

# **Evolving Bacterial Fitness with an Expanded Genetic Code**

*Tack, Cole, Shroff, Morrow & Ellington*  
**Scientific Reports**  
**2018**

Matti Ahokas, Eskild Fisker Angen, Inka Mattila, Linda van den Berg  
**Synthetic Biology Lecture Course**  
17.05.2021

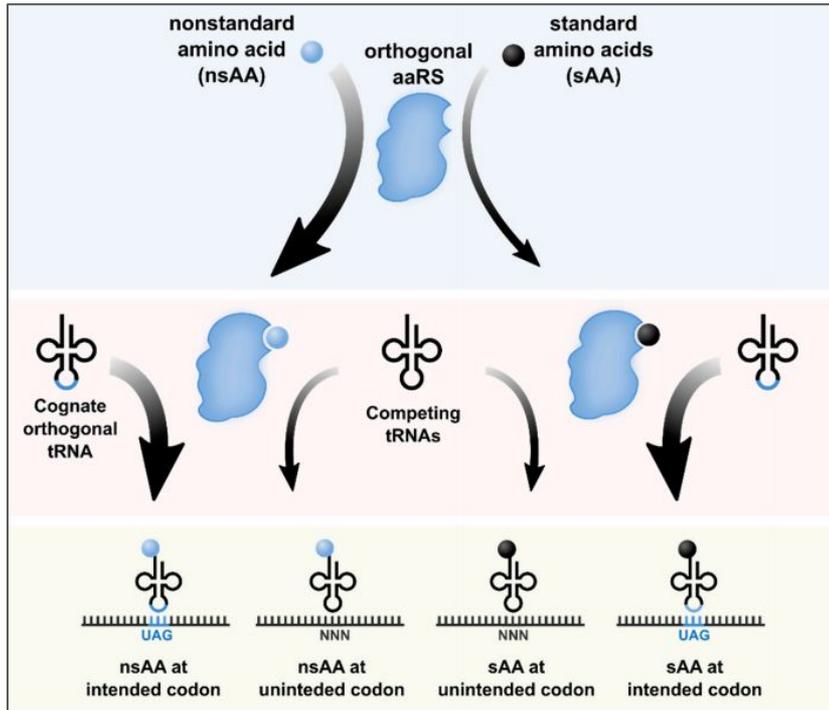
# Brief introduction to the topic

- There are 20 canonical amino acids in the standard genetic code.
- The development of the orthogonal translation systems (OTSs) allow incorporating noncanonical amino acids (ncAAs) to expand the genetic code.
- OTSs allow incorporating ncAAs by suppressing the amber stop codon (UAG).
- Cells containing an active OTS often exhibit fitness deficits.
- Some strains have found to be viable only when essential genes terminating with an amber stop codon were recoded to terminate with an alternative stop codon.

# Aim of the study

- **The aim of this study was to perform directed evolution experiments with an orthogonal translation system that inserts 3-nitro-L-tyrosine (3nY) across from amber codons (UAG), to create a 21 amino acid genetic code in which the amber stop codon encodes either 3-nitro-L-tyrosine or stop.**
- The 21 amino acid code is enforced through the inclusion of an addicted, essential gene.
- After 2000 generations of directed evolution, the fitness of the strain was largely repaired.

# OTS - Changing the translation machinery



From: Arranz-Gibert et al., 2019

**Problem:** Expand genetic code without causing cross-reactions with endogenous transcription, or translation machinery

**Solution:** Use enzymes from phylogenetically distant organism to favor ncAA incorporation

- **Engineered aaRS:tRNA pairs**
  - aaRS charges ncAA onto its cognate tRNA
  - tRNA promotes ncAA incorporation into protein

abbreviations:

aaRS = Tyrosyl-aminoacyl tRNA synthetase

ncAA/nsAA = Non-canonical/standard amino acid

sAA = Standard amino acid

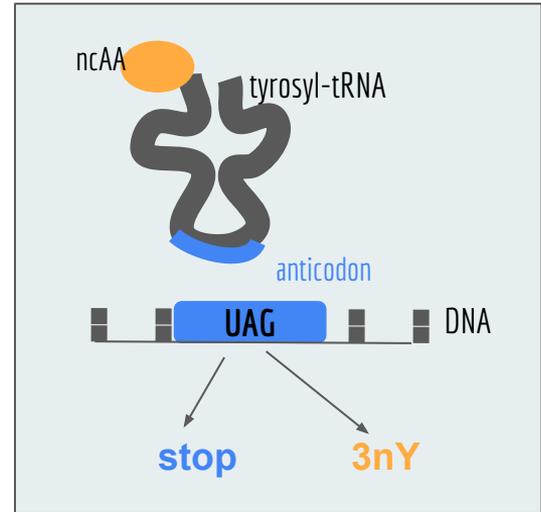
# Site-specific incorporation of 3nY

Heterologous components from *Methanocaldococcus jannaschii*:

- Tyrosyl-aaRS
- Suppressor tyrosyl-tRNA
  - Tyrosyl-tRNA anticodon complementary to UAG
  - Engineered to be specific for 3-iodo-L-tyrosine, compatible also with 3nY (*Sakamoto et al., 2009*)

## Aim:

- Cause ambiguity in codon translation
- Stop vs. incorporation of 3nY
  - Long term: Enforce switch to 3nY altogether



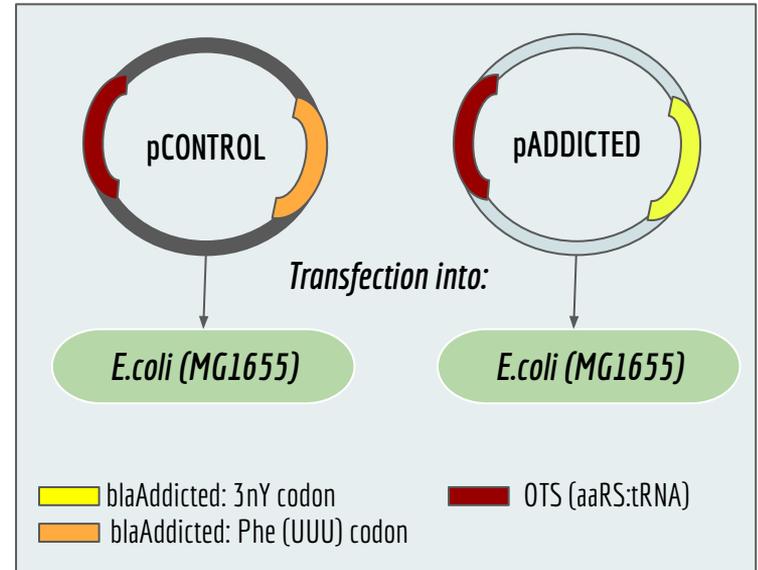
## Abbreviations:

3nY = 3-nitro-L-tyrosine

# OTS-addiction via a $\beta$ -lactamase variant

## Engineered beta-lactamase gene (*bla*):

- “*blaAddicted*” variant:
  - Confers moderate resistance to CAZ
  - Dependency on 3nY incorporation at AA position 162
- Control with Phenylalanine (UUU) codon:
  - Produces functional *bla*
  - No dependency on 3nY



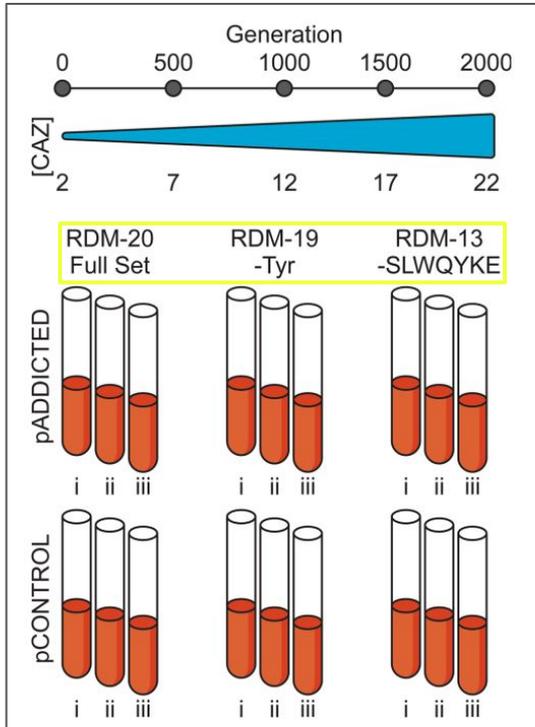
### Abbreviations:

AA= Amino acid

CAZ = ceftazidime

Phe = Phenylalanine

# Directed evolution



From: Tack et al., 2018

**Transfected *E.coli* to different AA media for 2000 generations:**

→ *SLWQYKE*: AAs codons accessible through SNPs in UAG

→ All media supplemented with 10mM 3nY

**Evolutionary pressure for 3nY retainment via:**

- Progressive increase in CAZ concentration  $1 \mu\text{g mL}^{-1}$  per 100 generations to enforce 3nY incorporation

**Pheno-/Genotyping via:**

- Growth rates = Cellular fitness before and after evolution
- WGS = Genomic adaptation after evolution
- GFP reporter assay = OTS retention/UAG suppression efficiency before and after evolution

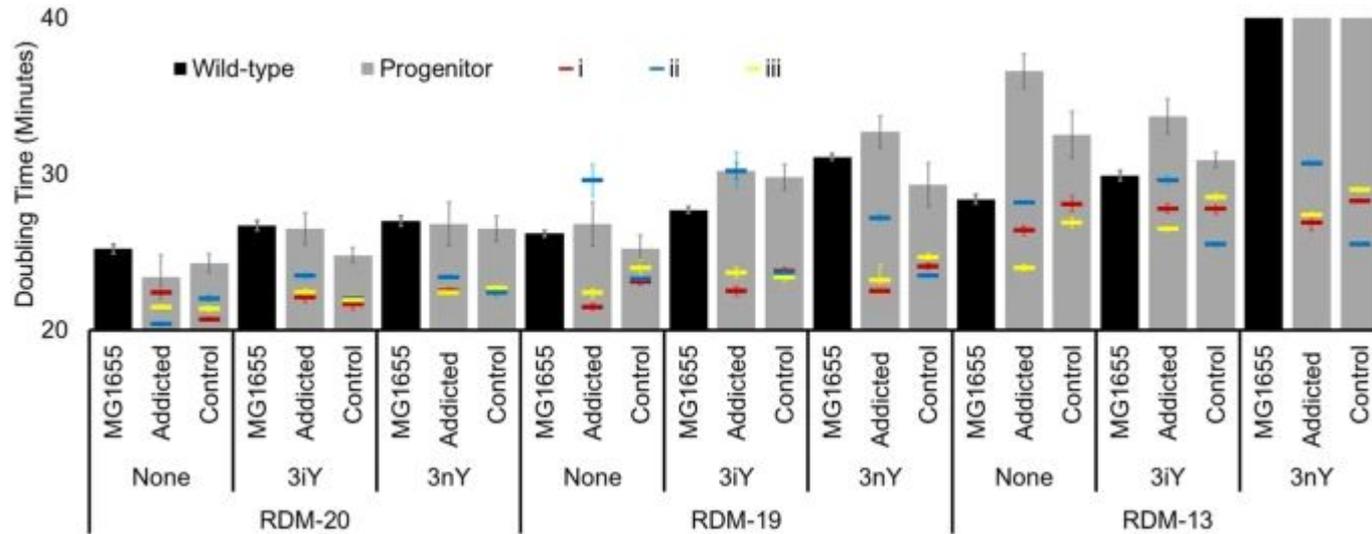
Abbreviations

SNP = Single nucleotide polymorphism

WGS = Whole genome sequencing

# Growth rates as a measure of fitness

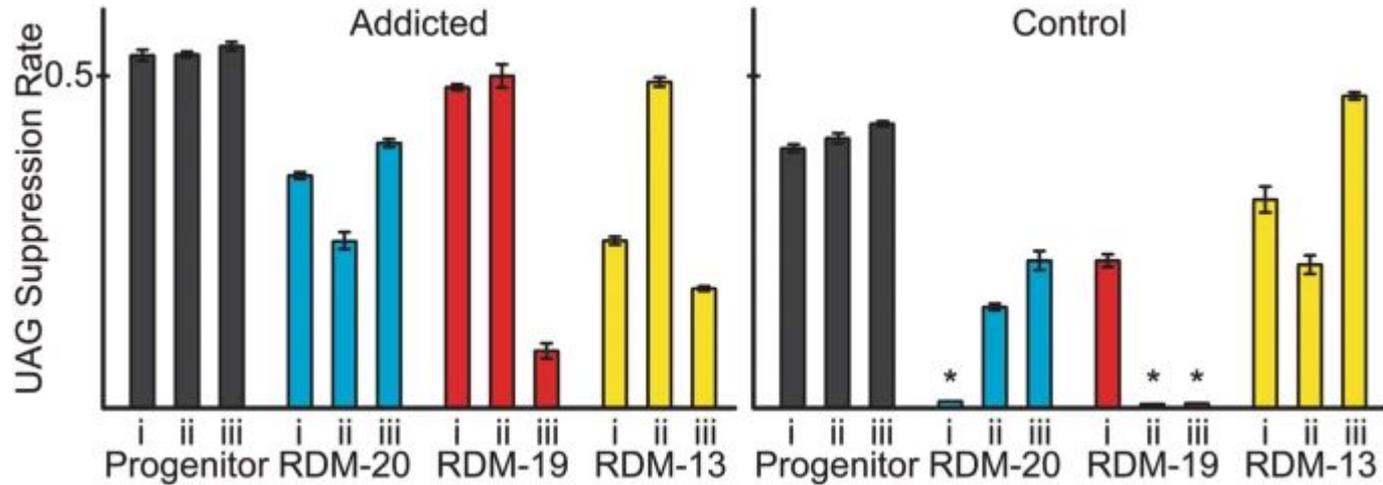
Effect of changing the media aa composition





# OTS-addiction and the use of non-canonical amino acids

UAG suppression measured with GFP



Addicted lines showed conservation of OTS

# Results

- Three different mixtures of amino acids environments were used (RDM-20, RDM-19, RDM-13)
- Three parameters were tested in each environment: without ncAA, with 3iY and with 3nY.
- Fitness was determined by calculating doubling times for *E. Coli* (MG1655) in these different mixtures.
- significant increases in doubling times was found in all three conditions(RDMs) when compared to media without ncAAs.
- the addition of 3iY to RDM-13 did not have a significant effect, with doubling times similar to growth in RDM-13 without ncAA.
- These results suggest that 3nY is mildly toxic and 3iY is less so.
- doubling times were not significantly affected by the addition of the OTS in the absence of ncAAs.

# Conclusion

Created a strain with the ability to utilize a non-canonical aa

Fitness deficit repaired during only 2000 generations

Better understanding of the evolution of adoption of aa

A step toward designing unique organisms

# References

**Arranz-Gibert et al.:** The Role of Orthogonality in Genetic Code Expansion. *Life (Basel)*. 2019; 9(3): 58.

**Tack et al.:** Evolving Bacterial Fitness with an Expanded Genetic Code. *Sci Rep*. 2018; 8(1):3288.

**Sakamoto et al.:** Genetic Encoding of 3-Iodo-L-Tyrosine in *Escherichia coli* for Single-Wavelength Anomalous Dispersion Phasing in Protein Crystallography. *Structure*. 2009; 17, 335–344.