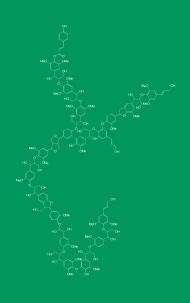


Lignin: Structure and characterization

Eero Kontturi Slides: Mika Sipponen D.Sc. (Tech.) Department of Forest Products Technology School of Chemical Technology Aalto University



Learning outcomes

- Where does lignin come from?
- Principles of lignin biosynthesis
- Classification of lignins
- Importance of lignin to plants
- Distribution of lignin in plant cell walls
- Overview of reactions under thermochemical conditions
- Industrial sources & relevance
- Isolation and characterization
- Current and emerging applications



Occurrence



Lignin

- Main component of plant biomass (in addition to cellulose and hemicelluloses)
- Makes 25-35% of wood dry weight and 15-25% in annual plants
- Structurally, it is a polydisperse mixture of methoxylated polyphenols
- Naturally brown, hence the colour of wood
- Dissolved from wood chips in chemical pulp production: combusted for chemical recovery and heat and power production
- New uses for lignin are sought in materials and chemicals

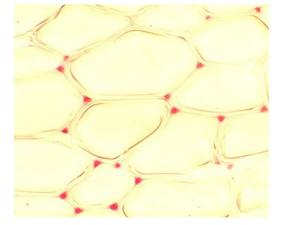


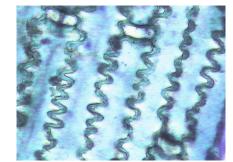




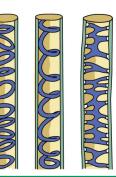
Occurrence and functions in plants

- Lignin is a natural resin that fills spaces between plant cells and strengthens cell walls by covering cellulose microfibrils
- Functions of lignin in plants:
 - Provides strength
 - Barrier against the attack of insects and microbes
 - Enables transportation of water and ions from the soil





Wheat straw xylem tissue seen in optical microscope

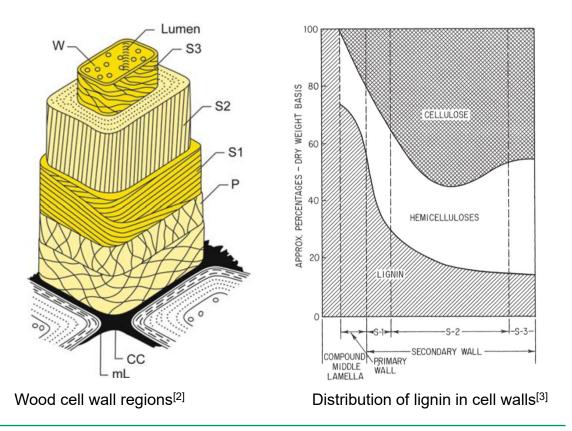


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http://sydney.edu.au/science/biology/learning/plant_form_function/revision_modules/2003A_Pmodules/module2/2AD1.shtml http://elte.prompt.hu/sites/default/files/tananyagok/StructureOfPlantsAndFungi/ch04s04.html

Distribution of lignin in plant cell walls

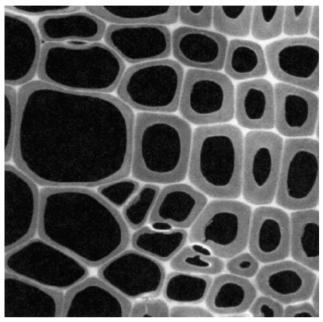
- CC, ML enriched in lignin, presumably because lignification is initiated there^[1]
- Secondary walls contain the largest proportion of the total lignin in cell walls



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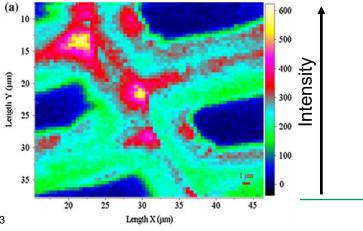
^[1] Donaldson, Phytochem. 57 (2001) 859–873
 ^[2] Agarwal, Planta (2006) 224:1141–1153 (ref. Sjöström, 1993)
 ^[3] Hale, 1969 "Structural and physical properties of pulpwood"

Imaging distribution of lignin in plant cell walls



Bright areas in pine wood arise from autoflurorescense of lignin^[1]

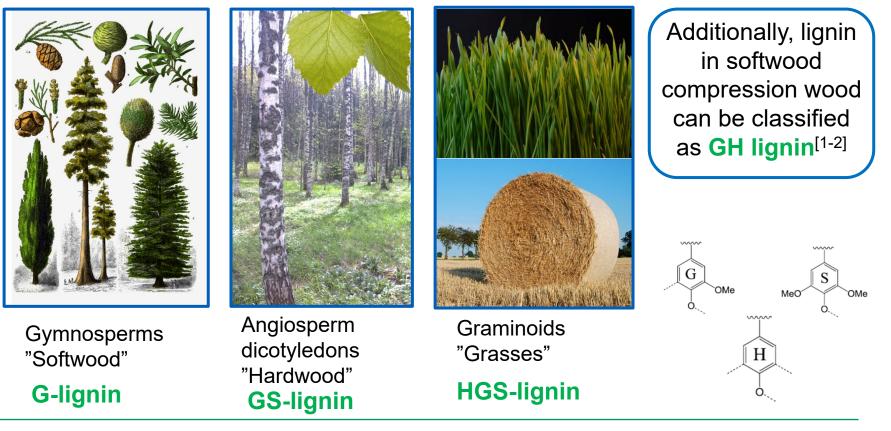
Raman imaging of spruce lignin at the area of six mature cells^[2]



21.9.2022

Aalto University School of Chemical Technology ^[1] Donaldson, Phytochem. 57 (2001) 859–873
 ^[2] Agarwal, Planta (2006) 224:1141–1153

Botanical classification of lignins



Aalto University School of Chemical Technology [1] Nimz et al., Holzforschung 35 (1981) 16-26.
 [2] Rolando et al., Thioacidolysis, Methods in lignin chemistry, 1992, pp. 334-349 https://www.ecn.nl/uploads/pics/Grote_rol_tarwestro_NW_01.jpg
 https://upload.wikimedia.org/wikipedia/commons/6/68/Grass_dsc08672-nevit.jpg
 https://upload.wikimedia.org/wikipedia/commons/6/68/Grass_dsc08672-nevit.jpg
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 https://upload.wikimedia.org/wikipedia/commons/thumb/4/40/Gymnospermae.jpg/330px-Gymnospermae.jpg
 http://c.80.138.132/skolor/skolbilder/filer/e0842627cbc4dc6bf2bbb27b66622986.Julgran.png
 http://www.giftofatree.ie/wp-content/uploads/2014/01/Betula-pendula-Youngii-Leaf.jpg

Biosynthesis and structure

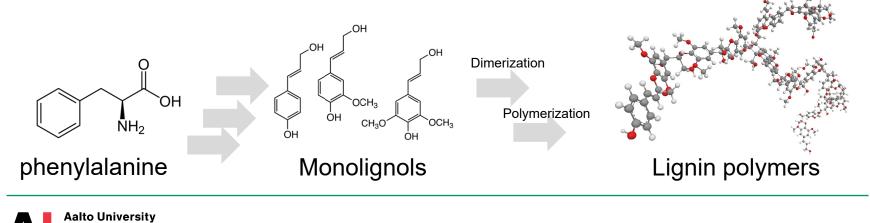
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Biosynthesis and the diversity of molecular structures

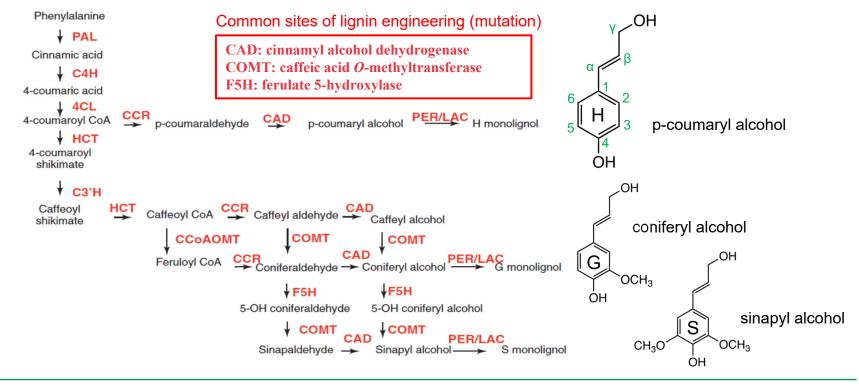
- Growing trees and other plants do not "just grow" they are living organisms with subtle control over biosynthesis
- Lignin is synthesized principally from three precursor monomers which are referred to as monolignols
- Amino acid phenylalanine is the starting point:

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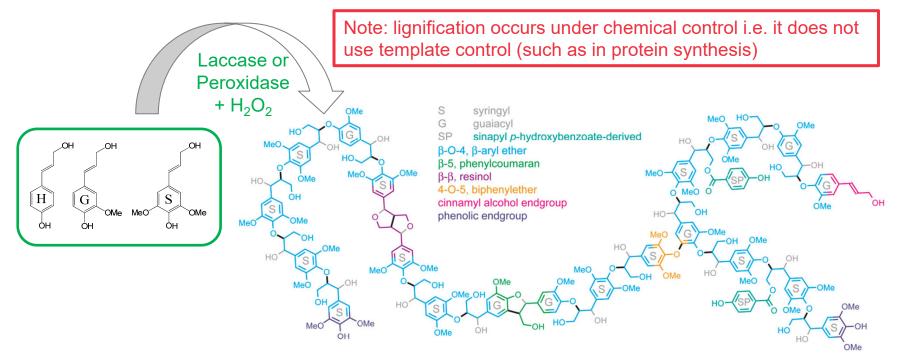


Several enzymes are needed in the monolignol synthesis



Aalto University School of Chemical Technology Zhao and Dixon, Trends Plant Sci. 2011

Lignin polymer is formed from monolignols in enzyme-initiated radical polymerization

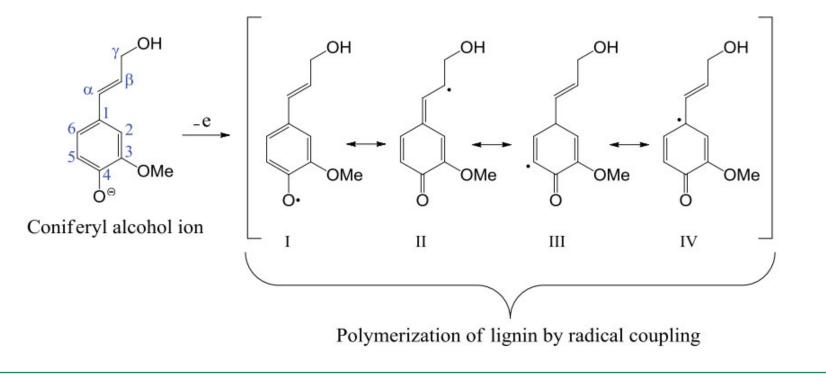


Result = mixtures of molecules (not one well-defined compound)



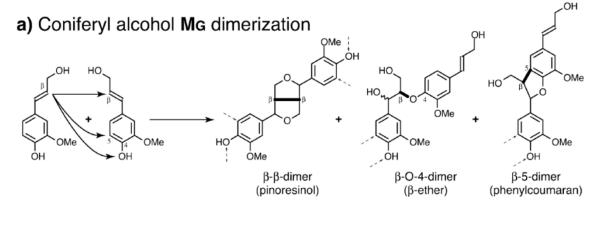
Vanholme et al., Plant Physiol. 153 (2010) 895-905.

Example of radical delocalization

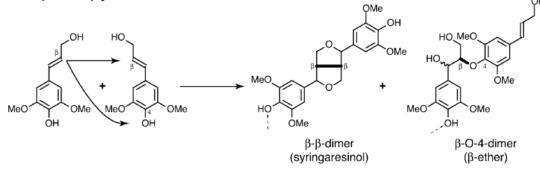


Aalto University School of Chemical Technology Sipponen, Doctoral thesis 2015

Dimerization reactions of monolignols



b) Sinapyl alcohol Ms dimerization



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Ralph et al., Phytochem. Rev. 3: 29-60, 2004.

Polymerization reactions of monolignols

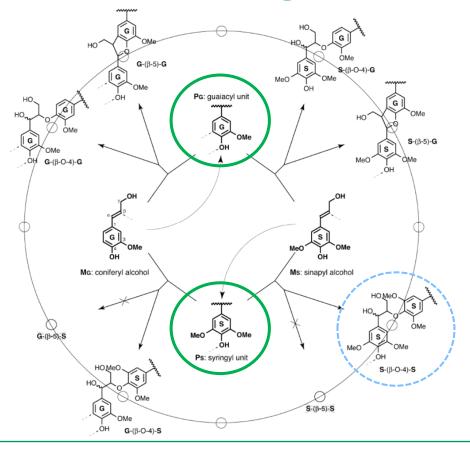
Radical coupling:

- Monolignols react endwise with the growing polymer
- The resulting structure of the lignin macromolecule is "random" in the sense that several linkage types are possible at each step
- S-S coupling is an exception since this predominantly gives rise to β-O-4 units

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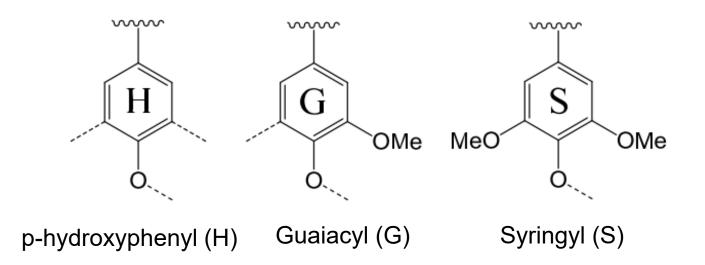
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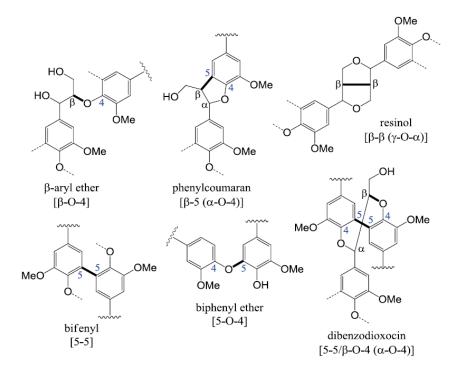
Ralph et al., Phytochem. Rev. 3: 29-60, 2004.

Naming of the units in the lignin polymer



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Major structural units of lignin polymer



 Bolded bonds are formed in radical coupling and other in dimerization reactions

Linkage type	Dimer structure	Approximate percentage
β-0-4	Phenylpropane β-aryl ether	45-50
β- <i>O</i> -4 α- <i>O</i> -4	Phenylpropane α -aryl ether	6-8
β-5	Phenylcoumaran	9-12
5-5	Biphenyl and dibenzodioxocin	18-25
4-0-5	Diaryl ether	4-8
β-1	1,2-Diaryl propane	7-10
β_β	β-β-Linked structures	3

 Units linked to another phenylpropane unit from the aryl ring positions 3 and 5 (2 and 6) are referred to as "condensed linkages"



Sipponen Doctoral thesis 2015 Chakar and Ragauskas Ind.Crops Prod. 20 (2004) 131-141.

d. 20 (2004) 131-141.

.OH

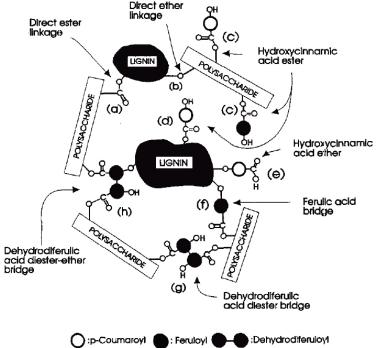
`OCH₃

G

OH

Covalent linkages between lignin and polysaccharides

- Lignin-carbohydrate complexes (LCCs) contain ether and ester bonds
- Linkages are mainly between
 hemicelluloses and lignin
- LCCs contribute to the cell wall integrity and difficulty of delignifying residual lignin^[1]
- Lignin carbohydrate network (LCN) must be disrupted for enzymatic saccharification of plant biomass



A schematic illustration of LC-linkages in grasses^[2]



^[1] Lawoko, Doctoral thesis, 2005 ^[2] liyama et al., Plant Physiol. 104 (1994) 315–320.

Fundamental properties and reactivity

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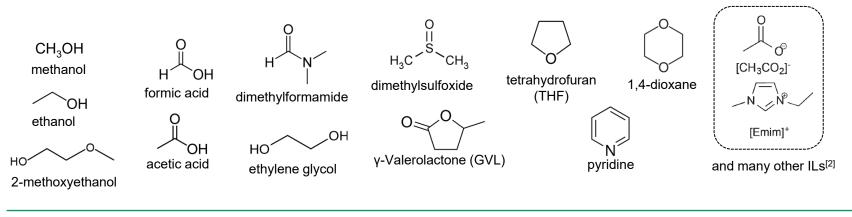
Industrial relevance

- Lignin is removed from fibers during cellulose pulp production
 - Provides surplus energy for the pulp mills
- Kraft, soda, sulfite, organosolv processes rely on distinct reactions
- Lignin is a structural constraint and inhibitor in biochemical conversion of plant biomass into ethanol etc.
- Biorefineries produce soluble and insoluble lignin
 - A spectrum of different hydrothermal and thermochemical lignocellulose pretreatment processes are under development



Fundamental properties and reactivity

- Lignins are negatively charged in aqueous solutions
 - pKa of phenolic hydroxyl groups vary between 7–10,^[1] depending on the neighbouring substituents; carboxylic acids have generally pKa < 4
- Solubility of lignin is important for industrial utilization and analytical work
 - Lignins are usually soluble in alkaline water and often (but not always) also in polar organic solvents and ionic liquids such as:

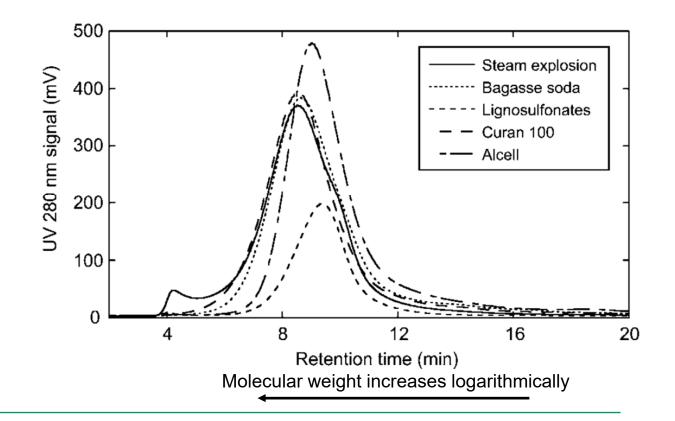




^[1] Ragnar et al., J. Wood Chem. Technol. 20, 277–305 ^[2] Hart et al., Green Chem., 2015, 17, 214–218

Polydispersity of lignin

- When isolated from wood, lignin contains molecules at various molecular weights
- Small lignin oligomers are naturally present also in plant biomass

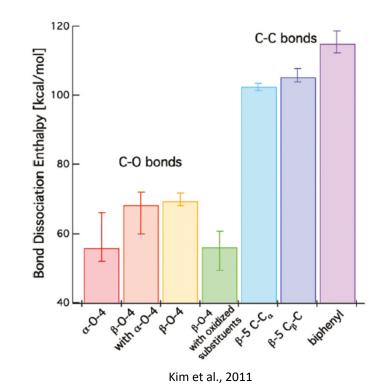




Baumberger et al., Holzforschung 61 (2007) 459-468

Stability of covalent bonds in lignin

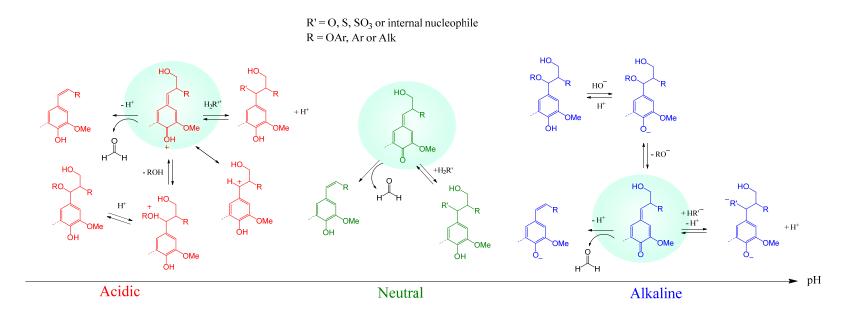
- Bond dissociation enthalpies (BDE) based on density functional theory calculations:
 - α -O-4 in GG pinoresinol, 284 kJ/mol^[1] β -O-4 (native) HG, 292 kJ/mol^[2] SS 295 kJ/mol^[3]
 - C–O are weaker than C–C bonds
 - Side-chain oxidation decreases BDE
- Note that lignin is more reactive in thermochemical conditions because of:
 - Formation of reactive quinone methide intermediates
 - Neighbouring group participation





[1] Elder, Energy Fuels 2014, 28, 1175–1182
 [2] Younker et al., ChemPhysChem 2011, 12, 3556 – 3565
 [3] Kim et al., J. Phys. Chem. Lett. 2011, 2, 2846–2852

Quinone methide intermediates are important reactive forms of lignin



Reactions of lignin through the quinone methide intermediate in acidic, neutral and alkaline media.^[1]

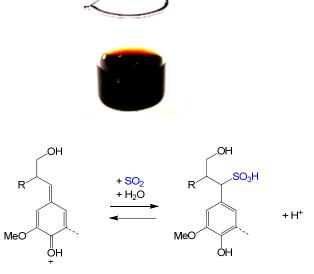


Lignin during pulping

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Sulfonation of lignin

- Formation in acid sulphite pulping
- Sulfoxyl groups (pKa<2) give water-solubility within a broad pH region
- Lignosulfonates are industrial chemicals
 - Plasticizers and additives in concrete and cement production
 - Viscosity reducing additive in oil drilling mud
 - Dispersants in pesticides, dyes, carbon black
 - Vanillin production (example follows later)



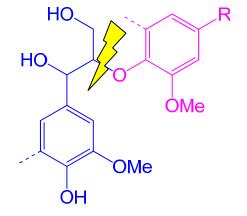
Sulfonation of lignin^[1]



^[1] J. Gierer, Wood Sci. Technol., 1985, 19, 289–312.

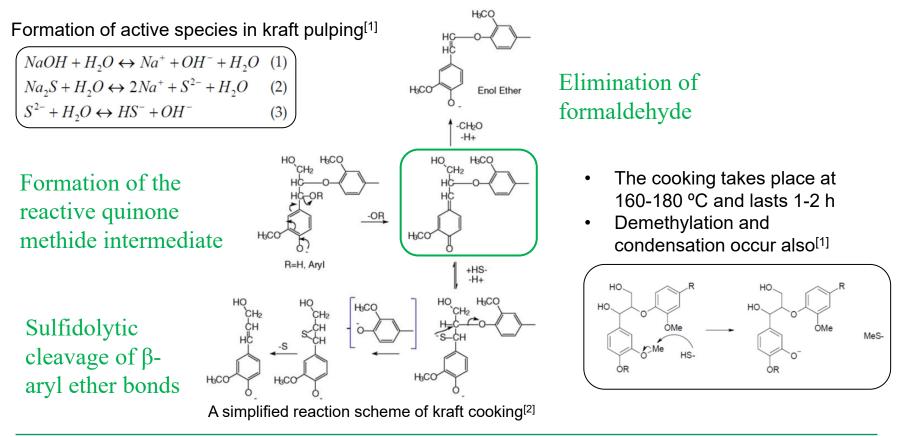
Cleavage of β–O–4 bonds in pulping processes

- The most important scission reaction in lignin chemistry, why?
 - Because β -O-4 bonds are so abundant and thermochemically labile
- Basis of delignification (depolymerisation
 → dissolution) in cellulose pulp production
 - Increases the content of phenolic hydroxyls
 - Side reactions form new C–C bonds





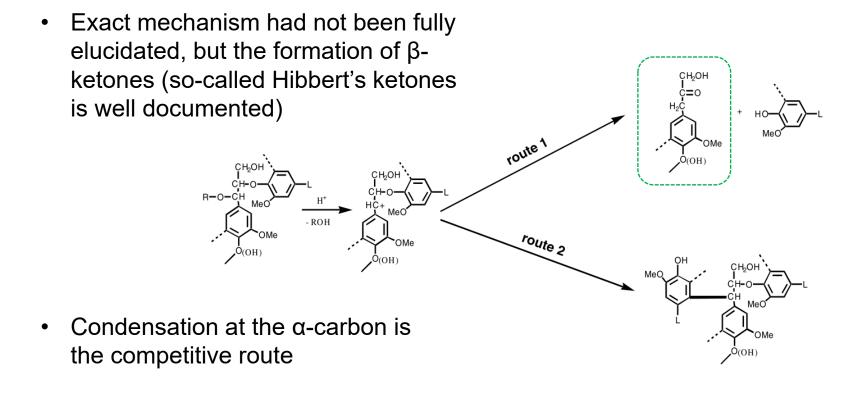
β-O-4 cleavage during kraft cooking



 Aalto University School of Chemical Technology
 [1] Vuorinen, CHEMISTRY OF PULPING AND BLEACHING, https://mycourses.aalto.fi/pluginfile.php/140064/mod_resource/content/1/Supporting%20material%201.pdf
 21.9.2022

 [2] Chakar and Ragauskas, Ind. Crops Prod. 20 (2004) 131-141.
 28.2022

β-O-4 cleavage in acidic conditions





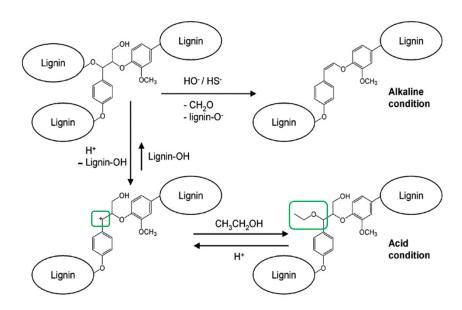
Li et al., Bioresour. Technol. 98 (2007) 3061-3068

Organosolv pulping: ethanol-water process as an example

- Ethylation of the carbocation at Cα reduces formation of condensed linkages
- This is one reason why organosolv lignins have low Mw, Tg, and dissolve well in organic solvents

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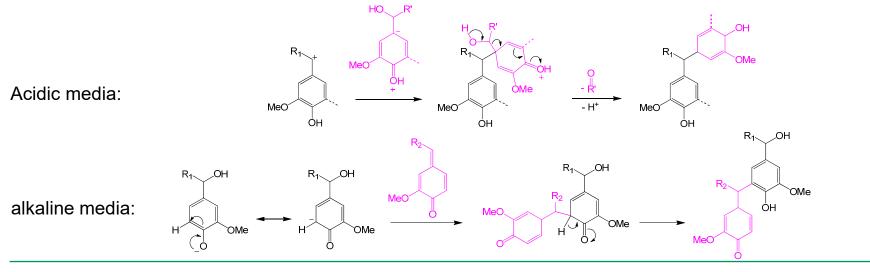


Proposed ethylation of lignin in organosolv pulping^[1]

^[1] Kubo and Kaddla, Macromolecules 2004, 37, 6904-6911

Formation of chemically stable "condensed" C–C linkages

- Generally, condensed linkages reduce solubility and increase molecular weight of lignin
- Occurs either in acidic or alkaline media, for example:





Gierer, Wood Sci. Tecnol. 19 (1985) 289-312

Isolation



Isolation and purification of lignin

Industrial lignin production

- Lignosulfonates (~1100 kt per year)
 - Borregaard LignoTech (Norway) is the main producer
- Kraft (sulfate) lignin (~100 kt per year)
 - WestRock (formerly MeadWestvaco) (US)
 - 2013: Domtar, North Carolina (US)
 - 2015: Stora Enso, Sunila (Finland)
 - 2016: West Fraser LignoForce 30 t/day demonstration plant



Stora Enso's Sunila mill http://www.hs.fi/webkuva/taysi/700/1374823934135?ts=763



Isolation of lignin from spent pulping liquors

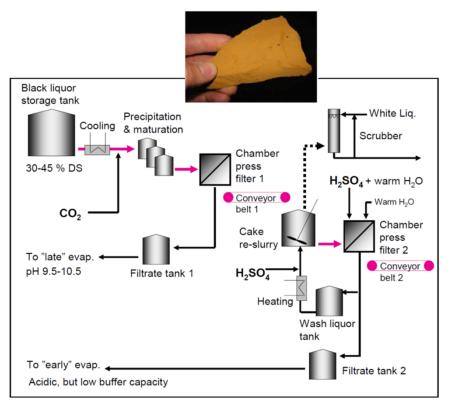
- Acid precipitation from alkaline solution: recall that lignin contains acidic groups which ionize above pH 4 and pH 7–10: precipitation occurs when anionic charge is reduced so that aggregation occurs
- Complexation with metals or cationic polymers
- Anti-solvent precipitation: lignin soluble in organic solvent (ethanol) precipitates when mixed with a miscible fluid (water)

- Membrane technology using small pore sizes ~1 kDa



Commercial lignin isolation processes

- Based on the capacity of CO₂ to reduce pH and precipitate lignin at pH 10
 - Carbon dioxide precipitation of lignin from pine kraft black liquor^[1]
- Lignoboost technology was further developed by Innventia and acquired by Valmet
- LignoForce technology oxidizes lignin before CO₂ precipitation^[2]



Tomani, Cellulose Chem. Technol., 44 (1-3), 53-58 (2010)

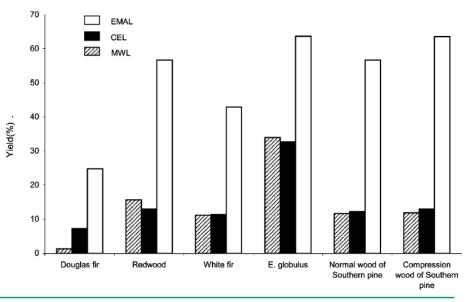


[1] Alén et al., Tappi 62: 11 (1979) 108-110.[2] https://www.lemaitrepapetier.ca/images/stories/documents/articles%20techniques/J-FOR_Vol2-issue4-ART1-The%20Lignoforce%20System.pdf35

Analytical lignin isolation methods

- Isolation of lignin from wood for characterization or as a model material causes always some structural alteration
- In the Milled Wood Lignin (MWL) method, wood is ground in a ball mill and subsequently extracted with aqueous Dioxane
- Cellulolytic enzyme lignin (CEL) and Enzymatic Mild Acidolysis Lignin (EMAL) procedures remove polysaccharides using hydrolytic enzymes before extraction of lignin with (acidic) aq. dioxane





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Guerra et al., J. Agric. Food Chem., 2006, 54, 5939–5947 Guerra et al., J. Agric. Food Chem., 2006, 54, 9696–9705

Lignin derivatizations

- In order to analyze structure of lignin or to render it more suitable to applications, chemical or anzymatic derivatizations are used
- Some common chemical derivatizations are:
 - Acetylation
 - Methylation
 - Carboxymethylation
 - Cationization
 - Cross-linking
- Various other organic compounds such as fatty acids can be grafted to lignin
- The enzyme laccase has been used to oxidize and cross-link lignins



Characterization

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Tools for characterization of lignin

- Combination of wet chemistry with spectroscopy, chromatography, and various other techniques
- Understanding the organic chemistry in use may help developing possible routes for chemical and materials production from lignin
- Characterization in liquid or solid state?
 - Liquid state: functional groups, molecular weight, interunit linkages Solid state: material properties, functional groups and elemental composition, imaging of the physical structure
- A few illustrative examples follow



Solution-state analysis of lignin

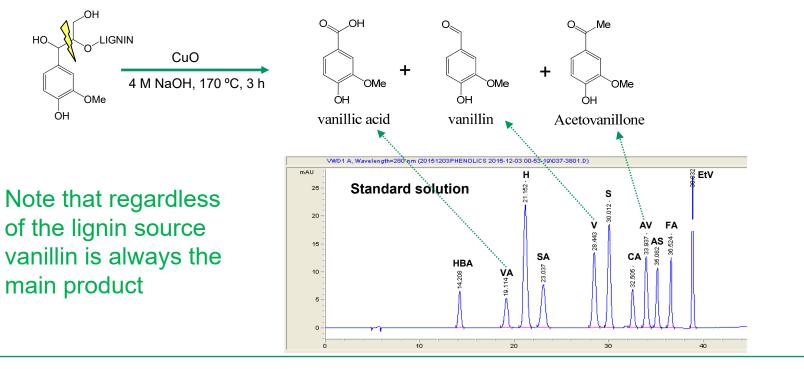
Chemical degradations

- Oxidations: ozonolysis, permanganate oxidation, nitrobenzene oxidation, **cupric oxide oxidation**
- Reductive treatments: **thioacidolysis**, DFRC
- Chromatography is used to quantify the products
- NMR spectroscopy
- Size-exclusion chromatography



Cupric oxide oxidation

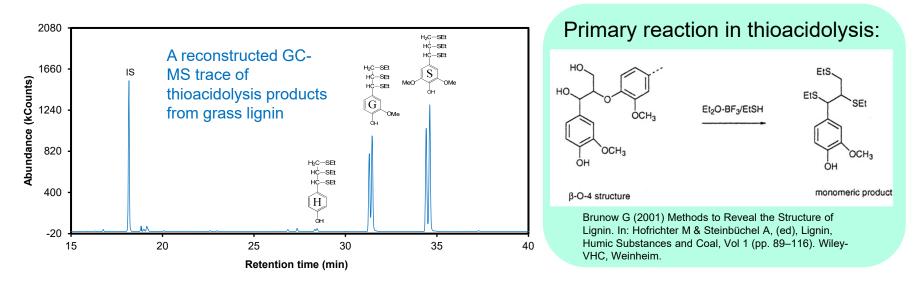
- A classical oxidative degradation method still in use today due to its simplicity
- Reaction mechanism involves radical oxidation





Thioacidolysis

- Comparable information as from CuO oxidation, but more specific to b-O-4 cleavage and yields less degradation side-products
- Solvolytic cleavage of lignin in the presence of ethanethiol
- Malodorous (EtSH) and harmful (Et₂O-BF₃) reagents limit applicability



Aalto University School of Chemical Technology Sipponen, Doctoral thesis 2015

¹³C NMR

- Nuclear Magnetic Resonance (NMR) spectroscopy measures chemical shifts (changes in frequency compared to the reference frequency) when the atom is subjected to a magnetic pulse at similar frequency as its natural resonance frequency
- ¹³C NMR delivers a wealth of quantitative structural information:^[1]
- However, time consuming in data collection and analysis

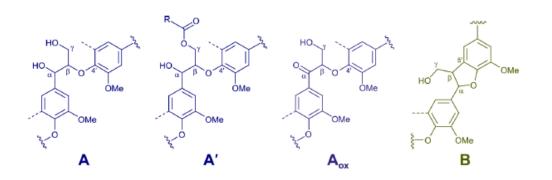
/	β-O-4 total	
	Pino/syringylresinol $(F_{\alpha})^{b}$ Phenylcoumarane (E_{α}) Sugars (C_{1}) $S_{2,6}$ G_{2} H_{4} Degree of condensation OMe	Primary aliphatic OH Secondary aliphatic OH 5-free phenolic OH 5-subst. phenolic OH Ar-H Oxygenated aliphatic Saturated aliphatic
	Non-conjugated CO Conjugated CO Non-conjugated COOR Conjugated COOR Total OH	EtO– Alkyl-O-Alkyl Side chain length Demethylation degree

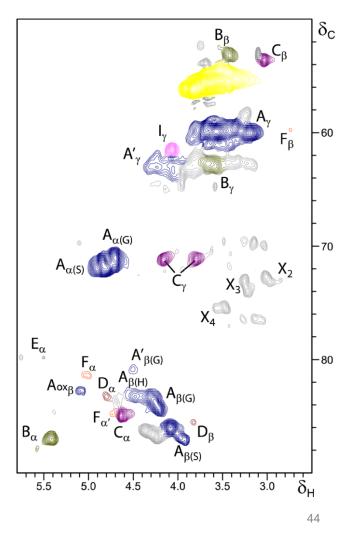


^[1] Balakshin and Capanema, RSC Adv., 2015, 5, 87187-87199

HSQC NMR (2D NMR)

 Integration of ¹H and ¹³C cross peaks gives semiquantitative information of the lignin interunit linkages as well as linkages to carbohydrates



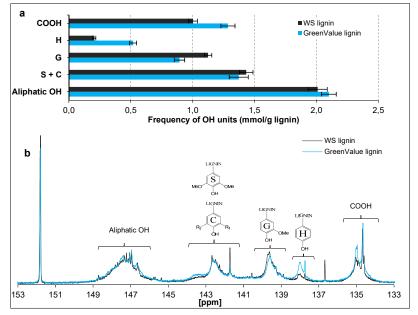




Del Río et al., J. Agric. Food Chem. 60 (2012) 5922-5935.

³¹P NMR, a powerful method for OH groups

• Derivatization of lignin in solution with a P-donor which is quantitatively measured against internal standard



TMDP = 2-Chloro-4,4,5,5-tetramethyl-1,3,2-dioxaphospholane **IS** = N-Hydroxy-5-norbornene-2,3-dicarboxylic acid imide

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LIGNIN IS SUBUNIT OMe ÓН ÓН TMDP `Me Me `Me Mé Ме Mé Ме `OMe `Me Me Me Me Mė Ме Mé Мe

Sipponen, Doctoral thesis 2015

Size exclusion chromatography

- Separation of molecules based on their hydrodynamic radii (size in solution)
- Organic and aqueous elution systems are in use
- Sample must dissolve fully and should not associate with the particles in the column packing material
- Molecular weight is calculated relative to known Mw standards or directly using laser scattering or viscometer



Analysis of lignin as a product

- Complex structure, but all-inclusive analysis is not necessary
- Key properties in relation to the application important
- Real application tests needed also
- Distinction of a high purity lignin from a low purity one:
 - Low amounts of contaminating carbohydrates, sulphur, inorganics
 - Good solubility and material properties



Applications

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Applications of lignin

- 2.10¹⁰ tons of lignin is produced annually in Nature [1]
- Only a fraction of the total is used in products, why?
 - Lack of regular (sterochemical) structure
 - Heterogeneous repeating units (H, G, S and their linkage patterns)
 - Poor understanding of how the degree of polymerization and structure change when lignin is isolated from cell walls
- However, recent advances in lignin isolation and characterization hold promise for increased use of lignin in materials



^[1] Funaoka, Macromol. Symp. 2003, 201, 213–221

R&D for commercialization of lignin is undergoing rapid growth

storaenso

New kind of gold from Nordic forests

Lignin is an organic material that binds the fibres and cells of wood. The word lignin may not be familiar to many, but there is a lot of anticipation and excitement around this material. Why?

PHOTO: Vesa Laitinen

excitement at the mill.

The first dedicated biorefinery investment is expected to be completed in early 2015. Mill Director OIII-Pekka Reunanen is pleased, and with good reason. Installing the new machinery in the architectural milieu that dates back to 1938 is an enjoyable challenge for the mill staff. "We won't be building new buildings; instead, the idle buildings in the mill area will be used to house the machinery required for the blorefinery," explains Reunanen.

F xperts say that lignin may eventually be

one of the new renewable materials

extracted out of the green gold of the Nordic

Finland will, in the future, extract lignin from

announcement made in July, there is an air of

forests. Stora Enso's Sunila Pulp Mill in

Following the biorefinery investment

pine and spruce.

Prior to making the 32-million-euro investment decision, Stora Enso carefully looked into areas where lignin extraction would be profitable. Sunila is Stora Enso's first, but probably not its last, biorefinery dedicated investment.

Stora Enso has been researching lignin and its attributes for more than a decade. "Lignin is a challenging and complex material; it's an honour to be a pioneer in this area. At the same time, we are taking a leap into new and previously uncharted markets, but Stora Enso is in a good position to develop commercially viable applications from lignin," says **Mikael Hannus** of the Biomaterials's Biorefinery and Bioenergy unit.

The initial markets are anticipated in, for example, the construction and automotive industries, where lignin offers a sustainable alternative to the phenois used in plywood and wood-panelling glues and the polyois used in foams. Other applications are also under development.





http://www.storaenso.com/rethink/new-kind-of-gold-from-nordic-forests http://www.upmbiochemicals.com/lignin/Pages/Default.aspx

Current uses of lignin

- Energy (it has value!) 26.7 MJ/kg (HHV)
- Vanillin, 3% from softwood (~10% of lignin)
 - Today, 20% of vanillin is produced from lignin and 80% from crude oil using the guaiacol route
- Pyrolysis of lignin \rightarrow bio-oil (15–25%), gas (20–50%), char (30–60%)
- Aromatic chemicals (BTX)
- Composites and polymer blends
- Micro- and nano-scale structures (particles, capsules, sheets etc.) for novel applications
- High-performance materials such as carbon fibres are also under development

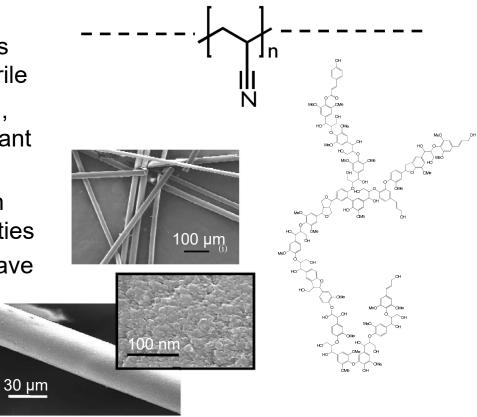


Carbon fibers from lignin

- Carbon fibers are high-value materials currently produced from polyacrylonitrile
- Lignin is more affordable raw material, but less linear polymer with predominant ether linkages
- → Challenges the production of carbon fibres with sufficient strength properties
- → However, this is promising results have been obtained in this research area

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Kadla et al., Carbon 40 (2002) 2913-2920

Summary of keywords

	aliphatic	umenzation		
condensation	phenolic	oxidation / reduction	radical coupling	
quinone methide intermediate	methoxyl	polydisperse branched	condensed / uncondensed units	
monolignols	β-Ο-4	hetero	ogeneous	
aror	aromatic lignin		anionic henyl	
lignosulfonate		p njenovjp	•	
kraft lig	nin	guaiacyl	H/G/S	
	1 mar	sy	ringyl	
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dimerization