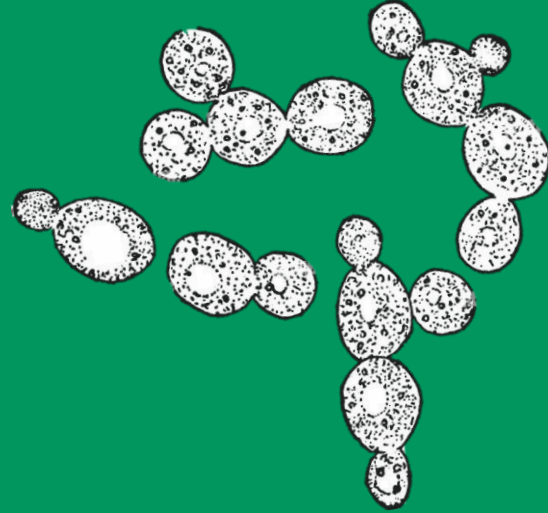


Synthetic Yeast 2.0

Group 13



Aalto University
School of Chemical
Engineering

Dinara Bozzhigitova

Oguzcan Ates

Hanna Dahl

Noora Karvonen

Submitted on 25.3.2023

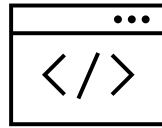
Outline



Yeast 2.0



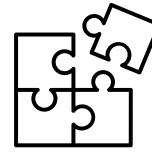
Design



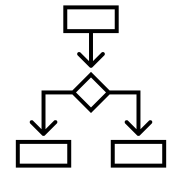
Programs



Wetlab



SCRaMbLE



Further
development

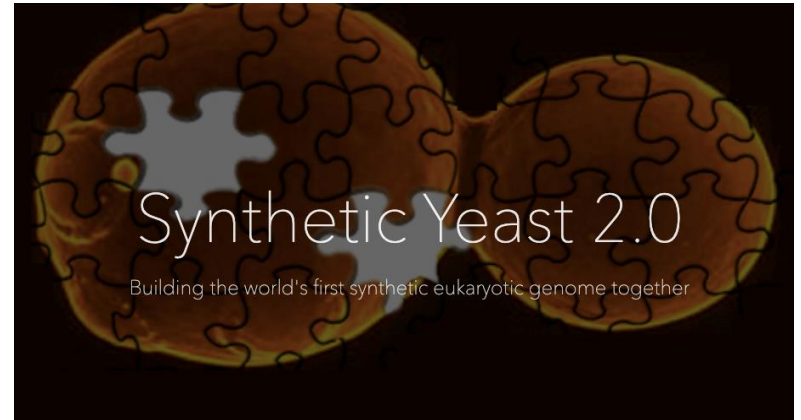
What is Yeast 2.0?

Synthetic Yeast 2.0

- The aim of project is to reconstruct a fully synthetic *Saccharomyces cerevisiae* yeast chromosome
- *S. cerevisiae* was chosen for the project because it is the most researched and understood genome
- 7 out of 16 chromosomes were successfully synthesised

By learning how to synthesize the genome, we can:

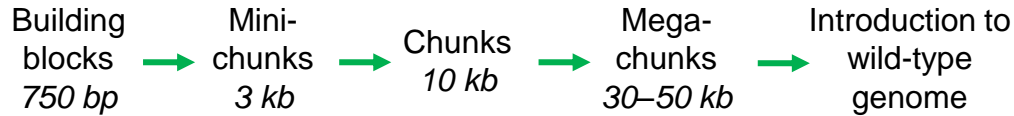
- **Answer** the fundamental questions about chromosomes properties
- **Develop** more efficient biofuel, medicines and other fermentation-related industries



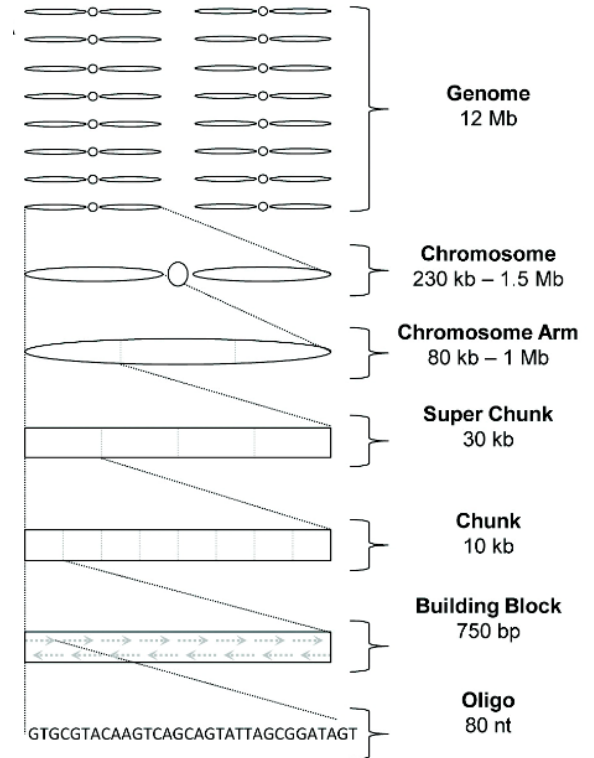
Synthetic Yeast 2.0 logo and slogan
<https://syntheticyeast.github.io>

Synthetic Yeast 2.0

- Changes to native genome are done using “**bottom-up**” **approach**. The alternations are done on a region approximately of 30 kb. This way, it is easier to monitor the viability and fitness of the cell.
- Assembly is done step-wise:



- Usually, 3–6 x 10 kb chunks are introduced at a time



The yeast genome division into increasingly smaller segments

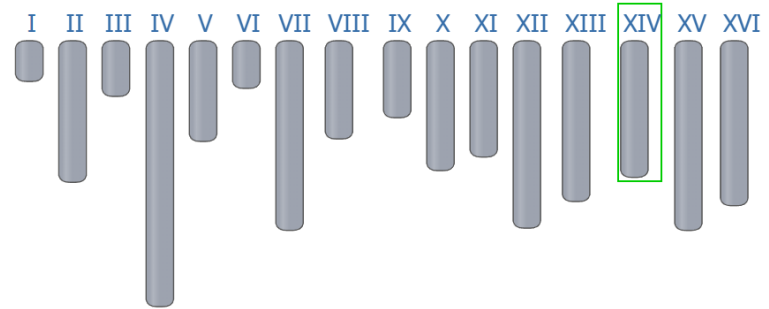
<https://doi.org/10.4161/bbug.19543>

Our design & Modifications

What was chosen and why?

Chromosome XIV, strain S228c

- Contains the **KRE1**
 - Cell wall glycoprotein involved in beta-glucan assembly
 - Serves as a K1 killer toxin membrane receptor
- **Studying the assembly of this gene might help scientists to identify K1 resistant cells, thereby:**
 - Developing more fit and robust cells
 - Use KRE1 for selective pressure for cultivation of transformed cells



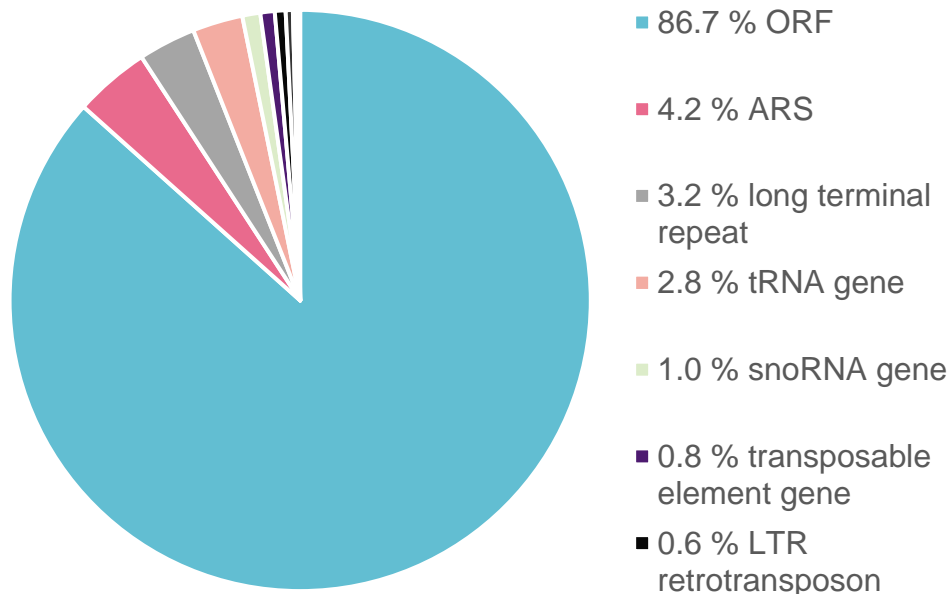
S. cerevisiae S288C chromosomes

https://www.ncbi.nlm.nih.gov/genome/gdv/browser/genome/?chr=XIV&id=GCF_000146045.2

Chromosome XIV

- Not synthesized yet
- Composition

Strain: S288C
Length: 784333 bases
NCBI RefSeq: BK006947.3

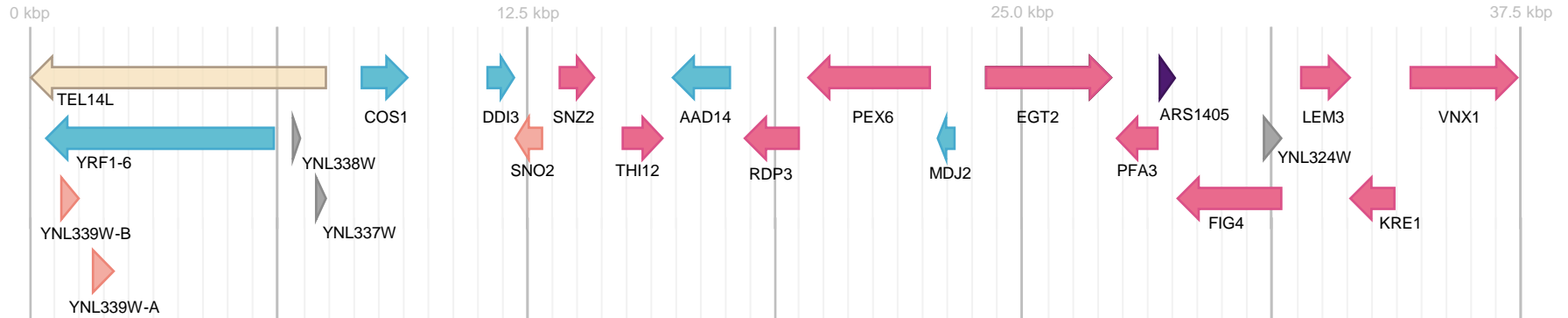


List of genes in the megachunk

<u>TEL14L-</u>	Telomeric region on the left arm of Chromosome XIV
<u>YRF1-6</u>	Helicase encoded by the Y' element of subtelomeric regions
<u>YNL339W-B , YNL339W-A</u>	Proteins of unknown function
<u>YNL338W, YNL337W</u>	Dubious open reading frame; unlikely to encode a functional protein,
<u>COS1</u>	Endosomal protein involved in turnover of plasma membrane proteins
<u>DDI3</u>	Cyanamide hydratase that detoxifies cyanamide
<u>SNO2</u>	Protein of unknown function
<u>SNZ2</u>	Protein involved in thiamine and pyridoxine biosynthesis;
<u>THI12</u>	Protein involved in synthesis of the thiamine precursor HMP
<u>AAD14</u>	Putative aryl-alcohol dehydrogenase;
<u>RPD3</u>	Histone deacetylase,
<u>PEX6</u>	AAA-peroxin;
<u>MDJ2</u>	Constituent of the mitochondrial import motor; associated with the presequence translocase
<u>EGT2</u>	Glycosylphosphatidylinositol (GPI)-anchored cell wall endoglucanase
<u>PFA3</u>	Palmitoyltransferase for Vac8p; required for vacuolar membrane fusion
<u>ARS1405</u>	Autonomously Replicating Sequence
<u>FIG4</u>	Phosphatidylinositol 3,5-bisphosphate (PtdIns[3,5]P) phosphatase; required for efficient mating and response to osmotic shock
<u>YNL324W</u>	Dubious open reading frame
<u>LEM3</u>	Membrane protein of the plasma membrane and ER
<u>KRE1</u>	Cell wall glycoprotein involved in beta-glucan assembly; serves as a K1 killer toxin membrane receptor
<u>VNX1</u>	Calcium/H ⁺ antiporter localized to the endoplasmic reticulum membrane

Original megachunk

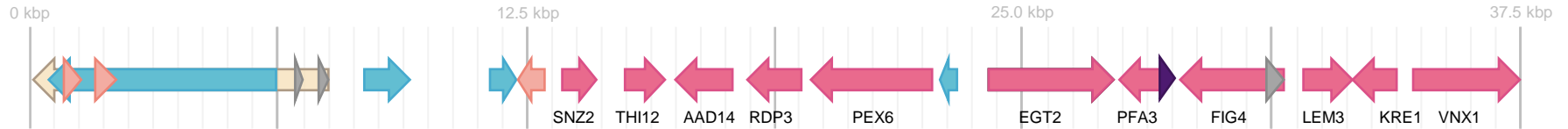
Chromosome **XIV**, chunk between 0–37.5 kbp



- Essential genes
- Non-essential genes
- Autonomously replicating sequence
- Dubious ORFs
- Protein of unknown function
- TEL

Original megachunk

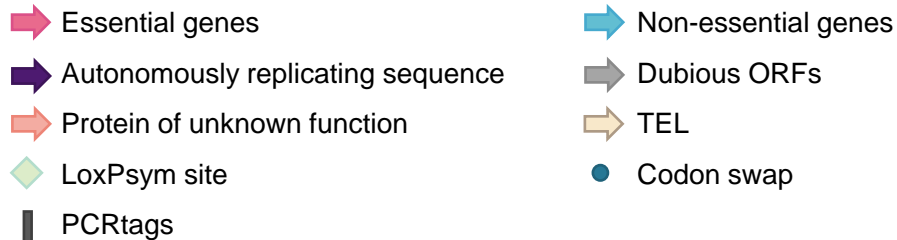
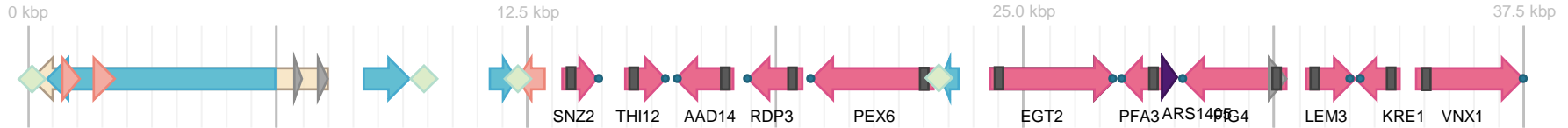
Chromosome XIV, chunk between 0-37.5 kbp



- ➔ Essential genes
- ➔ Non-essential genes
- ➔ Autonomously replicating sequence
- ➔ Dubious ORFs
- ➔ Protein of unknown function
- ➔ TEL

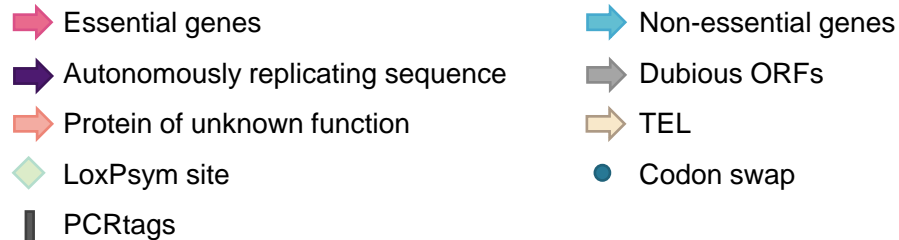
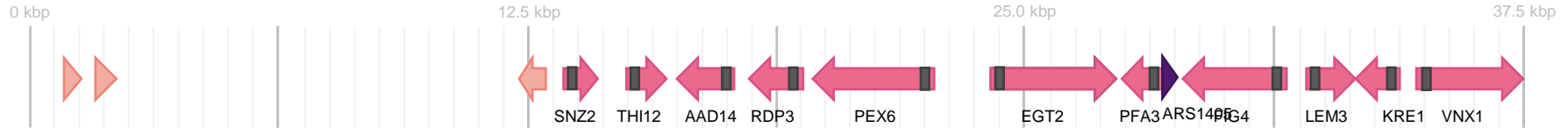
Editing

- **Deletion** of non-essential genes, introns and repeats (LTRs, Sub-TEs and transposons)
- **Re-location** of tRNA genes
- **Addition** of loxP sites
- **Re-coding** stop codons (TAG -> TAA) and including PCR tags



Constructed megachunk

- **Deletion** of non-essential genes, introns and repeats (LTRs, Sub-TEs and transposons)
- **Re-location** of tRNA genes
- **Addition** of loxP sites
- **Re-coding** stop codons (TAG -> TAA) and including PCRtags



Databases & Softwares

Databases & Softwares

Databases

- ***S. cerevisiae* genome database:**
<https://yeastgenome.org/>
 - Includes biological information about *Saccharomyces cerevisiae*
- **Yeast 2.0:**
<https://syntheticyeast.github.io/>
 - To learn about the designs of Sc 2.0

Softwares & Other tools

- **BioStudio:**
<https://metacpan.org/pod/Bio::BioStudio::Git>
 - The archive of genetic codes
- **DNA Atlas:**
<https://www.dnaatlas.com/>
 - To construct sequences and plasmid maps
- **ORF finder:** <https://web.expasy.org/translate/>
 - Translation of nucleotide to a protein sequence

Wet lab procedure

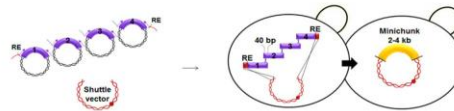
Wet lab procedure

1. Synthesis of building blocks



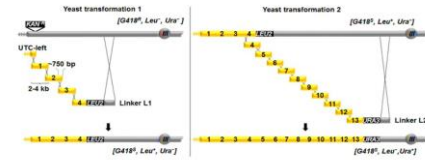
<https://doi.org/10.1126%2Fscience.1249252>

2. Assembly of minichunks



<https://doi.org/10.1126%2Fscience.1249252>

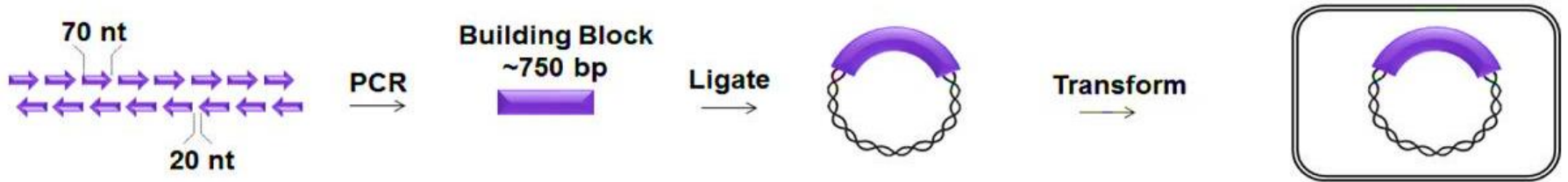
3. Replacement with minichunks



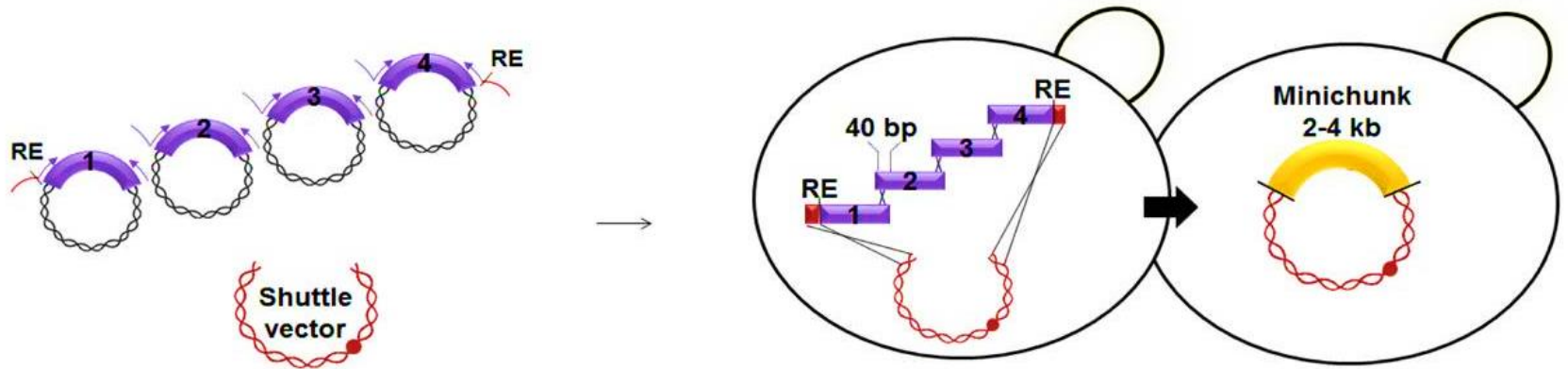
<https://doi.org/10.1126%2Fscience.1249252>

4. Confirmation using PCRtags

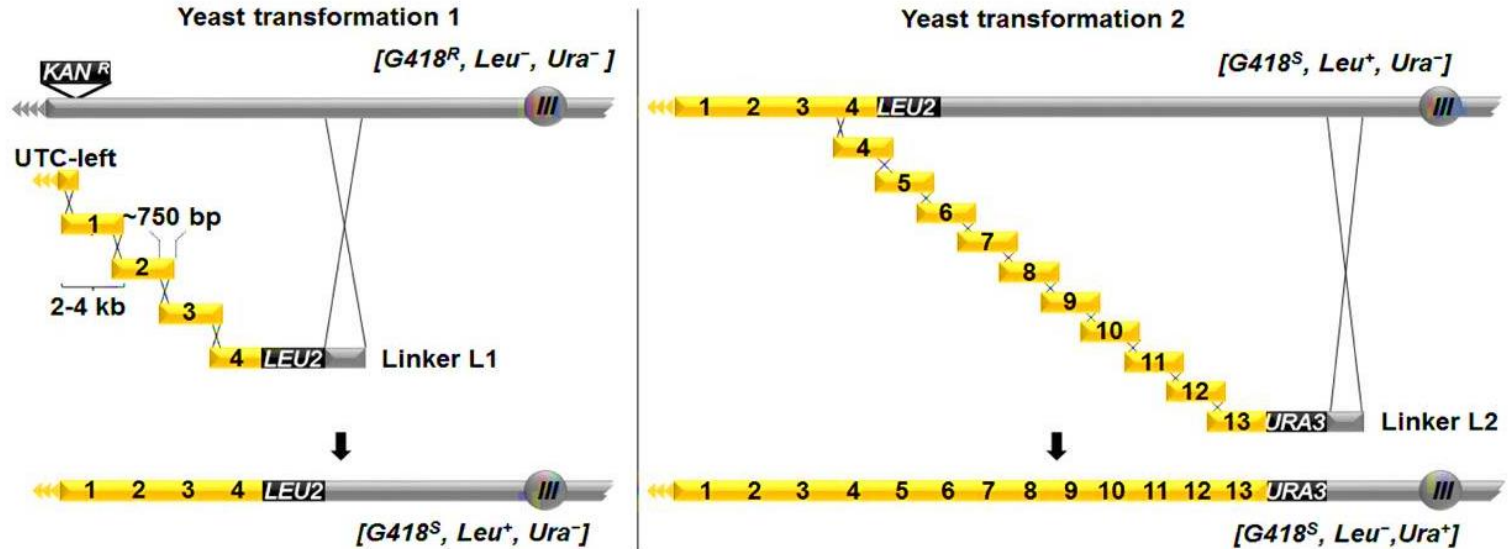
1. Synthesis of building blocks



2. Assembly of minichunks



3. Replacement with minichunks

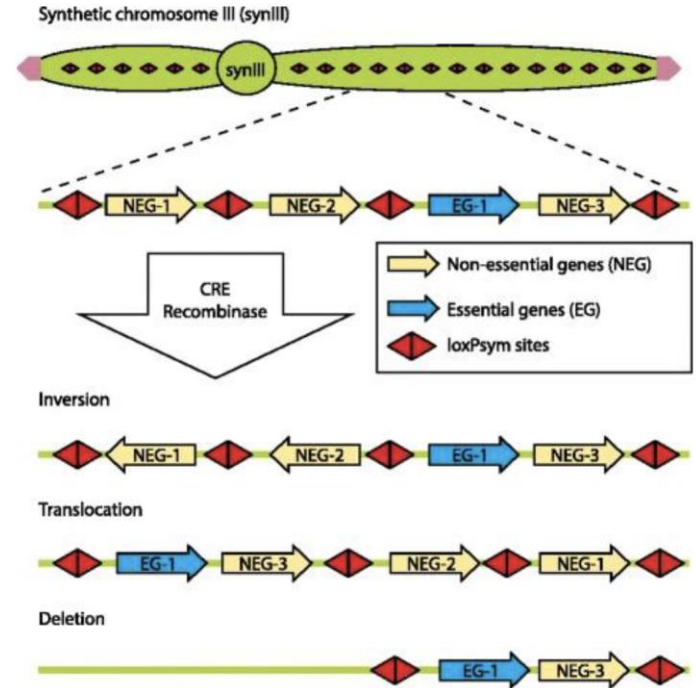


SCRaMbLE

*Synthetic Chromosome Rearrangement and
Modification by LoxP-mediated Evolution*

SCRaMbLE

- **Genetic engineering technique** that allows for the creation of novel genetic variations
- Generates combinatorial genomic diversity through rearrangements at designed recombinase sites
- Synthetic chromosome is designed with multiple **LoxP sites**, specific DNA sequences recognized by Cre recombinase enzyme
- Cre recombinase recognizes and recombines the LoxP sites, results in different combinations of DNA sequences
- Enables a **variety of random mutants in a short time**
- Used in yeast to generate a library of genetically diverse strains with novel phenotypes
- Has potential applications in biotechnology and synthetic biology



The pathway of Scramble (Annalaru et al, 2015).

Further Development

Further developments

Yeasts 3.0

- Research initiative constructing a comprehensive and high-quality genome scale model of yeast *S. cerevisiae*.
- Builds upon the work of the Yeast 2.0 project.

Biochemical and food industry

- The production of synthetic fuels, utilization of biomass for biodiesels
- Food ingredients
- Protein drugs



Thank you!
Questions?

References

- Annaluru, N., Ramalingam, S. & Chandrasegaran, S. 2015. Rewriting the blueprint of life by synthetic genomics and genome engineering. *Genome Biol* 16, 125. DOI:10.1186/s13059-015-0689-y.
- Bussey, H. (1991) "K1 killer toxin, a pore-forming protein from yeast," *Molecular Microbiology*, 5(10), pp. 2339–2343. Available at: <https://doi.org/10.1111/j.1365-2958.1991.tb02079.x>.
- *Chromosome XIV | SGD* (no date). Available at: https://yeastgenome.org/contig/Chromosome_XIV.
- Dai, J., Boeke, J. D., Luo, Z., Jiang, S., & Cai, Y. (2020). *Sc3.0: revamping and minimizing the yeast genome*. *Genome Biology*, 21(1).doi:10.1186/s13059-020-02130-z
- Dymond, J.S. and Boeke, J.D. (2012) "The *Saccharomyces cerevisiae* SCRaMbLE system and genome minimization," *Bioengineered*, 3(3), pp. 170–173. Available at: <https://doi.org/10.4161/bbug.19543>.
- Shen, Y., Stracquadanio, G., Wang, Y., Yang, K., Mitchell, L. A., Xue, Y., ... & Bader, J. S. (2016). SCRaMbLE generates designed combinatorial stochastic diversity in synthetic chromosomes. *Genome research*, 26(1), 36-49.
- *Synthetic Yeast 2.0* (no date). Available at: <https://syntheticyeast.github.io/>.
- *Yeast 2.0 - Bioplatforms* (2020). Available at: <https://bioplatforms.com/projects/synthetic-yeast-2-0/>
- Zhang, Y. *et al.* (2022) "Rapid evolution and mechanism elucidation for efficient cellobiose-utilizing *saccharomyces cerevisiae* through synthetic chromosome rearrangement and modification by loxpsym-mediated evolution," *Bioresource Technology*, 356, p. 127268. Available at: <https://doi.org/10.1016/j.biortech.2022.127268>.

