

The background of the slide features two large, overlapping, semi-transparent images of protein structures. These structures are rendered in a multi-colored ribbon style, with colors including blue, green, yellow, orange, and red. They are positioned in the top-left and bottom-right corners, framing the central text.

Biomolecules

ELEC-E3260

27th January

Mass Spectrometry techniques

Summary

- Brief History
- What is it?
- How is it working?
- Mass Spectrometry and Biomolecules
- Applications, advantages and disadvantages
- In-class activity

Brief History

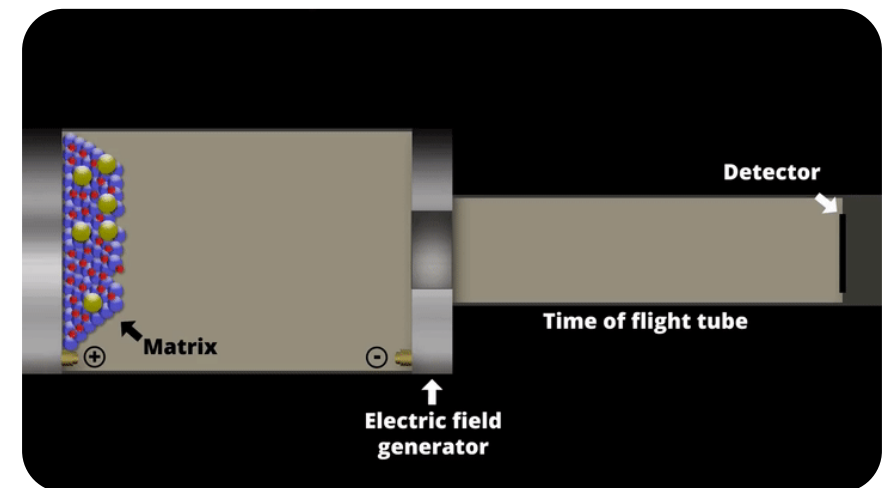
- Sir J.J. Thomson: The 'Father' of Mass Spectrometry. 1897-1919 Developed the Mass Spectrograph (with F.W. Aston).
- Nobel Prize of Physics for Thomson in 1906.
- The World's first commercial instrument became available in 1948.
- Time-of-flight (TOF) analysis (Wiley and Maclaren) and quadrupole analysis (Paul) were conceived in the early 1950's.
- In 1956, the first biologically important molecules were successfully analyzed.



The MS-2 - marketed by Vickers



Sir J.J. Thomson



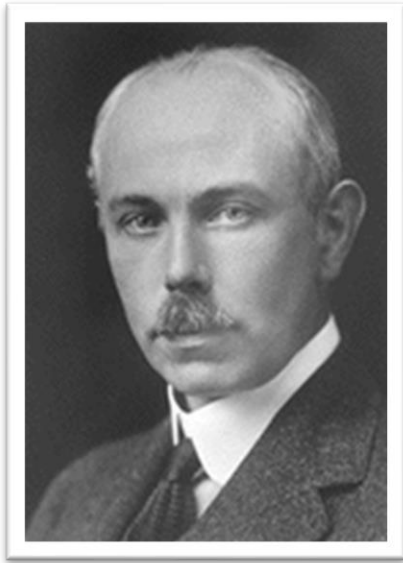
Some Nobel Prize Pionners For MS



J.J. Thomson

1906 Nobel Prize of Physics

For his development of Mass spectrograph



Francis William Aston

1922 Nobel Prize for Chemistry

For his discovery, by means of his mass spectrograph, of isotopes, in a large number of non-radioactive elements, and for his enunciation of the whole-number rule



Wolfgang Paul

1989 Nobel Prize for Physics

For the development of the ion trap technique



John Bennet Fenn

2002 Nobel Prize for Chemistry

For the development of soft desorption ionisation methods (ESI) for mass spectrometric analyses of biological macromolecules



Koichi Tanaka

2002 Nobel Prize for Chemistry

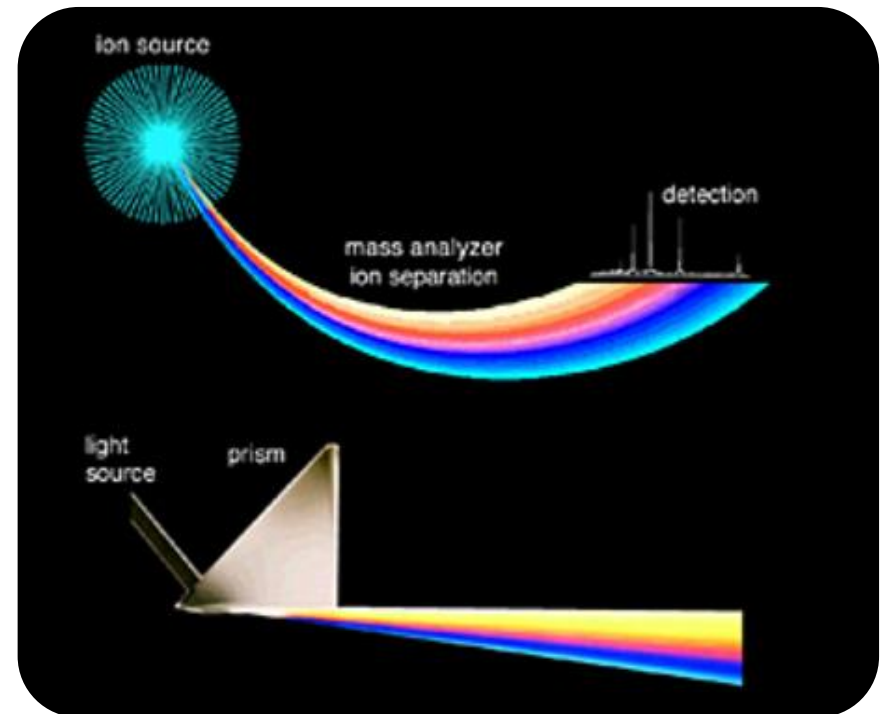
for the development of soft desorption ionisation methods (MALDI) for mass spectrometric analyses of biological macromolecules

What is it?

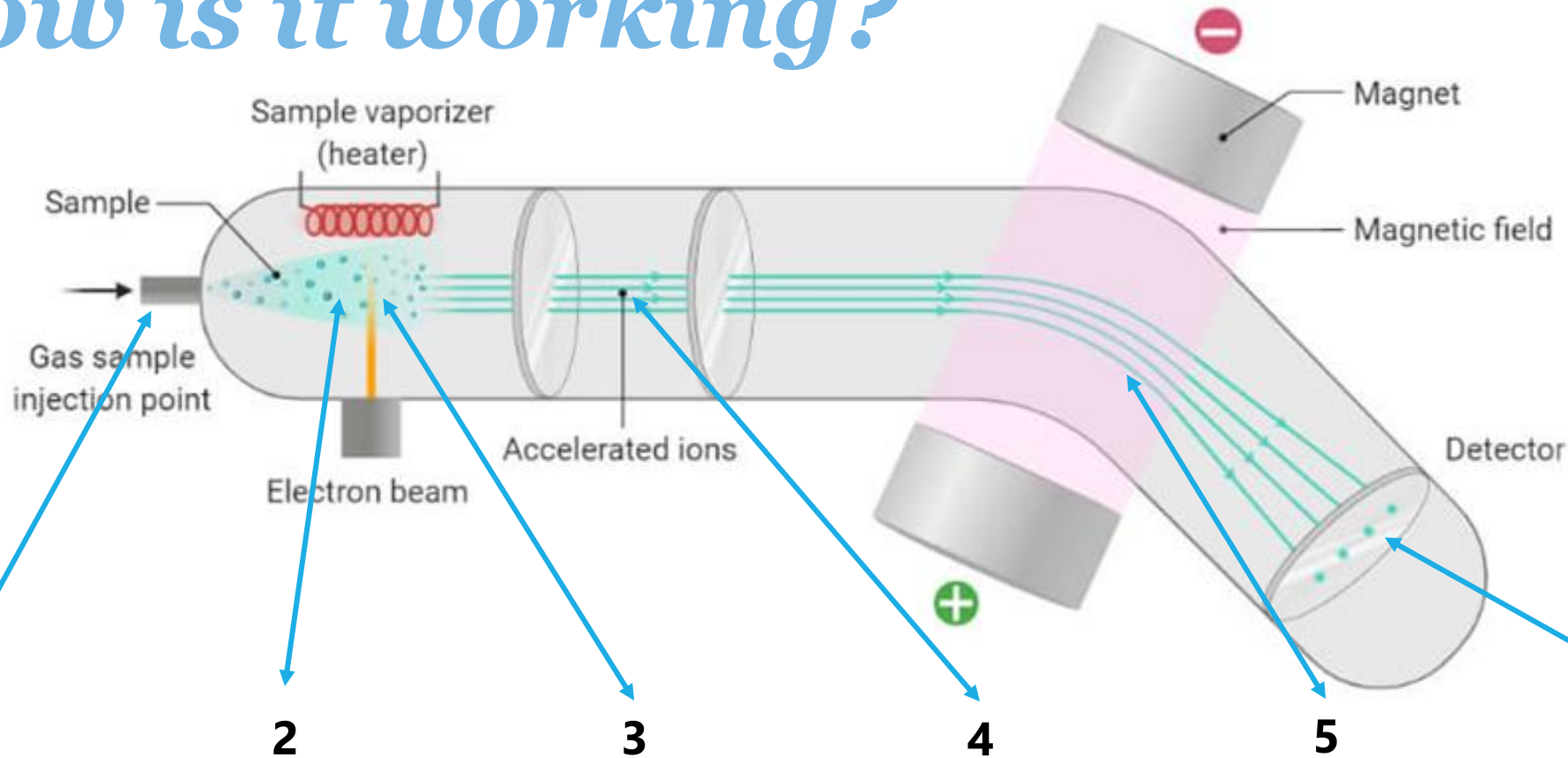
"Mass spectrometry is the art of measuring atoms and molecules to determine their molecular weight. Such mass or weight information is sometimes sufficient, frequently necessary, and always useful in determining the identity of a species. To practice this art, one puts charge on the molecules of interest, i.e., the analyte, then measures how the trajectories of the resulting ions respond in vacuum to various combinations of electric and magnetic fields."

- MS can:
 - Quantify known materials
 - Identify unknown compounds
 - Elucidate the structure and chemical properties of different molecules
 - Distinguished different isotopes of the same element

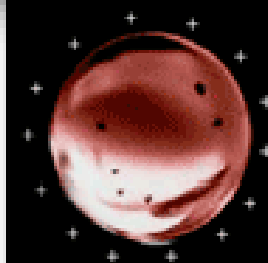
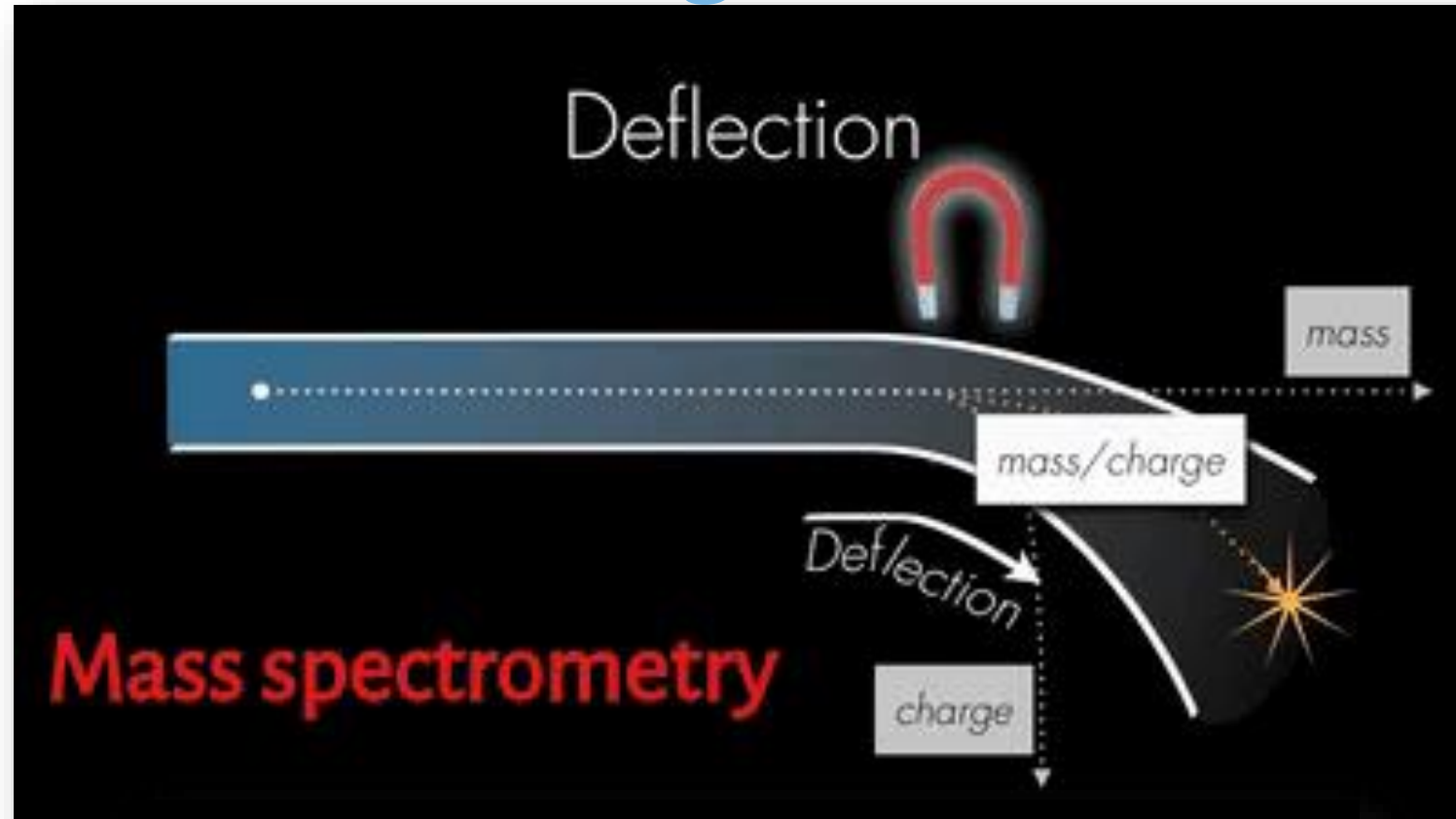
By their mass to
charge ratios
(m/z or e) and
relative
abundances



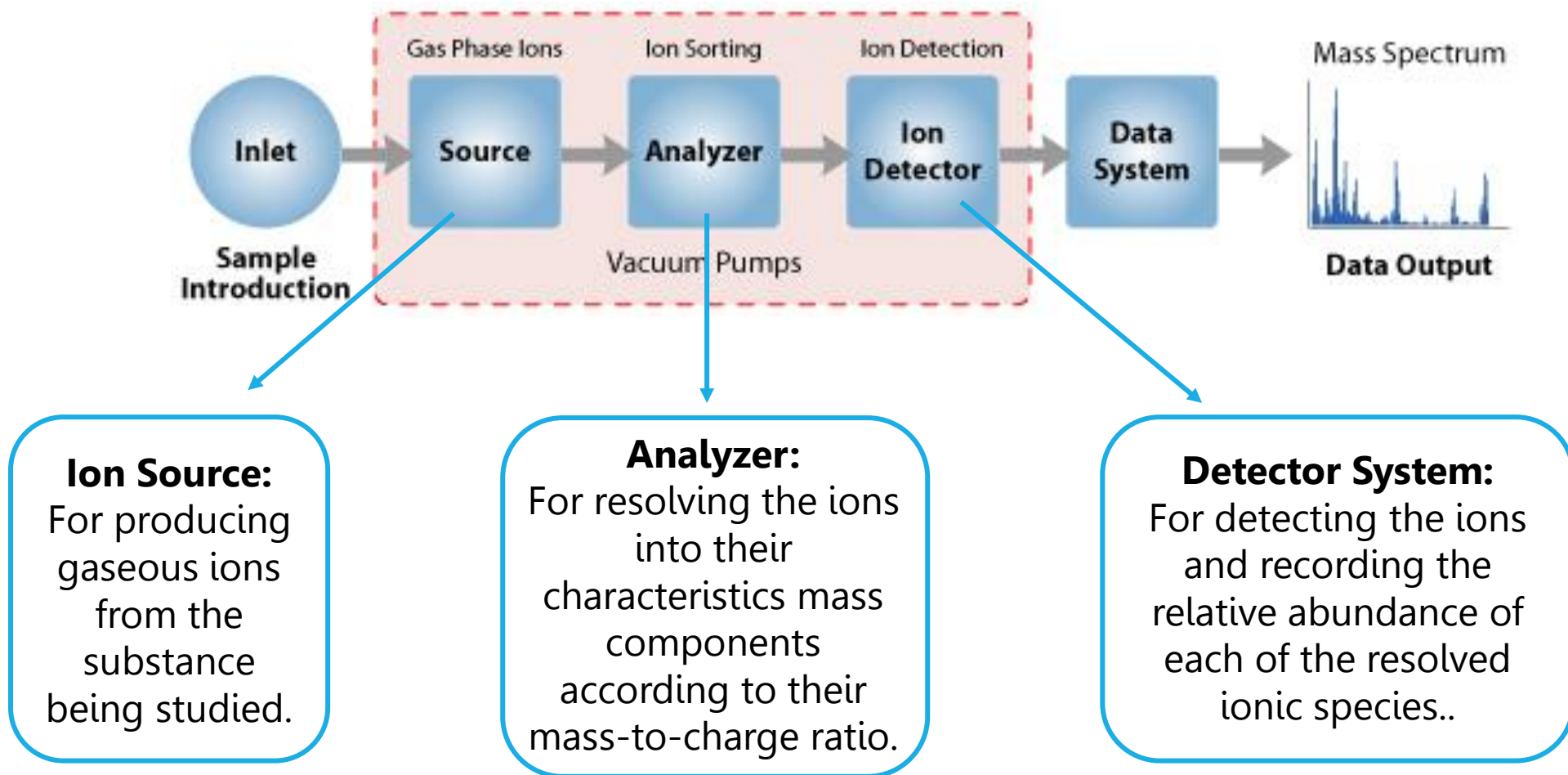
How is it working?



How is it working?



What is it ?



Ion Sources

GAS PHASE METHOD

- ***Electron Ionization (EI)***: Harsh method of molecule fragmentation and ionization and is used for relatively volatile and low molecular weight sample.
- ***Chemical ionization (CI)***: Very soft ionization technique and does not lead to extensive fragmentation.
- ***Direct analysis in real time (DART)***: Ability to analyze materials of different shapes and sizes with no prior sample preparation and in ambient conditions.
- ***Inductively coupled plasma (ICP)***: Ability to ionize almost all elements

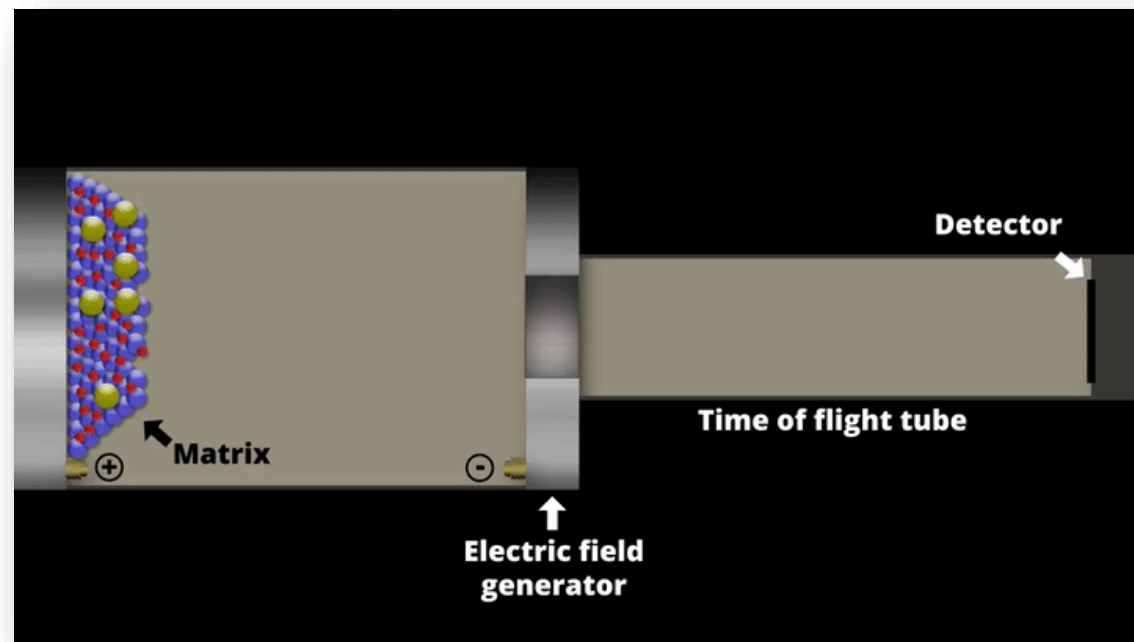
DESORPTION METHOD

- ***Matrix assisted laser desorption ionization (MALDI)***: One of the major "soft" ionization methods, particularly useful for the analysis of large or labile molecules.
- ***Fast atom bombardment (FAB)***: Soft ionization technique, able to produce positively and negatively charged ions.
- ***Thermal ionization***: The most common primary ion source and can be focused with electrostatic ion optics for secondary ion MS.
- ***Plasma ionization***: Commonly used to produce beams of gaseous ions.
- ***Liquid metal ion (LMIS)***: Ion beam with the smallest spot sizes and highest brightness, particularly advantageous in MS imaging.

SPRAY METHOD

- ***Electrospray ionization (ESI)***: Soft ionization technique suitable for analysis of large molecules and macromolecules.
- ***Desorption electrospray ionization (DESI)***: Droplets are directed to a sample held at ambient pressure.

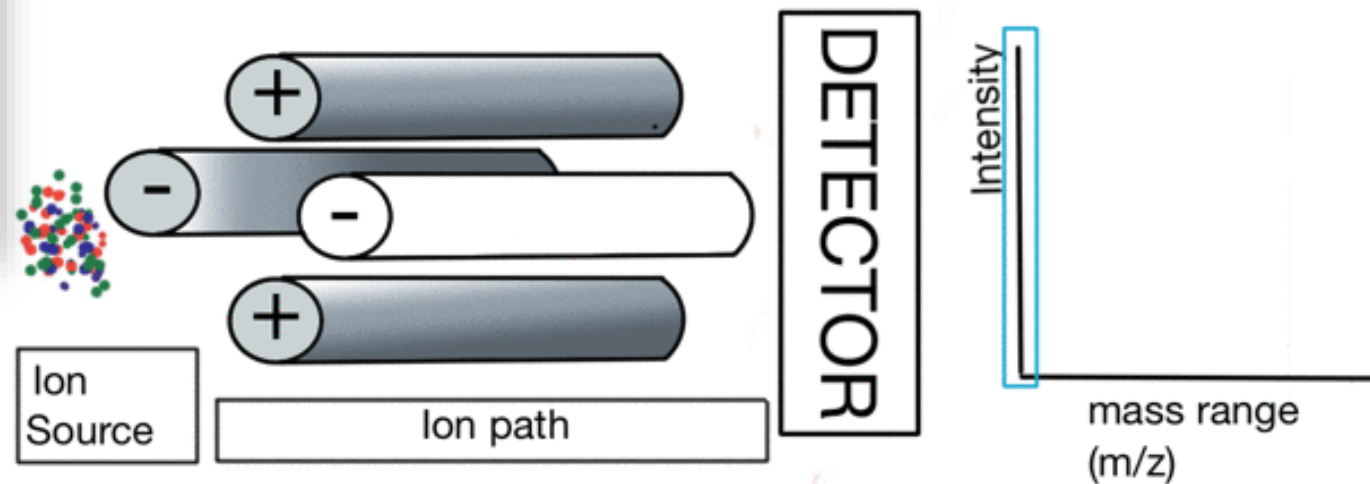
MASS ANALYZERS



TOF

Quadrupole

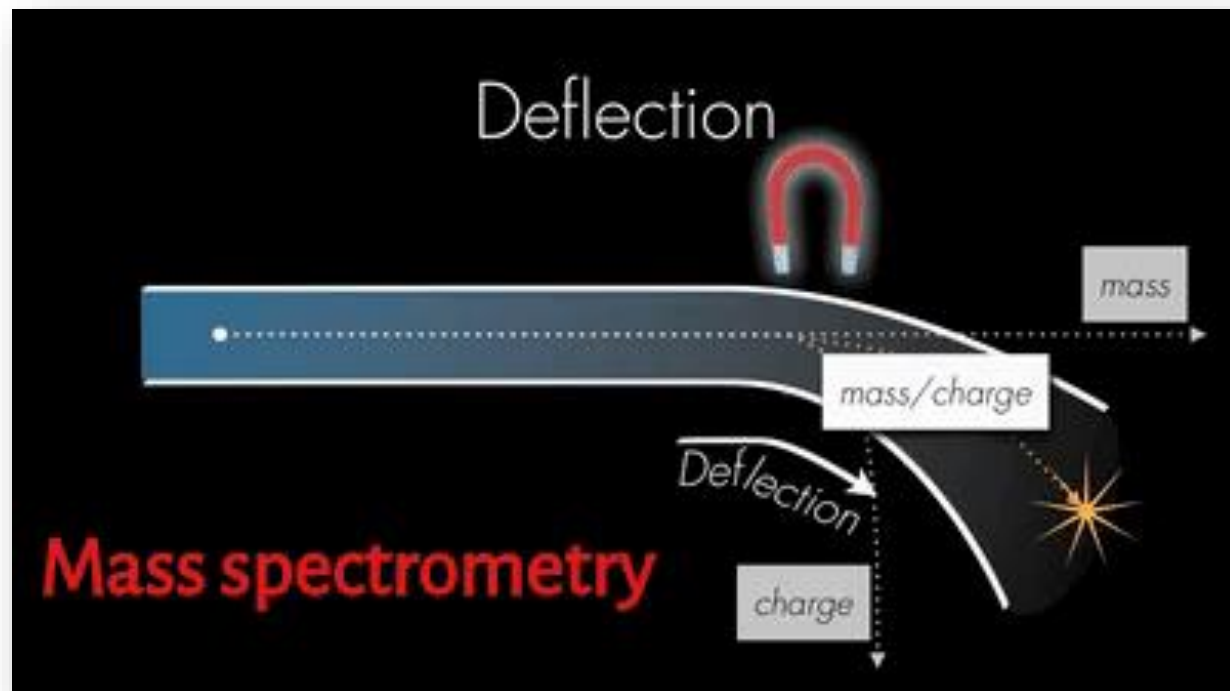
The quadrupole **SCANS** through a range of conditions...



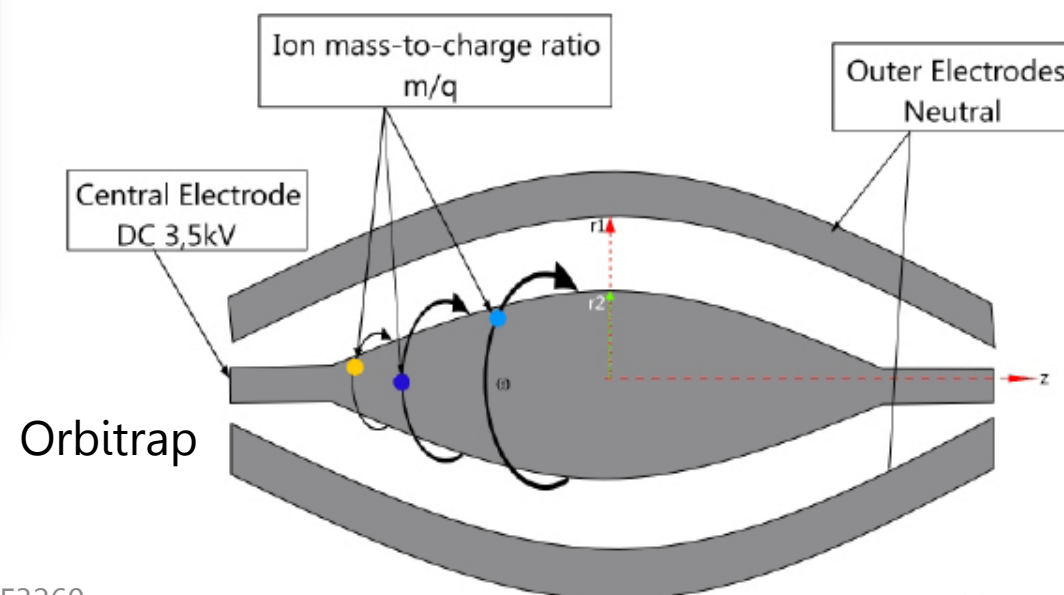
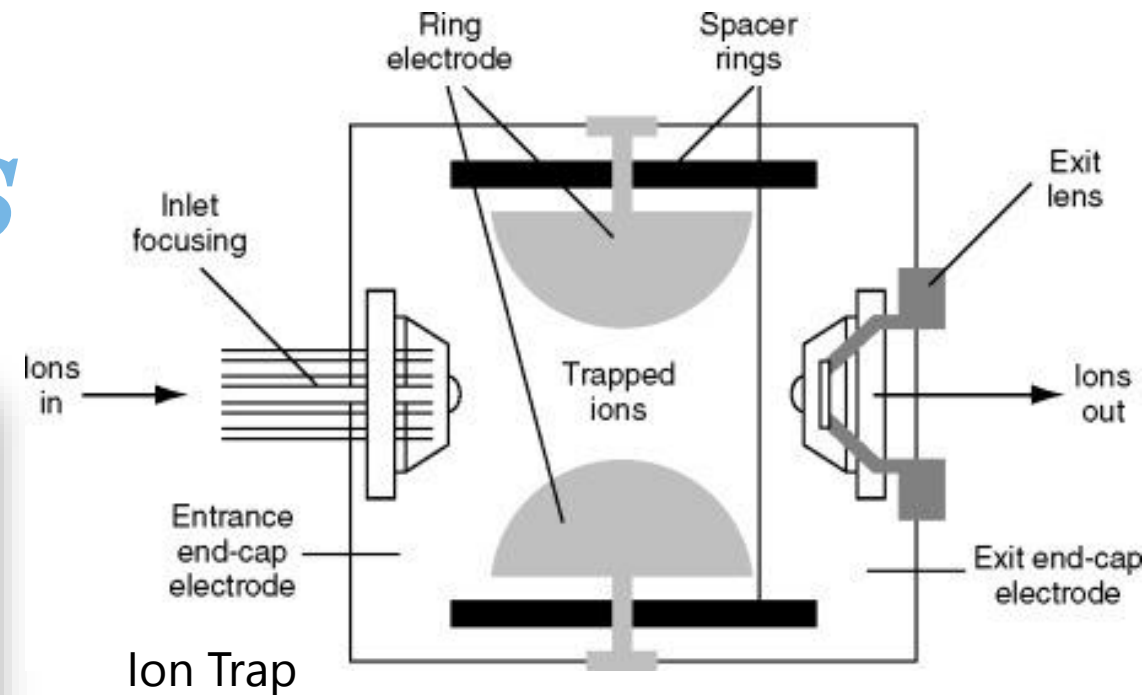
... that lets ions of a specific mass to charge ratio reach the detector.

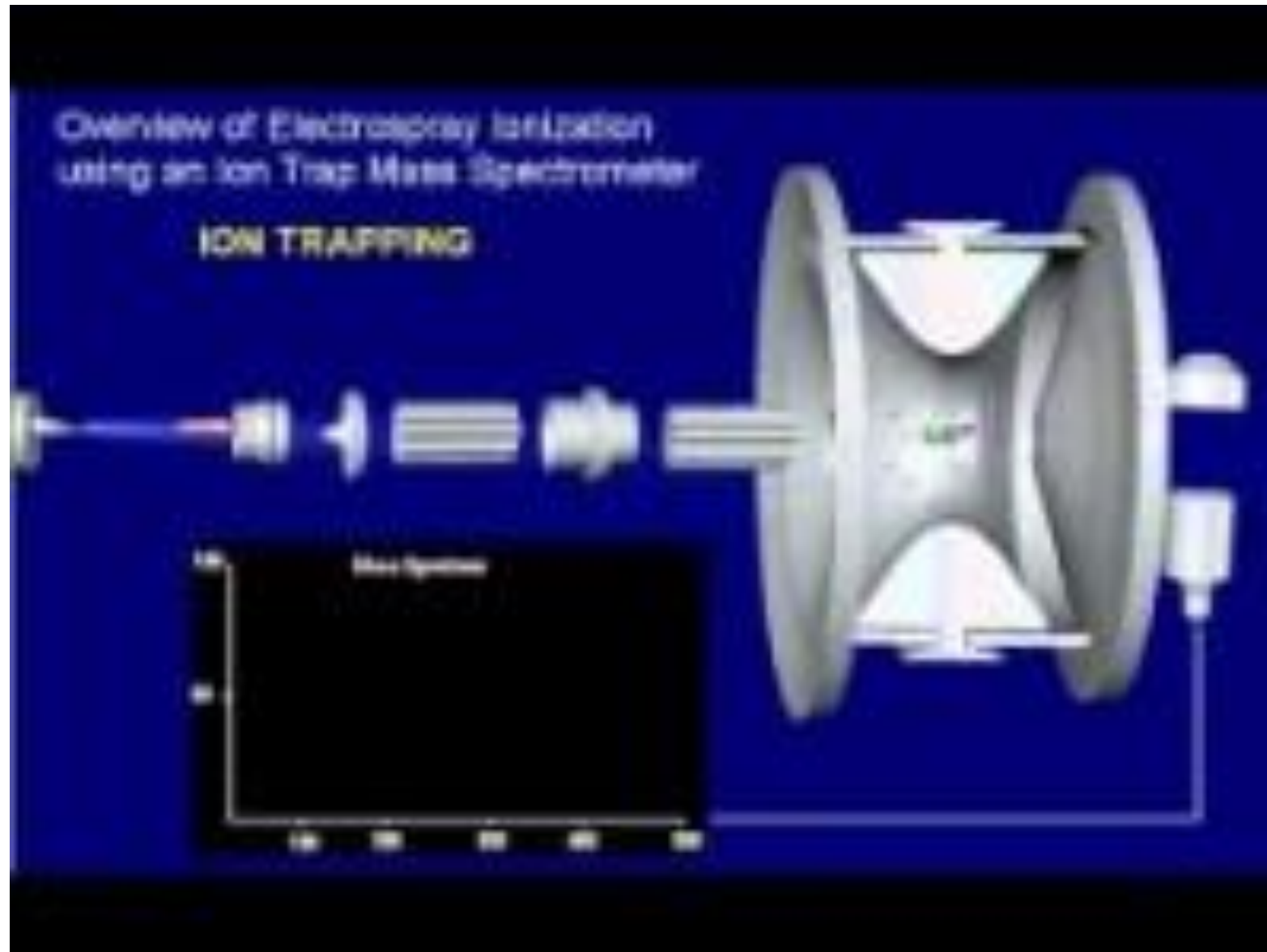
All the other ions go to waste (over a few milliseconds).

MASS ANALYZERS

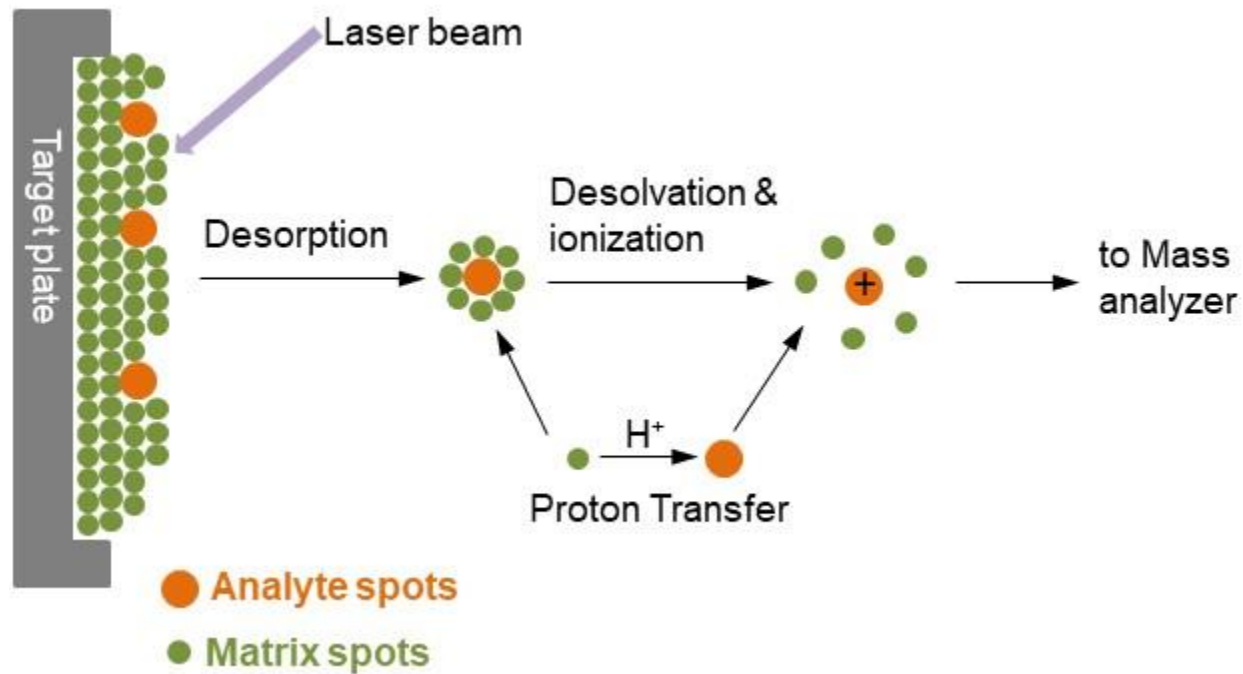


Magnetic Sector

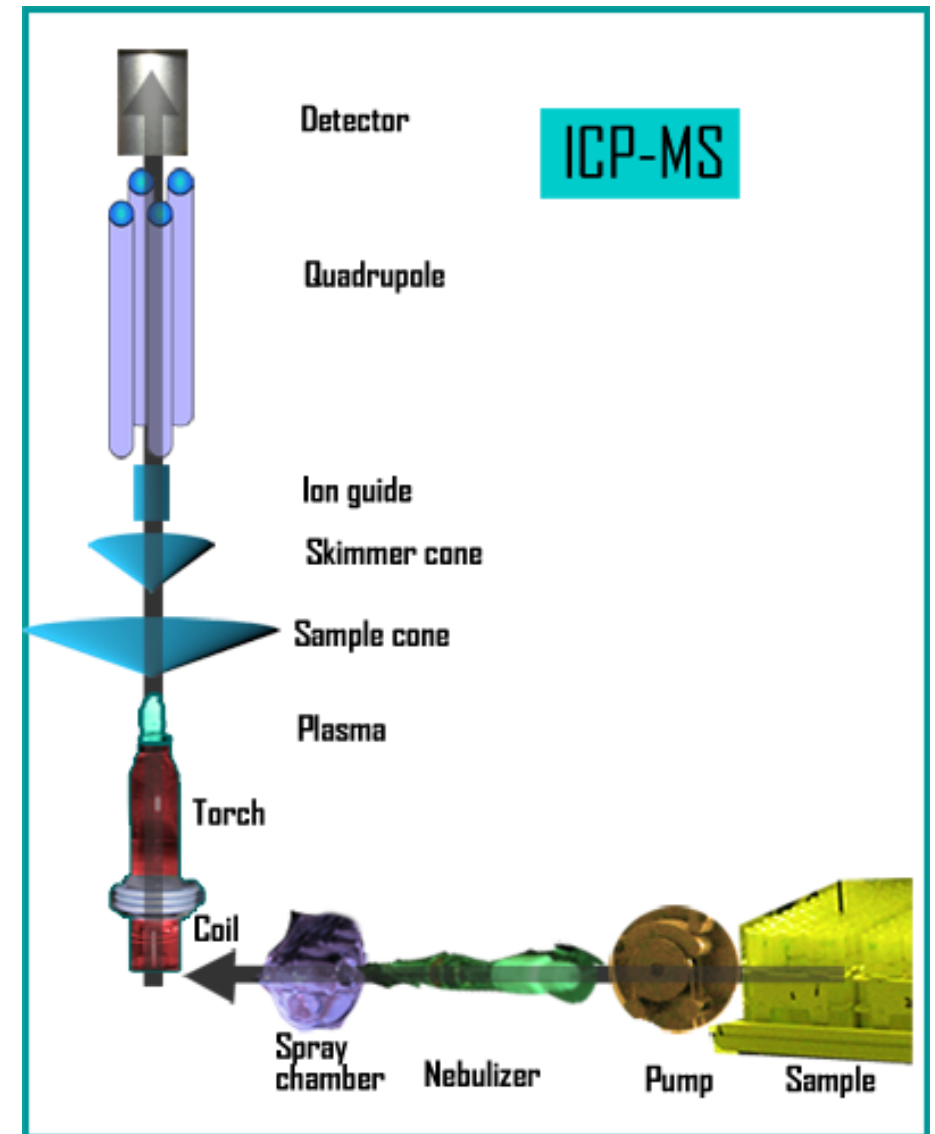




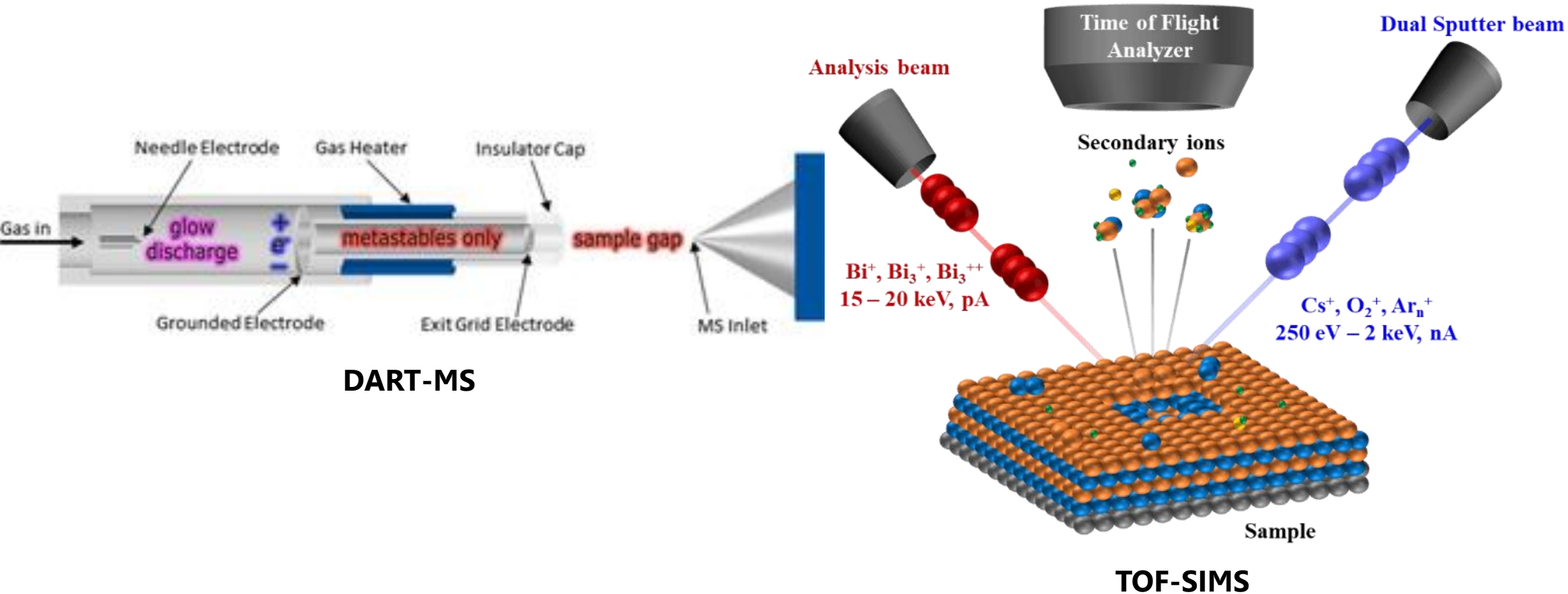
Pairing ionization techniques with mass analyzers



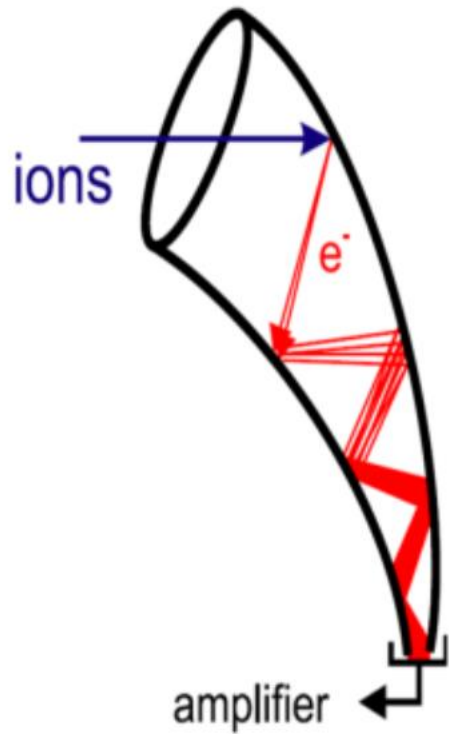
MALDI-TOF



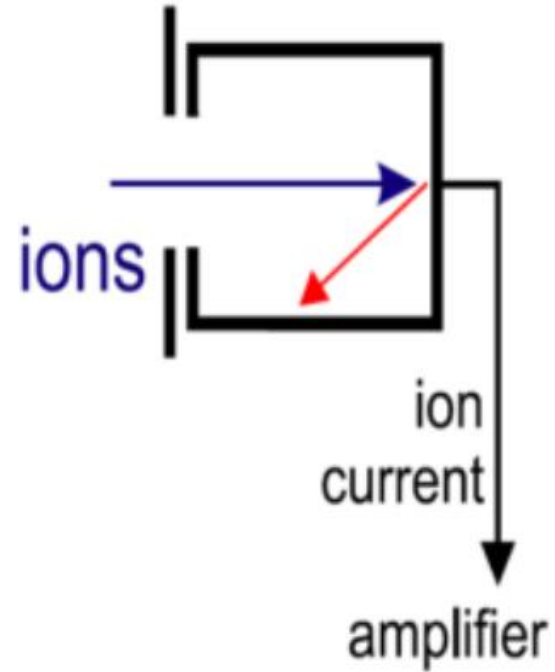
Pairing ionization techniques with mass analyzers



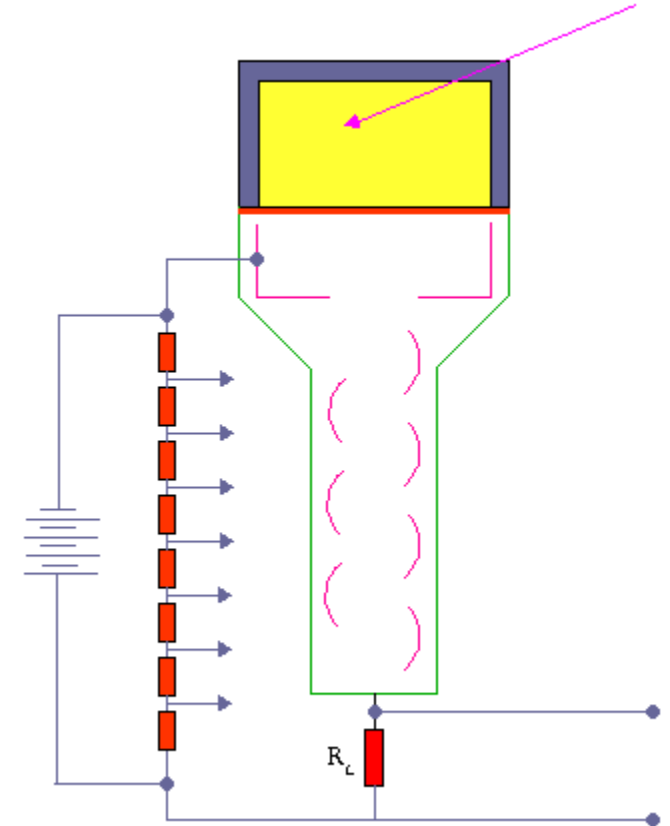
MASS DETECTORS



Electron Multiplier

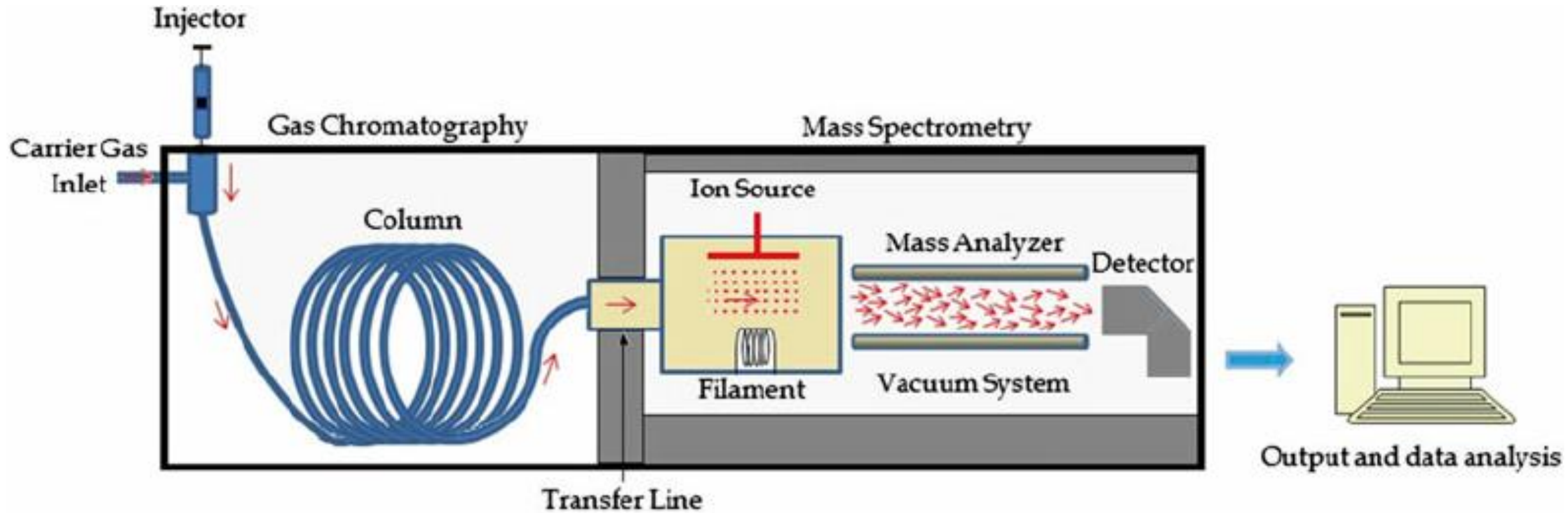


Faraday Cup

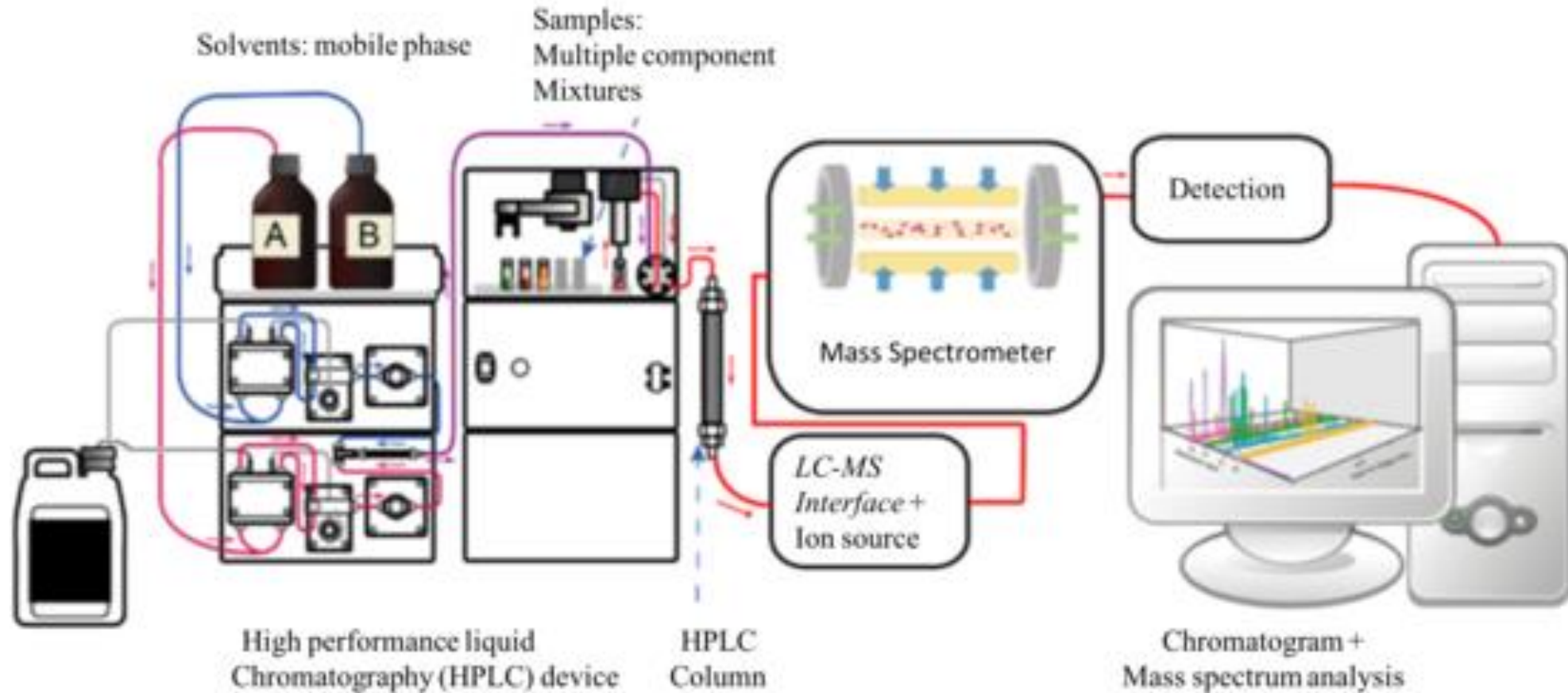


Photomultiplier Conversion Dynode

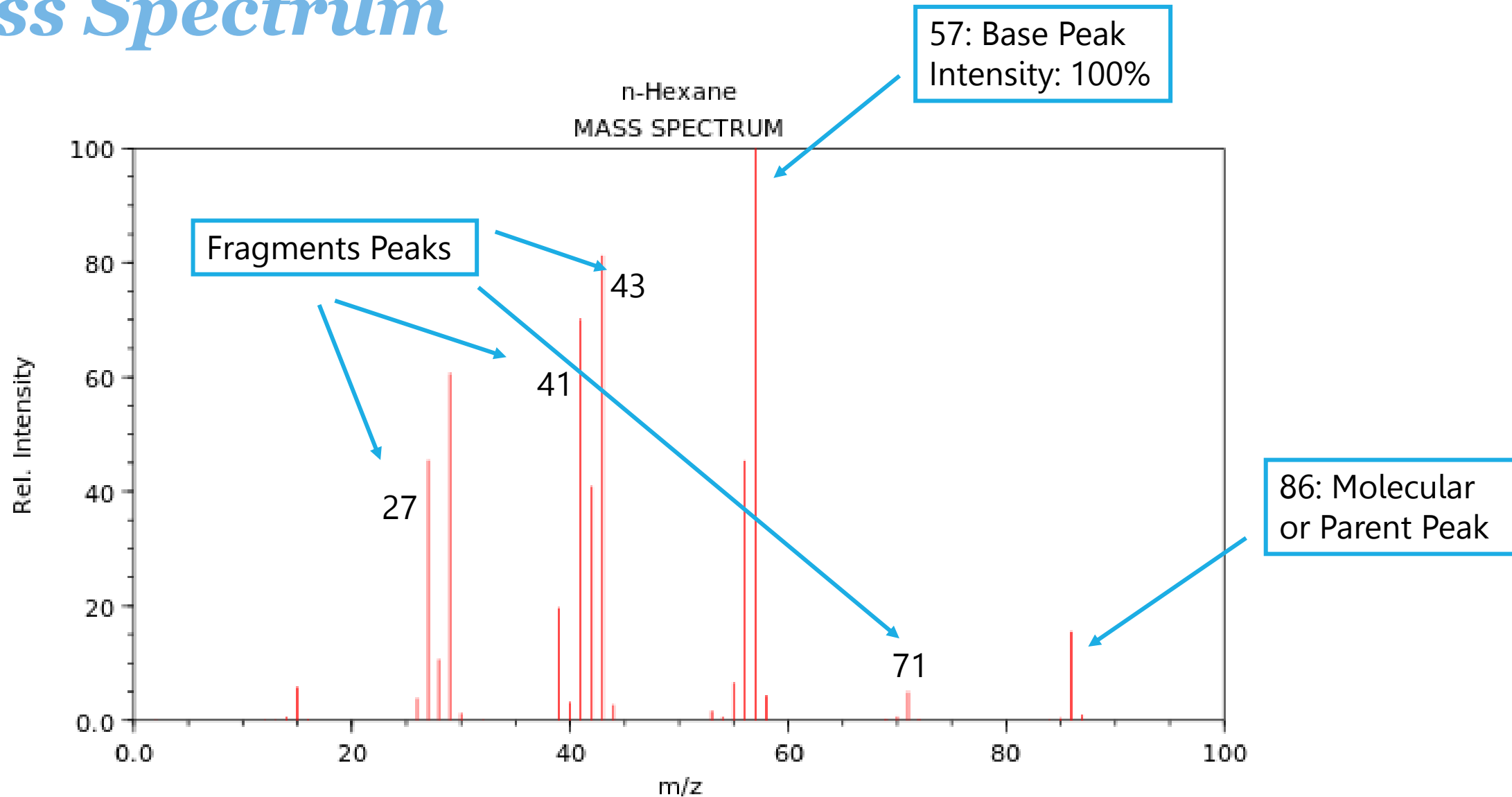
Combining mass spectrometers with other techniques: GC-MS



Combining mass spectrometers with other techniques: LC-MS



Mass Spectrum



MS And Biomolecules

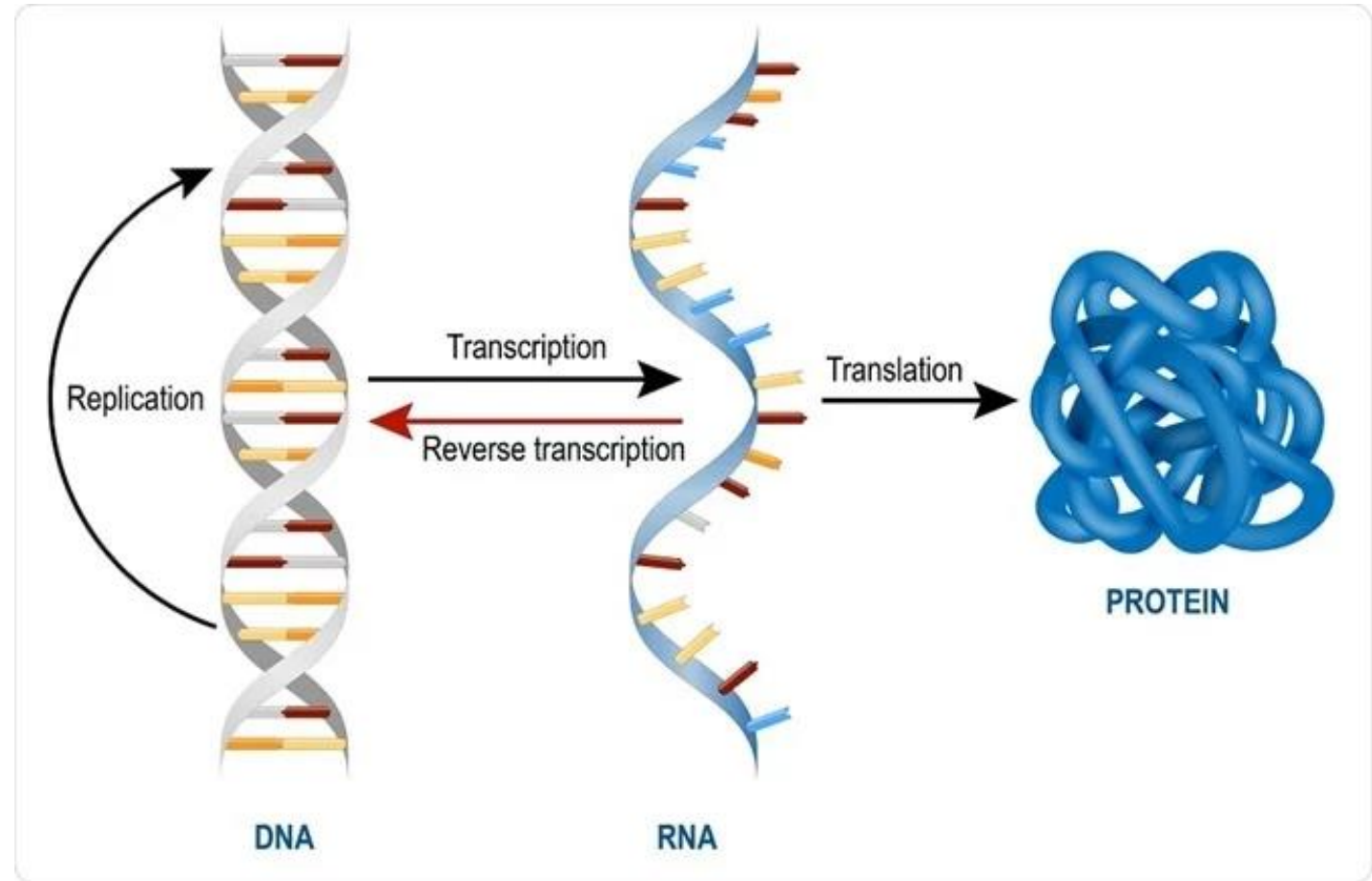
Proteomics

- Determine protein structure, function, folding and interactions.
- Identify a protein from the mass of its peptide fragments.
- Detect specific post-translational modifications throughout complex biological mixtures using workflows for phosphoproteomics and protein glycosylation.
- Quantitate proteins (relative or absolute) in a given sample.
- Monitor enzyme reactions, chemical modifications and protein digestion.

<i>Polypeptide sequence</i>	<i>Mass/amu</i>	<i>Δ</i>	<i>Amino acid</i>
X-Ser-Gly-Trp-Glu-Asp-Leu-Ile-Lys-Met	10531	131	Met
X-Ser-Gly-Trp-Glu-Asp-Leu-Ile-Lys	10400	128	Lys/Gln?
X-Ser-Gly-Trp-Glu-Asp-Leu-Ile	10272	113	Leu/Ile?
X-Ser-Gly-Trp-Glu-Asp-Leu	10159	113	Leu/Ile?
X-Ser-Gly-Trp-Glu-Asp	10046	115	Asp
X-Ser-Gly-Trp-Glu	9931	129	Glu
X-Ser-Gly-Trp	9802	186	Trp
X-Ser-Gly	9616	57	Gly
X-Ser	9559	87	Ser
X	9472		

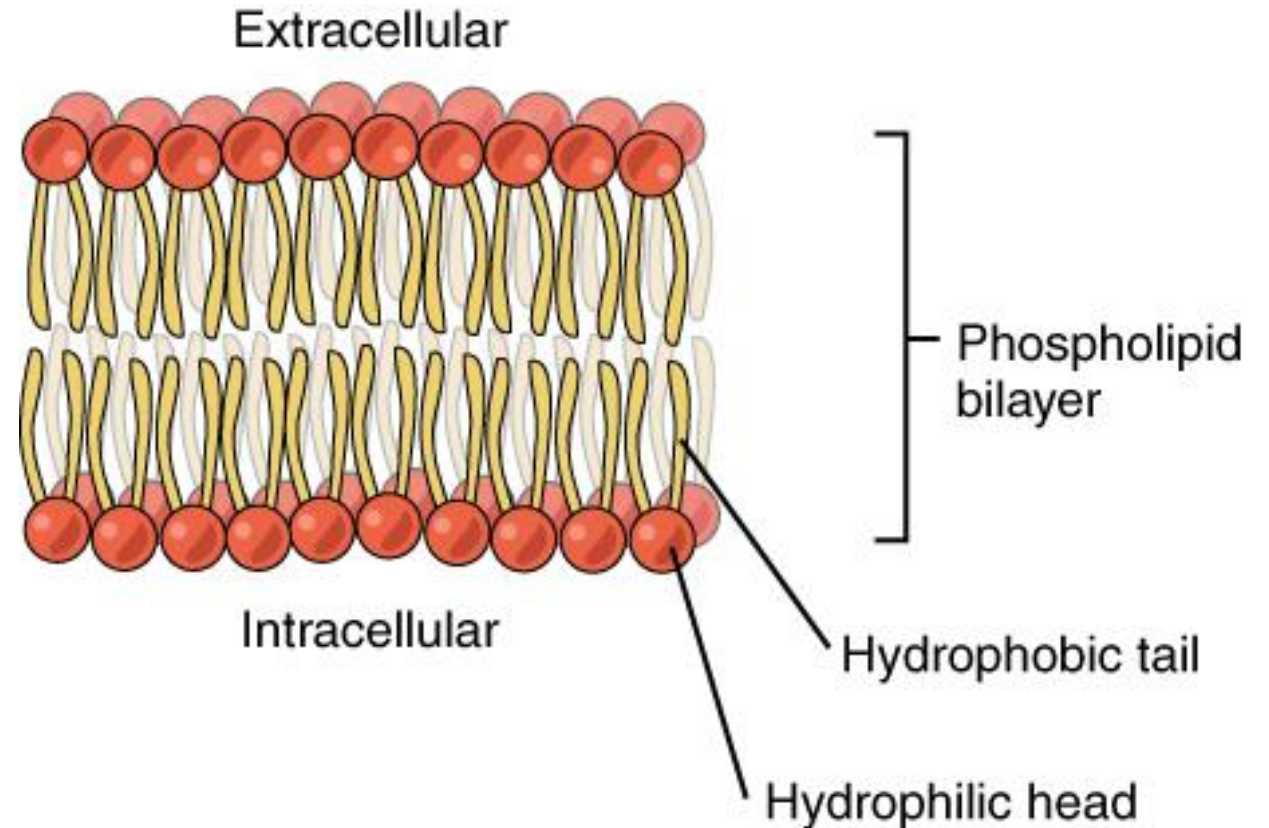
Oligonucleotides

- Identify the covalent modifications and determining their structure
- Identify their position in the oligonucleotide
- Determination of the molecular weight of oligonucleotides
- Determination of their sequences



Lipidomics

- Determine the molecular weight
- Determine elemental composition
- Determine the position of branching
- Determine nature of substituents in the lipid structure.



Applications

- Quality control
- Identify structures of biomolecules
- Sequence biopolymers
- Determination of the molecular mass of Biomolecules
- Monitoring gases in patients' breath during surgery.
- Identification of drug abuse and metabolites of drugs of abuse in blood, urine, and saliva.
- Analyses of aerosol particles.
- Determination of pesticides residues in food.

Advantages and Limitations

Advantages:

- Extremely sensitive
- Identify unknown components
- Combining with other techniques
- Very precise and rapid
- Very small sample quantities
- Gives the relative molecular mass of every molecule

Drawbacks:

- Costly
- Need a skilled technician
- Not a portable system
- Unable to differentiate among isomers of the molecule with the same charge-to-mass ratio
- Chiral columns may be required to separate enantiomer
- Difficult to recognize hydrocarbons that generate parallel ions
- Unable to separate optical and geometric isomers

Any Questions ?



In-Class Activity

Here you have the mass spectrum of the pentane (C_5H_{12}), consisting of CH_3 and CH_2 groups, try to explain me the spectrum obtained.

