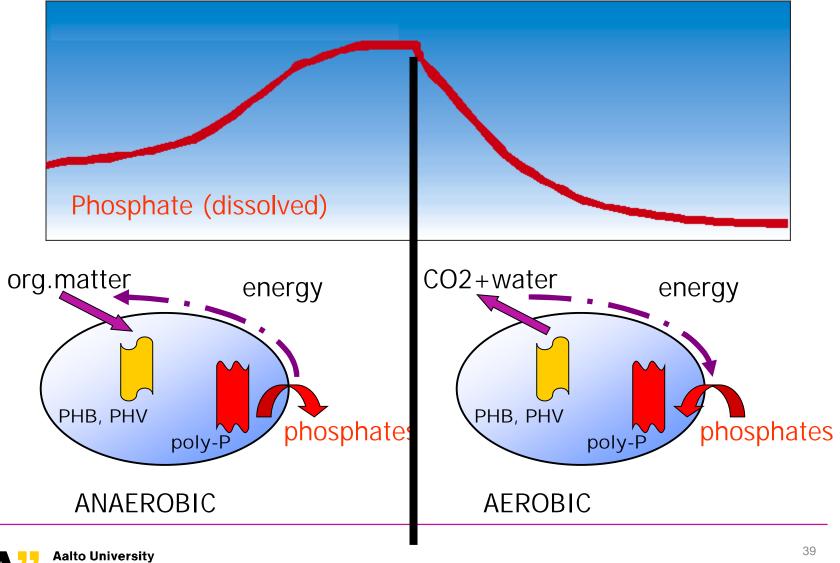
# **Biological phosphorus removal**

- Phenomenon was discovered by accident in India 1959
- Observed in full-scale plant in South Africa in the 70s also by accident
- Based on microbes capable of storing polyphosphates
- Require alternating anaerobic (not even nitrates) and aerobic conditions and carbon source in the anaerobic phase.
- PAOs phosphorus accumulating organisms
- Competition with GAOs (Glycogen accumulating organisms) especially in warm temperatures



# BioP

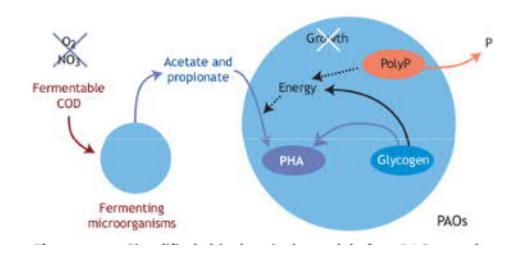
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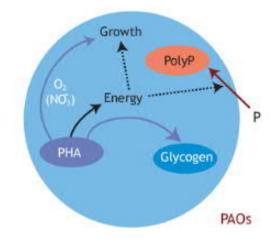




#### **ANAEROBIC CONDITIONS**

### **AEROBIC CONDITIONS**

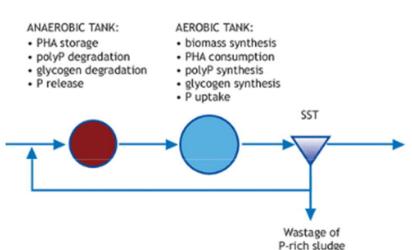




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# **Principles of bioP**

- Phosphorus accumulating organisms (PAOs) store organic matter as polyhydroxyalcanoates (PHA) in anaerobic conditions using energy from poly-P inside the cell
- In aerobic conditions PAOs store more poly-P than needed for the normal metabolism using stored PHA
- Phosphorus is removed with the sludge (3-8 % of P)





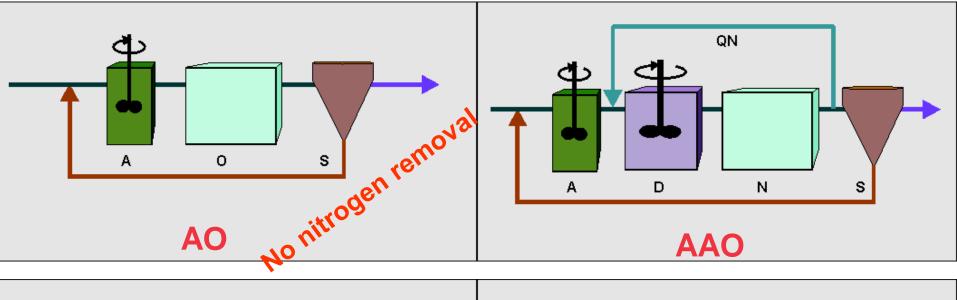
# Important aspects in bioP processes

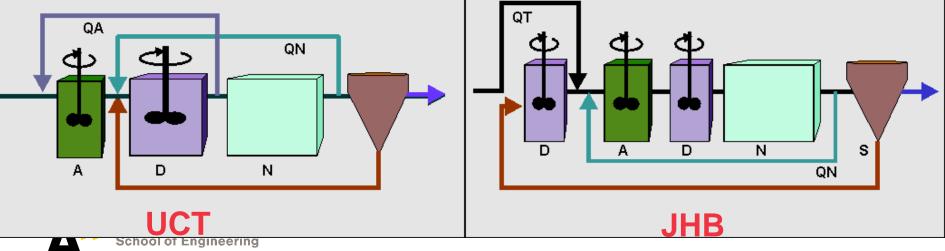
- Minimizing oxygen in the anaerobic zone
- Minimizing nitrates and nitrites in the anaerobic zone.
- Increase volatile fatty acids (VFA) concentration in the anaerobic zone. (VFA is taken up and forms PHA)

- Minimizing solids in the effluent (high P content)
- Maximizing phosphorus uptake = short SRT and good oxygen concentration pattern



### **Biological P removal processes**

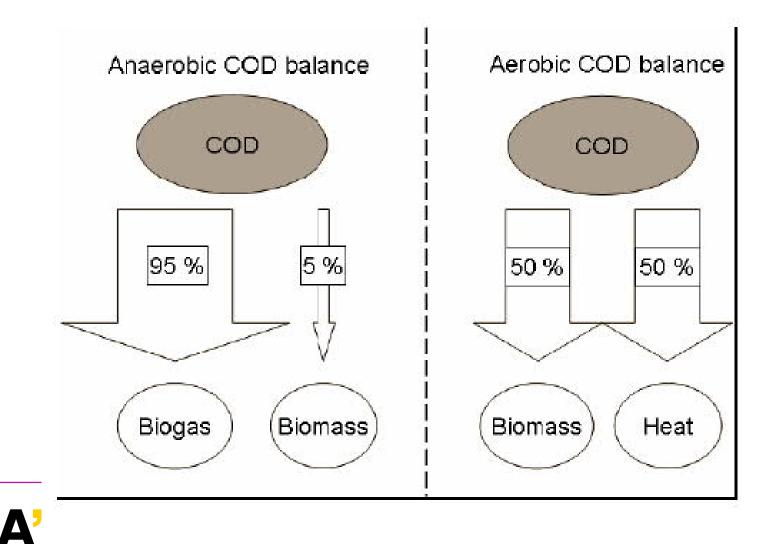




# Anaerobic processes



### Why anaerobic treatment?



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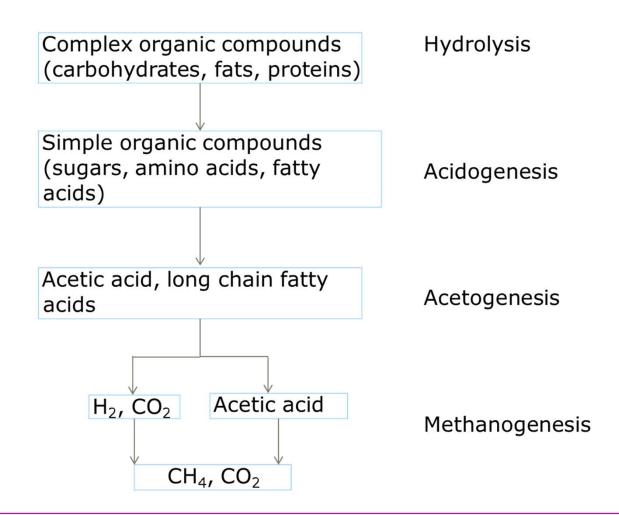
# Pros and cons of anaerobic digestion

- + CO<sub>2</sub> as electron acceptor
- + no need for aeration
- + Low sludge yield
- + Produces methane, 90% can be used as energy (9000 kcal/m<sup>3</sup>)
- + high loading→ less space
- + Works with certain organic compounds that can not be degraded in aerobic conditions

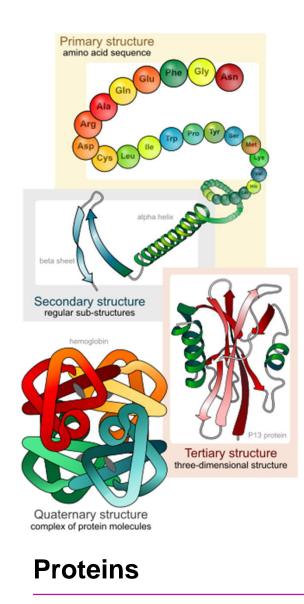
- Slow process (HRT about 30 d)
- Sensitive to toxic substances
- Long start-up
- Requires high substrate concentrations



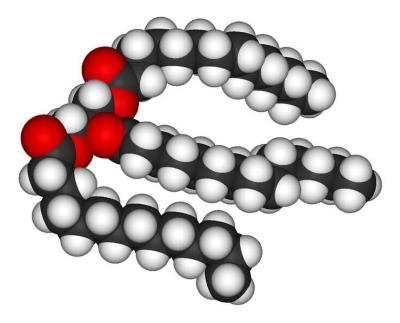
### **Anaerobic digestion**



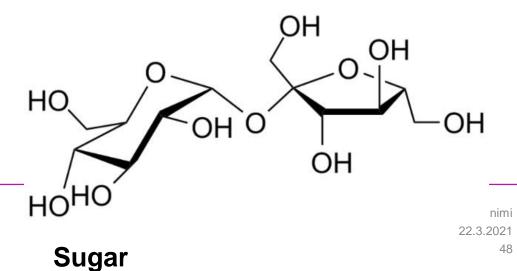








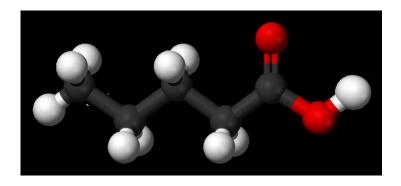
Fats (triglyseride molecule)



# **Hydrolysis**

- First step of the anaerobic digestion
- Different groups of bacteria produce extracellular enzymes to cut the larger organic molecules into smaller ones
- Larger molecules = proteins, fats, carbohydrates
- Smaller molecules = small molecule sugars, amino acids, short chain fatty acids

# Hydrolysis products, example valeric acid

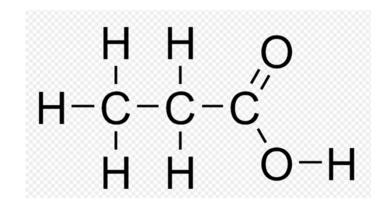




# Acidogenesis

- Second step of the anaerobic digestion
- Acidogenesis
- Bacteria degrades the organic molecules further to short-chain fatty acids and alcohols
- Ammonium, hydrogen and CO<sub>2</sub> also produced

End product in this step, for example propanoic acid





# **Acetogenesis and methanogenesis**

- Third step of the anaerobic digestion
- Acetogenic bacteria degrades the short chain fatty acids to acetic acid (and hydrogen and CO<sub>2</sub>)

Last step of the anaerobic digestion Methanogenic bacteria use

acetic acid, CO<sub>2</sub> and hydrogen to produce biogas (=methane)

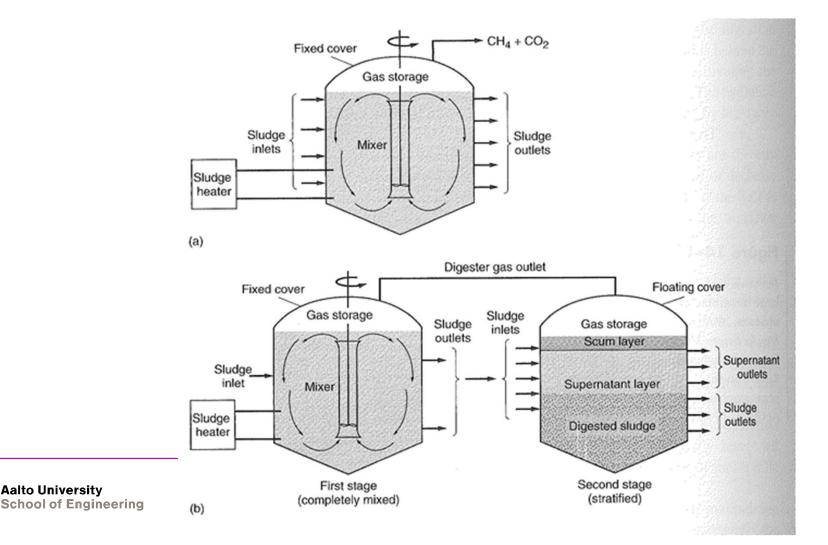


# **Pre-fermentation**

- In order to produce VFA = volatile fatty acids
- VFAs are enhancing denitrification and biological phosphorus removal
- Can be done with influent waste water, raw sludge, waste activated sludge or a industrial influent



# Anaerobic processes for sludge digestion (biogas plants)



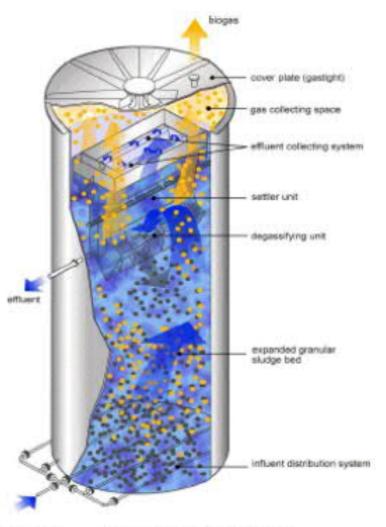
# **Digestion processes**

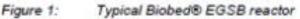
Mesophilic 33 – 37 °C Retention time about 21 days

Thermophilic 54 – 55 °C Retention time about 14 days Requires more energy



# Anaerobic processes in wastewater treatment



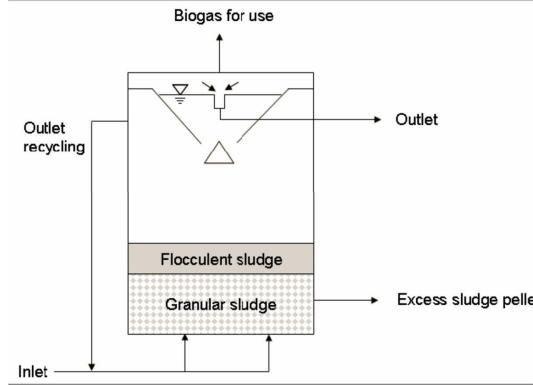




Typical Biobed<sup>®</sup> EGSB plant at Lapin Kulta, Haparanda (Finland)

# UASB reactor (Upflow anaerobic sludge blanket)

- Granular biomass is created in the reactor
- Biomass is kept in suspension by the gravity of the granules and the upflow of wastewater





# **Reading material**

**Biological wastewater treatment (Course book):** 

Chapters

2.1

- 2.2.7 2.2.8
- 2.3

2.4

7.1 – 7.4

