

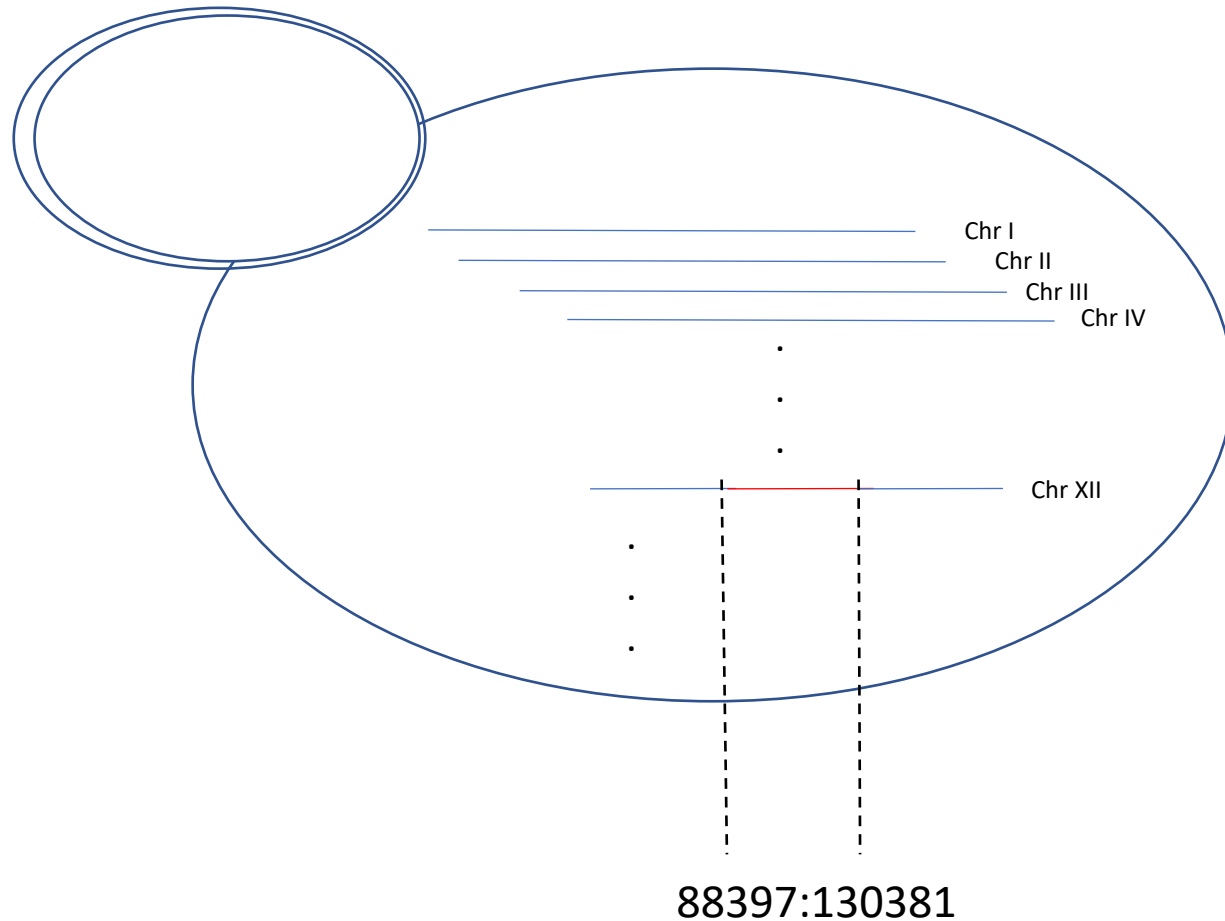
Genetic modification on
Saccharomyces cerevisiae:
Project Sc. 2.0

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Yeast 2.0

- Synthesizing and optimizing the genome of baker's yeast (*Saccharomyces cerevisiae*)
- Aim: to create an "artificial" yeast species that is better suited for biotechnological applications than the natural form
- Removing parts of DNA and adding useful parts to make the genome more efficient
- Potential to facilitate the manufacture of biotechnological products and contribute to the development of new therapies and medicines.
- Can serve as a model to study the complex relationships between genotype and phenotype

Yeast 2.0 modified region



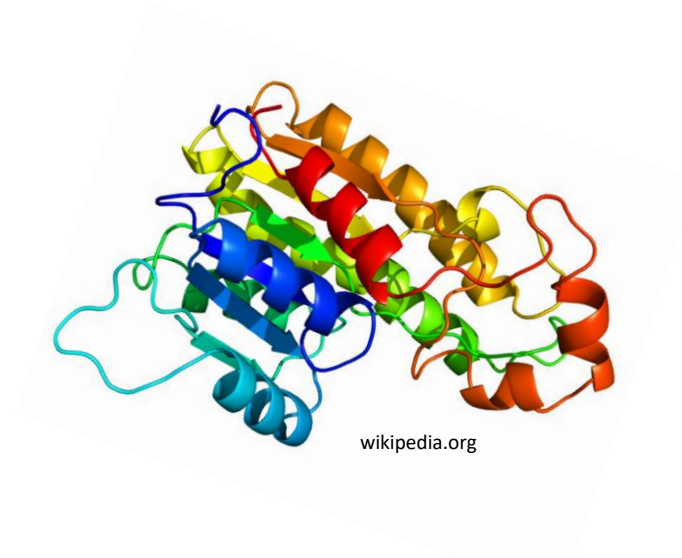
42 kb length region to be synthetically replaced

Contain essential genes for:

- Proteins assembly
- Structural protection

Region content

Chromosome XII - 88397:130381



Heat shock protein (HSP104)

After Heat or Environmental stresses



- Resolubilization
- Refolding
- Dissociation

Of aggregates of damaged proteins

HSP family chaperone (SSA2)

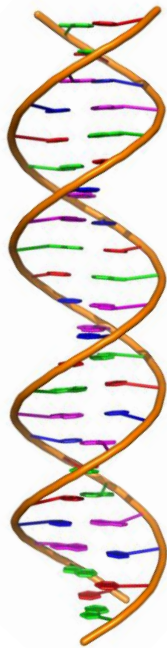
Involved in:



- Protein folding
- Vacuolar import of protein
- Degradation of short-lived proteins

Other genes in the region

Code for proteins



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Unknown function

Cytoskeletal actin organization

Kinases

Mitochondrial

Transmembrane transport

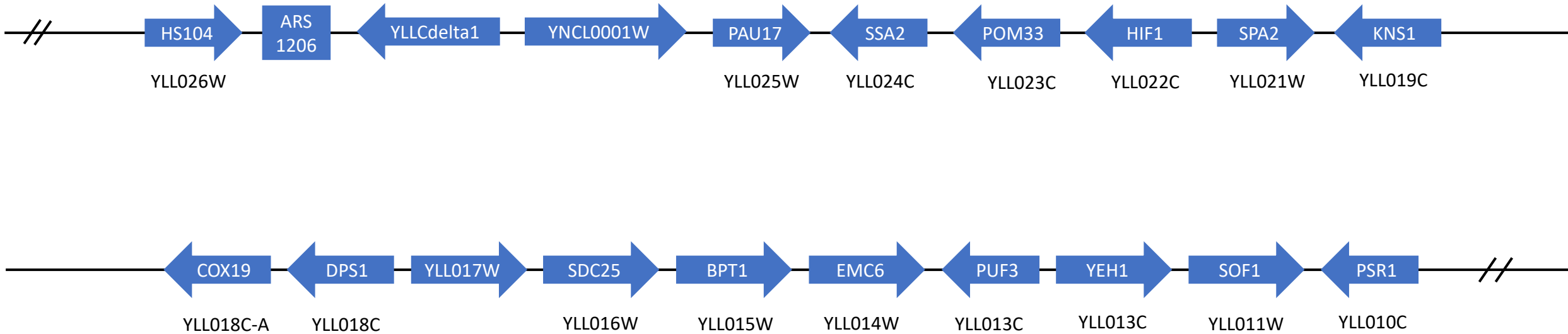
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Non-essential

Targeted gene sequence

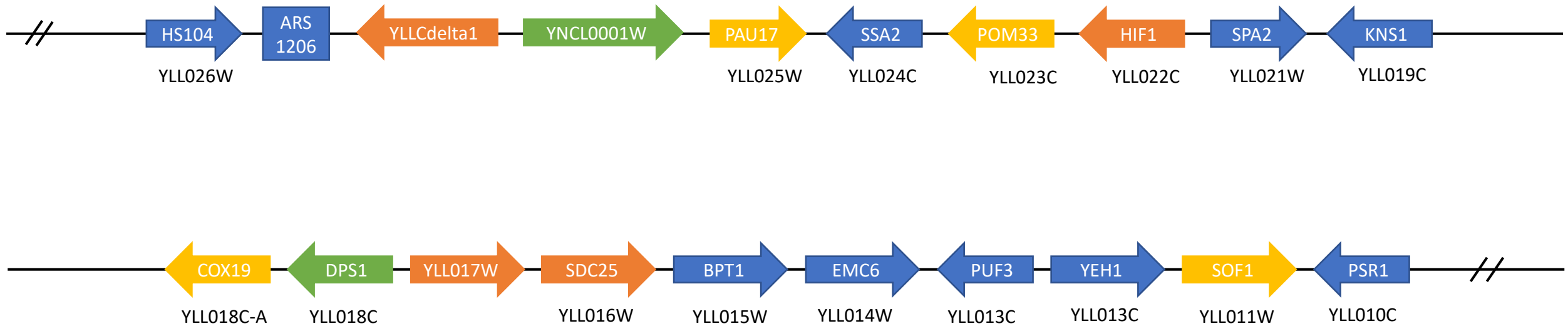


Needed modifications:

- Synthetic telomers
- Removal of introns
- Removal of non essential genes
- Replacement of TAG stop codons
- Deletion of transposons and SPR3 gene
- Relocation of tRNA genes

Targeted gene sequence

No telomers
No introns



- Essential genes
- Non-essential genes
- tRNA genes
- Genes with AUG stop codon

Synthetic megachunk



- Deletion of 3 non-essential genes
- Deletion of 1 transposon
- Removal of 4 TAG stop codons
- Relocation of 2 tRNA genes

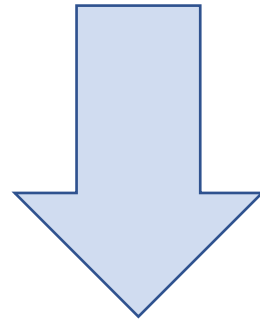
- Essential genes
- Non-essential genes
- tRNA genes
- Genes with AUG stop codon

Relocation of tRNA genes

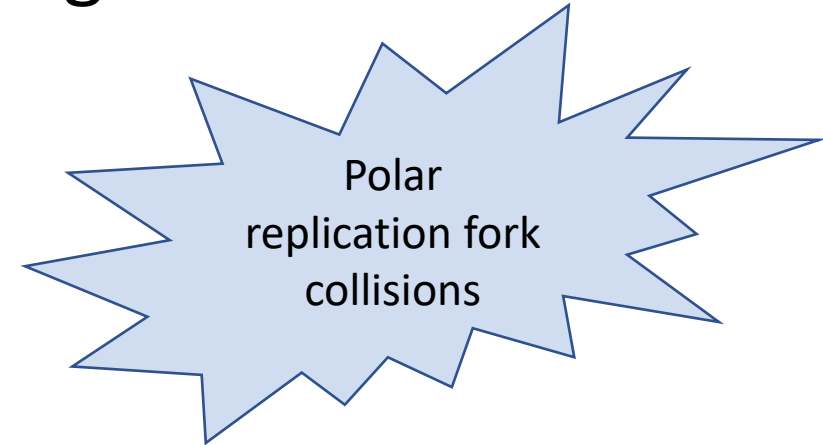
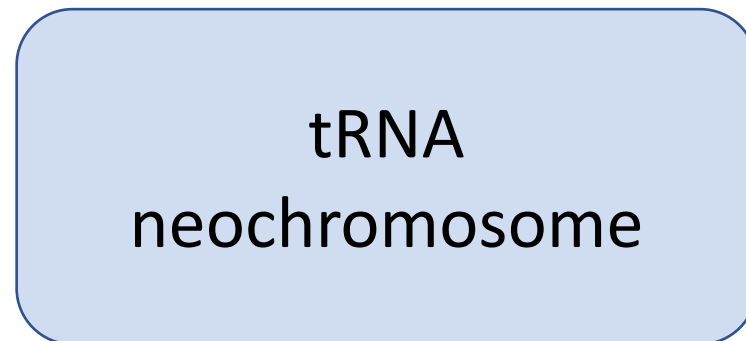


- Rationale:

Constant transcription of tRNA genes leads to more DNA breakage and instability

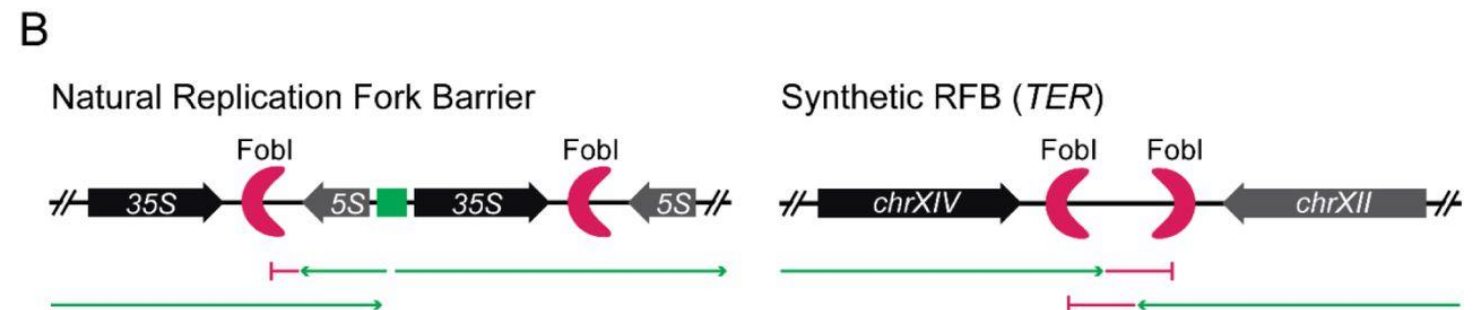
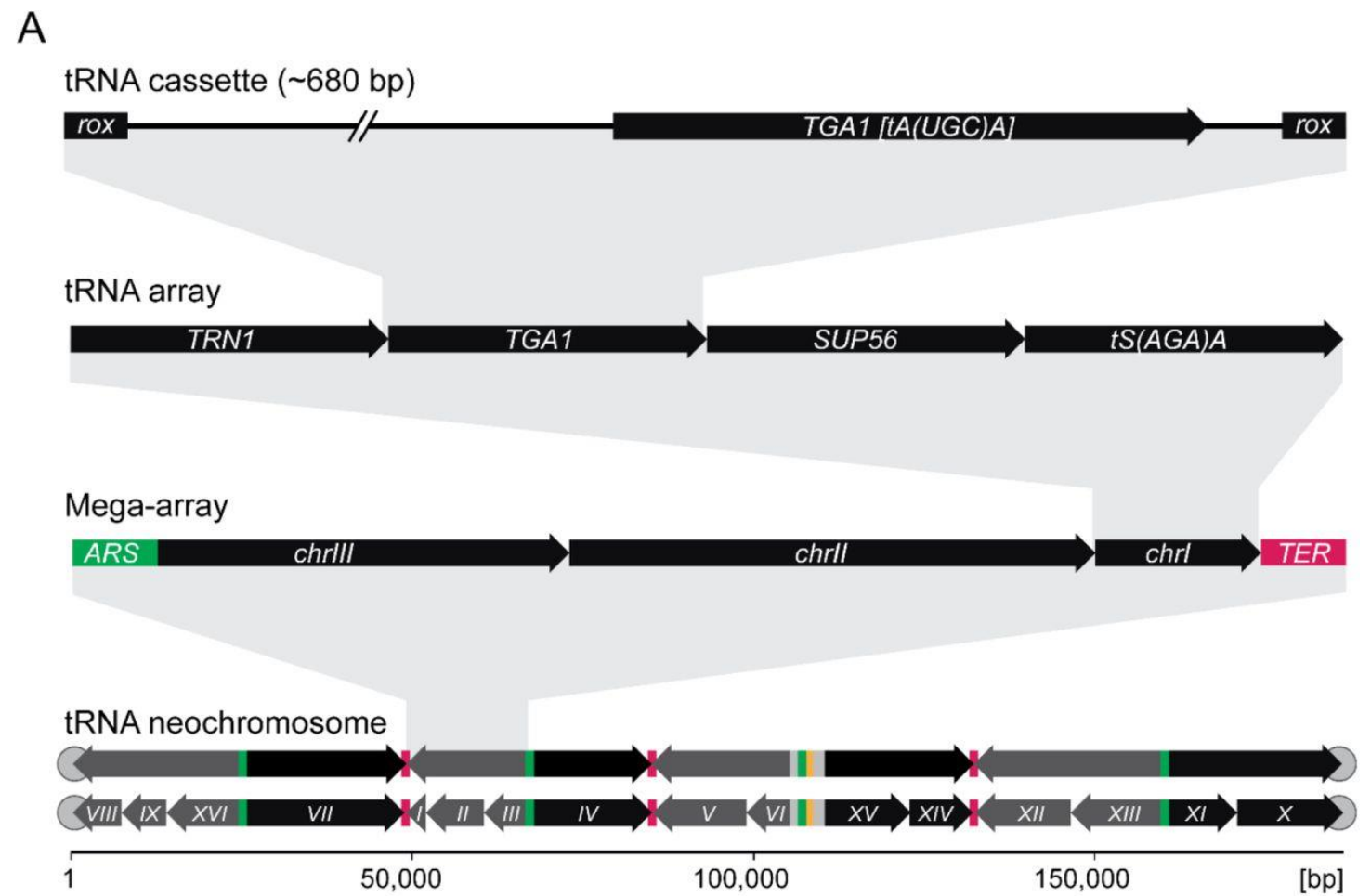


- Solution

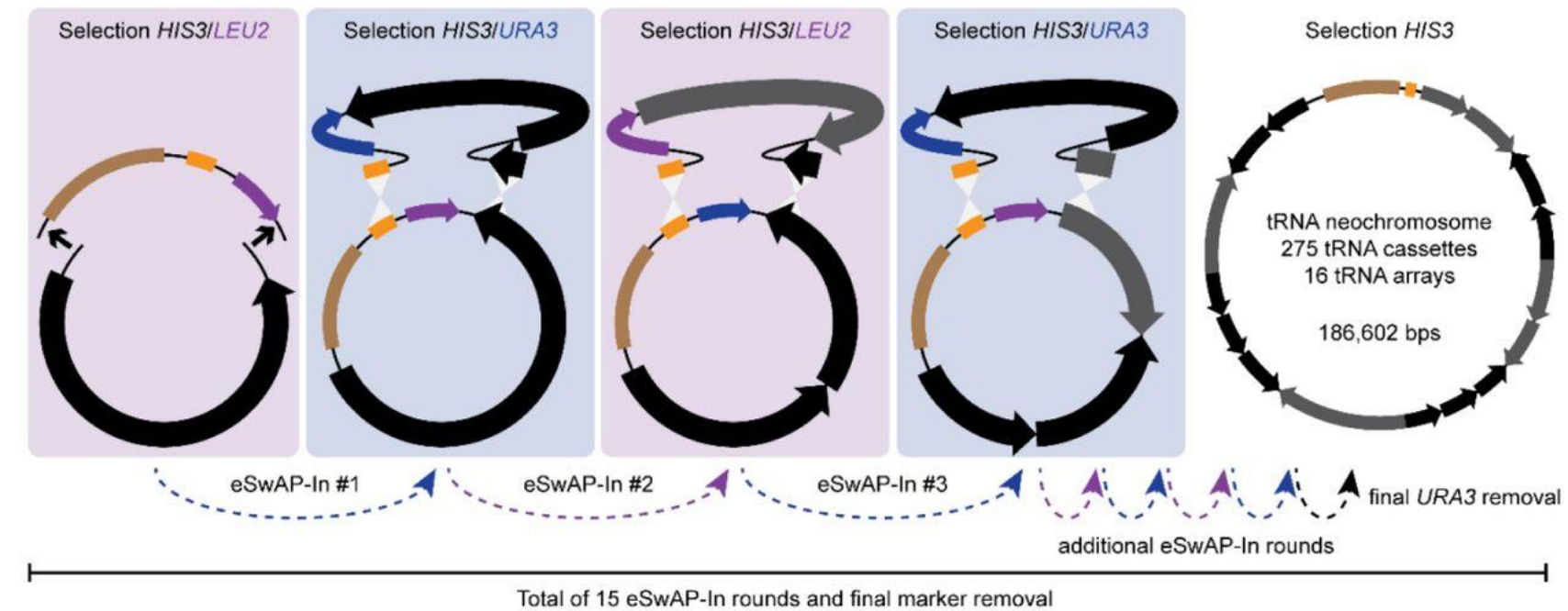


Design of a tRNA neochromosome

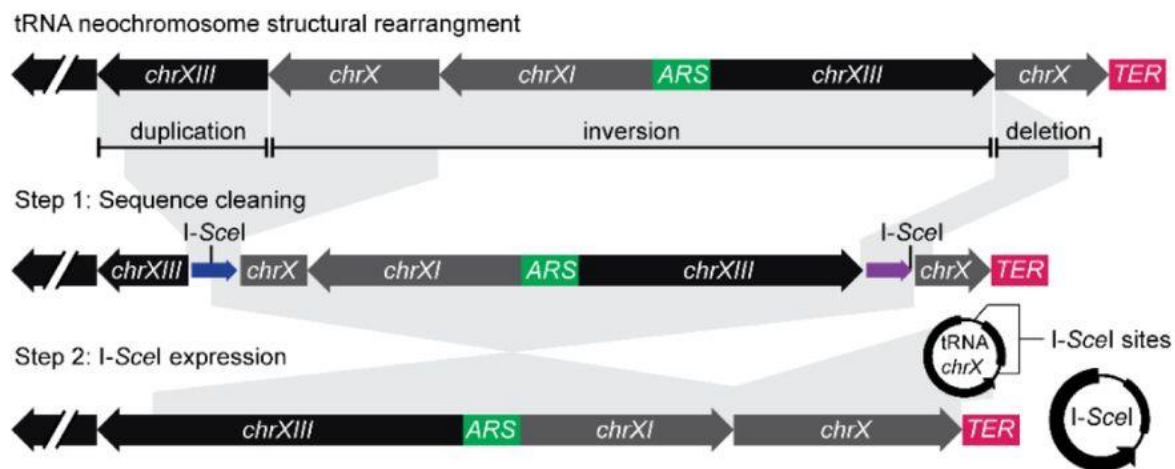
- Orthogonality
- tRNA genes do not face the replication fork



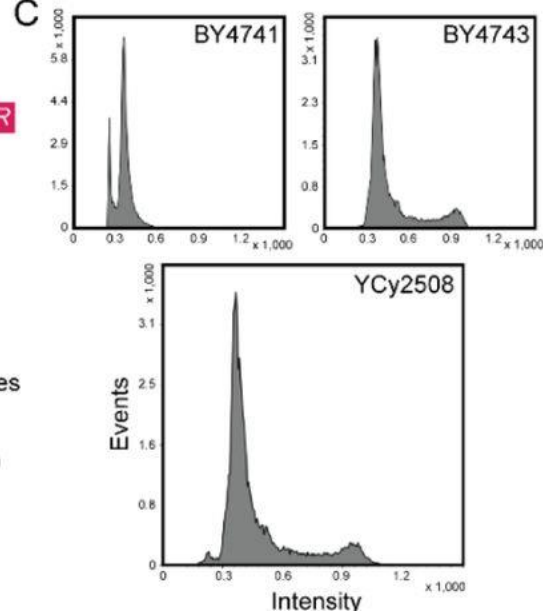
A



B



C



Method

- E-SwAPin Method
- Linearization

PCR Tags

Unique sequence that is added to the ends of the PCR primers

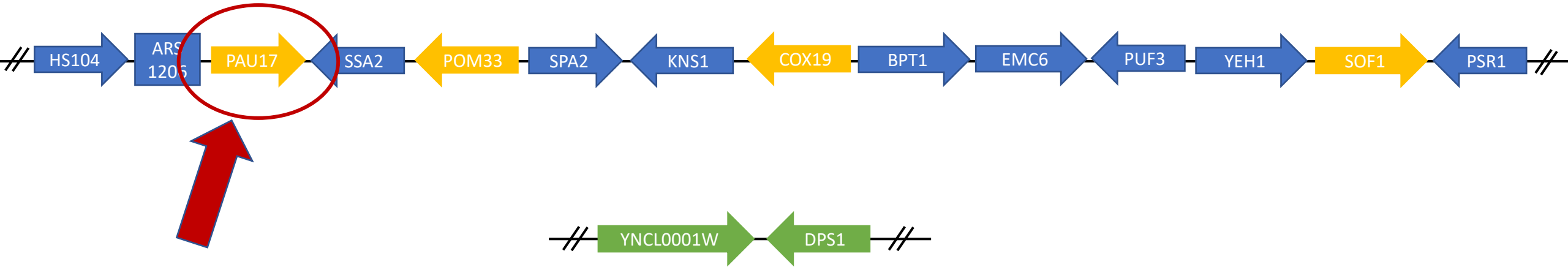
- **Rationale:**

- Allows us to check that the synthetic substitution of the genome has occurred
- Allows us to monitor changes to the genome after SCRaMbLE etc.

- **Considerations:**

- Usually ~60% different from native sequences (minimum 33%) (Ensures they do not interfere with PCR synthesis)
- Melting temperatures between 58 °C and 60 °C

Synthetic megachunk



*Giving examples of primers
for this region*

PCR Primer Sequences for PAU17 / YLL025W

primer	start	len	product size	tm	seq
primer-left-0	356	22	596	59.03	ACACTGCTGTACCAAAGTAGGA
primer-right-0	951	21	596	59.10	GTTGCCAACCCAATCATGTCT
primer-left-1	436	22	569	57.01	TGGTTTCGACGGTATAGATGAA
primer-right-1	1004	20	569	58.86	ACACCACTGCTACCAAGGAA
primer-left-2	369	21	581	57.57	AAACTAGGATTTGACGCCCTT
primer-right-2	949	22	581	58.23	TGCCAACCCAATCATGTCTAAA
primer-left-3	333	20	670	59.05	CGCTCTATCTGCAGACGGTA
primer-right-3	1002	21	670	58.95	ACCACTGCTACCAAGGAAGAA
primer-left-4	323	20	694	58.74	CCATCTCCAGCGCTCTATCT
primer-right-4	1016	20	694	57.74	GGTTAGACAGCAACACCACT

Computer tools

SGD

Saccharomyces cerevisiae whole genome

Benchling

Molecular biology experiments design (primers, plasmids, synthetic sequence...)

NCBI gene
bank

Sequence, information about a gene of interest

UNIPROT

Sequence, information about a protein of interest

GenScript

DNA primer design

Thank you!

Are there any questions?

