CHEM-E4109 MODERN METHODS IN **BIOCATALYSIS**

chapter #6: Aldol chemistry

Jan Deska Bioorganic Chemistry

www.deskalab.com

Updated Schedule

- 1.3. Introduction & basic principles
- 4.3. Oxidoreductases I (reduction catalysis)
- 8.3. Oxidoreductases II (oxygenation catalysis)
- 11.3. Transferases (transaminases)
- 15.3. Lyases I (decarboxylases and transketolases)
- 18.3. Lyases II (aldolases)
- 22.3. Directed evolution (no live session, video only)
- 25.3. Hydrolases (lipases, esterases, epoxide hydrolases)
- 29.3. Muta- and semisynthesis
- 1.4. Enzymes in non-natural reactions

Lyases - features

enzymes of class EC 4 catalyze the cleavage of chemical bonds (non-hydrolytic!) (...and of course, like always, some can do the reverse reaction)



Aldol-catalyzing enzymes

- mainly involved in biosynthesis and degradation of sugars and related biomolecules
- 2 major classes that differ in their basic activation mode
 - ✓ class I mechanism: enamine catalysis or
 - ✓ class II mechanism: enolate catalysis



Aldol-catalyzing enzymes

- mainly involved in biosynthesis and degradation of sugars and related biomolecules
- 2 major classes that differ in their basic activation mode
 - ✓ class I mechanism: enamine catalysis or
 - ✓ class II mechanism: enolate catalysis



 current main drawback: narrow scope of accepted donors (nucleophiles)

class I aldolase: enamine catalysis



class I aldolase: enamine catalysis



class I aldolase: enamine catalysis



class I aldolase: enamine catalysis



class II aldolase: enolate catalysis



class II aldolase: enolate catalysis



class II aldolase: enolate catalysis



class II aldolase: enolate catalysis



2-Deoxyribose-5-phosphate aldolase

DERA (Lactobacillus brevis)

- homo dimer, 259 amino acids per subunit (other organisms exploit tetrameric DERAs)
- class I aldolase
- independent on cofactors or metal ions
- physiological role: catabolism of glycosides and deoxyribonucleotides
- catalyzes the reversible cross-aldol reaction between <u>acetaldehyde</u> and other acceptor aldehydes



Synthesis of the statin core structure



Jan Deska Bioorganic Chemistry

Greenberg, Varvak, Hanson, Wong, Huang, Chen, Burk, Proc. Natl. Acad. Sci. 2004, 101, 5788-5793.

Synthesis of azasugars



Dean, Greenberg, Wong, Adv. Synth. Catal. 2007, 349, 1308-1320.

Synthesis of azasugars



Dean, Greenberg, Wong, Adv. Synth. Catal. 2007, 349, 1308-1320.



Dean, Greenberg, Wong, Adv. Synth. Catal. 2007, 349, 1308-1320.

Enamine-based biomimetics as the mother of Organocatalysis

based on stoichiometric enamine chemistry



Yamada, 1969: asymmetric enamine reaction (not yet catalysis)



The Hajos-Parrish-Eder-Sauer-Wiechert reaction

first highly enantioselective organocatalytic reaction



The Hajos-Parrish-Eder-Sauer-Wiechert reaction

first highly enantioselective organocatalytic reaction



The Hajos-Parrish-Eder-Sauer-Wiechert reaction

first highly enantioselective organocatalytic reaction



high enantioselectivity (List 2000)



List, Lerner, Barbas, *J. Am. Chem. Soc.* **2000**, *122*, 2395-2396. Northrup, MacMillan, *J. Am. Chem. Soc.* **2002**, *124*, 6798-6799.

high enantioselectivity (List 2000)



with α -substituted donors: high anti-diastereoselectivity (MacMillan 2002)



List, Lerner, Barbas, *J. Am. Chem. Soc.* **2000**, *122*, 2395-2396. Northrup, MacMillan, *J. Am. Chem. Soc.* **2002**, *124*, 6798-6799.

Nobel Prize 2021 in Chemistry

"for the development of asymmetric organocatalysis"



Benjamin List



Max-Planck-Institut Mülheim, Germany



David W. C. MacMillan

Princeton, USA

Houk, Martin, List, J. Am. Chem. Soc. 2003, 125, 2475-2479.

origin of stereoselectivity (simplified Houk-List model)



origin of stereoselectivity (simplified Houk-List model)



Houk, Martin, List, J. Am. Chem. Soc. 2003, 125, 2475-2479.

origin of stereoselectivity (simplified Houk-List model)



Jan Deska Bioorganic Chemistry

Houk, Martin, List, J. Am. Chem. Soc. 2003, 125, 2475-2479.

origin of stereoselectivity (simplified Houk-List model)



Jan Deska Bioorganic Chemistry

Houk, Martin, List, J. Am. Chem. Soc. 2003, 125, 2475-2479.

syn-selective aldol reactions

by stabilization of anti-Z-enamine



81%, 92% ee, 88% syn



Jan Deska Bioorganic Chemistry

Ramasatry, Zhang, Tanaka, Barbas J. Am. Chem. Soc. 2007, 129, 288-289.

syn-selective aldol reactions

by stabilization of anti-Z-enamine



81%, 92% ee, 88% syn



Jan Deska Bioorganic Chemistry

Ramasatry, Zhang, Tanaka, Barbas J. Am. Chem. Soc. 2007, 129, 288-289.

syn-selective aldol reactions

by destabilization of Zimmerman-Traxler arrangement



Jan Deska Bioorganic Chemistry

Kano, Yamaguchi, Maruoka, Angew. Chem. Int. Ed. 2007, 46, 1738-1740.

replacement of acceptor = other α -functionalizations of aldehydes and ketones

Imine acceptors = Mannich reaction



Jan Deska Bioorganic Chemistry

Hayashi, Tsuboi, Ashimine, Urushima, Shoji, Sakai, Angew. Chem. Int. Ed. 2003, 42, 3677-3680.

replacement of acceptor = other α -functionalizations of aldehydes and ketones

Imine acceptors = Mannich reaction



Jan Deska Bioorganic Chemistry

Hayashi, Tsuboi, Ashimine, Urushima, Shoji, Sakai, Angew. Chem. Int. Ed. 2003, 42, 3677-3680.

proline as template for the development of next generation organocatalysts



alpha-functionalization of aldehydes and ketones



alpha-functionalization of aldehydes and ketones



alpha-functionalization of aldehydes and ketones



alpha-functionalization of aldehydes and ketones



alpha-functionalization of aldehydes and ketones



alpha-functionalization of aldehydes and ketones



enamine activation in radical chemistry



enamine activation in radical chemistry



Beeson, Mastracchio, Hong, Ashton, MacMillan, Science 2007, 316, 582-584.

Remember the acyloin reaction?

Acyloin reaction

- PDCs also catalyze C-C-coupling between α-ketoacids and aldehydes
- biochemical equivalent to the Benzoin reaction



Remember the acyloin reaction?

Acyloin reaction

- PDCs also catalyze C-C-coupling between α-ketoacids and aldehydes
- biochemical equivalent to the Benzoin reaction



- formal addition of HCN yields cyanohydrines
- analogous to Stetter amino acid synthesis

Cyanohydrine-converting enzymes

- hydroxynitrile lyases (EC 4.2.1.x) mainly found in higher plants
- main role: defence mechanisms based on cyanogenesis
- 2 major classes that differ in their dependence on FAD
 - ✓ FAD not acting as redox mediator in these enzymes



cheap sources for HNLs: almonds, cherry, manioc, rubber tree



mechanism in e.g. manioc and *Hevea* sp.

Jan Deska Bioorganic Chemistry



mechanism in e.g. manioc and *Hevea* sp.

> Jan Deska Bioorganic Chemistry



mechanism in e.g. manioc and *Hevea* sp.

> Jan Deska Bioorganic Chemistry



mechanism in e.g. manioc and *Hevea* sp.

> Jan Deska Bioorganic Chemistry

Synthetic applications: addition of HCN to aldehydes



calcium pantothenate: important animal feed additive

> Jan Deska Bioorganic Chemistry

Bracco, Busch, von Langermann, Hanefeld, Org. Biomol. Chem. 2016, 14, 6375-6389.

Synthetic applications: addition of HCN to aldehydes



Jan Deska Bioorganic Chemistry

Bracco, Busch, von Langermann, Hanefeld, Org. Biomol. Chem. 2016, 14, 6375-6389.

Synthetic applications: HCN-free hydrocyanations?

acetone cyanohydrine as masked HCN



Jan Deska Bioorganic Chemistry

Bracco, Busch, von Langermann, Hanefeld, Org. Biomol. Chem. 2016, 14, 6375-6389.