CHEM-E4109 MODERN METHODS IN **BIOCATALYSIS**

chapter #10: enzymes in non-natural reactions

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That's nice, right? But how far can you go?

Synthetic Biology & Metabolic Engineering can...

- effectively regulate pathways that are intrinsic in life
- access structures that already found somewhere in nature

Synthetic Biology & Metabolic Engineering fails to...

- provide solutions for truly synthetic targets
- offer bio-based solutions for many traditional chemistries
- engage in anything that lacks precedence in biosynthesis

SCOPE & LIMITATIONS OF ENZYMES

The bright side...

- highly productive enzymatic systems for the preparation of chiral building blocks, e.g. alcohols, amines, amino acids,...
- mild reaction conditions in absence of hazardous solvents or reagents
- modern biotechnology allows for effective catalysts tuning
- easy to combine multiple biocatalysts one-pot: design of catalytic cascades

SCOPE & LIMITATIONS OF ENZYMES

The dark side...

- few privileged enzymatic transformations of synthetic relevance
- some biosynthetically encoded but inadequately applicable reactions
- many abiotic purely synthetic molecular manipulations

limited reaction portfolio results in limited applicability in synthetic strategies

Biosynthesis vs chemical synthesis

- few privileged enzymatic transformations of synthetic relevance
- **some** biosynthetically encoded but inadequately applicable reactions
- **many** abiotic purely synthetic molecular manipulations
- = limited reaction portfolio results in limited applicability in synthetic strategies



Biosynthesis vs chemical synthesis

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De novo design

- identification of configurations of low-energy transition states
- mapping of a general binding pocket architecture



generic transition state



functional groups and orientation to stabilize the envisioned transition state

De novo design

- identification of configurations of low-energy transition states
- mapping of a general binding pocket architecture
- ✓ computer-guided selection of stable protein scaffolds to match the necessary shape
- ✓ introduction of required amino acid functionalities to the positions



Baker et al, *Science* 2010, *329*, 309



Nature as Inspiration



Ronald Breslow

"In Biomimetic Chemistry, we take what we have observed in Nature and apply its principles to the invention of novel synthetic compounds that can achieve the same goals... As an analogy, we did not simply make larger versions of birds when we invented airplanes, but we did take the idea of the wing from Nature, and then used the aerodynamic principles in our own way to build a jumbo jet."

Synthetic compounds that mimic biological materials' functions or properties:

- bioactivity (medicinal chemistry)
- light-activated (photovoltaics & photocatalysis)
- natural binding modes (organocatalysis)

Reverting biomimetics?

- modern synthetic organic chemistry as a blueprint for natural catalysts
- chemoinspired artificial functions of biocatalysts



Chemistry

a) Coelho, Brustad, Kannan, Arnold, *Science* 2013, *339*, 307-310; b) Coelho, Wang, Ener, Baril, Kannan, Arnold, Brustad, *Nature Chem. Biol.* 2013, *9*, 485-487.

General concept for cyclopropanation, alkylation, amination...



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Zhang et al. Nature 2019, 67

Directed evolution as key for effective biocatalysts





Zhang et al. Nature 2019, 67

Sure... but why bother

key chemical transformations



unprecedented in nature, yet template for the design of new enzymes

targeted activity mining & evolution

novel enzymatic activities



artificial biocatalytic modules enabling non-natural transformations

Sure... but why bother





ex vivo cascade design



synthetic tools combining native and abiotic functions

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artificial biocatalytic modules enabling non-natural transformations

Sure... but why bother



Traditional organic synthesis

• 2017: Independent syntheses of Angiopterlactone B by Lawrence and Bhattacharya



Thomson, Nichol, Lawrence, *Org. Lett.* **2017**, *19*, 2199-2201 Kottamagari, Gonnade, Bhattacharya, *Org. Lett.* **2017**, *19*, 3564-3567

Our assembly line in action

Achmatowicz'ase glucose oxidase chloroperoxidase D-Glc purely enzymatic method for conversion of biorefinery furans Thiel, Doknic, Deska, Nature Commun. **2014**, *5*, 5278 Borrowing Hydrogen'ase alcohol dehydrogenase NADP+ HO (+)-walterolactone enantioconvergent isomerization of Achmatowicz pyranones

Liu, Merten, Deska, *Angew. Chem. Int. Ed.* **2018**, *57*, 12151



What are interesting reactions to aim for?

(no need to agree with me on that one)

Pericyclic reactions?

Three kinds of pericyclic reactions

- Common features
 - ✓ all pericyclic reaction are concerted (= electron reorganization takes place in one single step)
 - either thermal or photochemical reactions
 - highly selective and generally not affected by catalysts or solvents
- Cycloaddition reactions
 - ✓ reaction between two pi-containing molecules to form a cyclic product



- \checkmark two fewer pi bonds in the product
- two additional sigma bonds in the product

Pericyclic reactions?

Three kinds of pericyclic reactions

- Common features
 - ✓ all pericyclic reaction are concerted (= electron reorganization takes place in one single step)
 - either thermal or photochemical reactions
 - highly selective and generally not affected by catalysts or solvents
- Sigmatropic rearrangements
 - intramolecular migration of groups by rearrangement of pi bond(s)



- \checkmark a sigma bond is broken while a new sigma bond forms
- ✓ a pi bond, or a conjugated pi system in the product
- ✓ reversibility! check equilibrium position

Pericyclic reactions?

Three kinds of pericyclic reactions

- Common features
 - ✓ all pericyclic reaction are concerted (= electron reorganization takes place in one single step)
 - either thermal or photochemical reactions
 - highly selective and generally not affected by catalysts or solvents
- Electrocyclic reactions
 - ✓ intramolecular reaction where a conjugated linear pi system is closed at the ends



- \checkmark a new sigma bond is formed, one pi bond less in the product
- ✓ reversibility! check equilibrium position

Biological mimicry of catalytic principles relevant for pericyclic reactions

Here: two conceptional work-in-progress projects

 Sigmatropic rearrangements by hijacking copper-dependent monooxygenases CH amination by a biocatalytic alternative to allylpalladium- or nitrene insertion-type C-N bond formation



Oxidative sp_3 -CH amination strategies

via allylpalladium species



biochemically challenging!

via nitrene insertion



enzymatically implemented by using sulfonyl azides

Arnold et al., *Angew. Chem. Int. Ed.* **2013**, *52*, 9309-9312

Frauenhoffer, White, *J. Am. Chem. Soc.* **2007**, *129*, 7274-7276. Zalatan, Du Bois, *J. Am. Chem. Soc.* **2008**, *130*, 9220-9221.

Oxidative sp³-CH amination strategies

via Alder-ene reaction of N-containing enophiles



Palmer, Frazier, Read de Alaniz, *Synthesis* **2014**, *46*, 269-280 (review). Morozova, Shumakovich, Shleev, Yaropolov, *Appl. Biochem. Microbiol.* **2007**, *43*, 523-535 (review).

Oxidative sp³-CH amination strategies

via Alder-ene reaction of N-containing enophiles



 hydroxamic acid derivatives and hydroxylamines commonly used as redox mediator in enzyme-driven aerobic oxidation reactions



Palmer, Frazier, Read de Alaniz, *Synthesis* **2014**, *46*, 269-280 (review). Morozova, Shumakovich, Shleev, Yaropolov, *Appl. Biochem. Microbiol.* **2007**, *43*, 523-535 (review).

A Copper-dependent Alder-Enase?

O N H H		enzyme(s) H ₂ O		-OH		
	0	25 I	50 I	75 I	100	
laccase						conversion
(<i>R. vernicifera</i>)						selectivity
laccase						
(A. bisporus)						
laccase						
(1. versicolor)						
laccase (CotA)						
(B. licheniformis)						
chloroperoxidase*						
(<i>C. fumago</i>)						
horseradish						
peroxidase*						* - with alucese 8

* = with glucose & glucose oxidase

A Copper-dependent Alder-Enase?

O H H H	¹ / ₂ O ₂	enzyme(s) H ₂ O		-OH		
	0	25 I	50	75 I	100 J	
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laccase (<i>T. versicolor</i>)						
laccase (CotA) (<i>B. licheniformis</i>)						
chloroperoxidase* (<i>C. fumago</i>)						_
horseradish peroxidase*						*
						= WITN GIUCOSE &

giucose oxidase

Two potent biocatalysts for C-N bond formations



CotA yield: 51%

CotA yield: 59%

HRP yield: 77% CotA yield: 63%

Towards a mechanistic understanding

enzyme-induced vs enzyme-assisted





diffusion of nitroso species?

small molecules as electron mediators?

intimate copper-substrate interaction

SET's vs 2e⁻ processes?

Pericyclic vs addition/elimination

Kinetic & stereochemical studies

regioselectivity: defined rearrangement, no free radical intermediates



Kinetic & stereochemical studies

regioselectivity: defined rearrangement, no free radical intermediates



enantioselectivity: not awesome (but that's what we have directed evolution for, right?)



pH 7.0: CotA = 2% *ee*, Tv = 5% *ee* pH 5.5: CotA = 27% *ee*, Tv = 43% *ee*

Summary 'Alder enase'

- ✓ exploiting intrinsic oxidase activity for generation of reactive intermediates
- ✓ highly effective biocatalyst for C-N bond forming reactions without biosynthetic precedence
- ✓ rational activity mining based on knowledge of laccase's reactivity principles
- o directed evolution required to solve existing challenges (e.g. enantioselectivity)

Sigmatropic reactions beyond chorismate mutase

(hopefully more versatile than native mutases)

Onium ylide formation as interesting target reaction

historical, Stevens (1928): 1,2-benzyl shift of ammonium ylides



Kirmse (1968): oxonium ylide formation from metal carbenes: the basics...



... and synthetic applications



1) Stevens, Creighton, MacNicol, *J. Chem. Soc.* **1928**, 3193; 2) Kirmse, Kamps, *Chem. Ber.* **1968**, *101*, 949; 3) Hodgson, Angrish, Erickson, Kloesges, Lee, *Org. Lett.* **2008**, *10*, 5553.

Copper oxidases as artificial Kirmse mutases?



Pathways for diazo activation



major challenge:

- oxonium ylides not necessarily compatible with aqueous media
- with water being an absolute prerequisite for those kinds of proteins









Role of free copper or other proteins



- shutdown of product formation
- no free copper ions responsible for observed effect

- inverted selectivity with free copper or Cu-leaching proteins
- no reactivity with recombinant galactose oxidase !!!

What else could be involved



• other low MW Copper Oxidase?

Collection of protein fractions by UPLC

- ✓ GalOx region shows no Kirmse activity
- only the low retention first fractions
 exhibit the previously observed effect
 from the crude GalOx mixture

rich impurity pattern in commercial GalOX
 no similarities to recombinant *F. gr.* GalOx

The most likely candidate

- *Fusarium graminearum* encodes for at least 18 lytic polysaccharide monooxygenases
- carbohydrate-active mononuclear copper enzymes with *auxiliary activity* (AA9, AA10, AA11)
- cleave cellulose and related biopolymers (chitin, chitosan, xyloglucan,...)



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Vaaje-Kolstad, Westereng, Horn, Liu, Zhai, Sørlie, Eijsink, Science 2010, 330, 219-222.

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Vaaje-Kolstad, Westereng, Horn, Liu, Zhai, Sørlie, Eijsink, *Science* **2010**, *330*, 219-222.



So how would a random LPMO perform?

The most likely candidate

- Fusarium graminearum encodes for at least 18 lytic polysaccharide monooxygenases
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- cleave cellulose and related biopolymers



Vaaje-Kolstad, Westereng, Horn, Liu, Zhai, Sørlie, Eijsink, *Science* **2010**, *330*, 219-222.



Chasing the actual catalyst

- reconstruction of initial reactivity by recombinant & purified enzymes
 but
- selectivity and activity still behind the crude enzyme system



- selection of gel cut-outs with characteristic protein fractions
- 2 tryptic digest to yield peptide fragments
- 3 MS/MS analysis & bioinformatics

hits in section III with high level of confidence for: cellulose-active LPMO (AA9) & chitin-active LPMO (AA11)

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production of two new family 11 LPMOs in P. pastoris

Chitinase as the real biocatalyst?

chitinase closely related to the tested PcLPMO9



Hemsworth, Henrissat, Davies, Walton, Nat. Chem Biol. 2014, 10, 122-126.



Chitinase as the real biocatalyst?

chitinase closely related to the tested PcLPMO9









prefers polymeric chitin



prefers polymeric cellulose



prefers small soluble chitin oligos!

Chitin-active monooxygenases as Kirmse mutase



- recombinant system matches the previously successful commercial black box almost spot-on
- ✓ highly effective (TTN ~ 20 000)

Substrate scope



Summary 'Kirmse mutase'

- ✓ oxidative enzyme as catalyst in non-oxidative transformation
- stabilization of highly reactive intermediates that seem incompatible with aqueous media
- ✓ prime example for serendipity in science
- ✓ complete failure of rational design

That's it... We're almost done

Thank you for your participation and your input through the learning diaries

What's left to do?

- prepare your seminar presentations
- join the seminar on your day of choice (13.4. or 14.4.)
- please answer the Webropol feedback survey (available now, until 27.4.2021)