CS-E5875 High-Throughput Bioinformatics Machine learning for scRNA-seq analysis

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Contents

- Neural networks: basics
- Cell type identification
- Variational autoencoder
- Single-cell variational autoencoder

Generalized linear model for binary-valued data

- ▶ Recall again the generalized linear modeling (GLM) framework
- Consider data D = {(x₁, y₁),..., (x_n, y_n)), where x_i = (x_{i1},..., x_{ik})^T denotes the explanatory variables and the response variable y_i can have only two possible value: {0,1}
- Binary data can be modeled using the Bernoulli probability density function:

$$\operatorname{Ber}(y \mid p) = p^{y}(1-p)^{1-y},$$

where p is the probability of success, or the mean (parameter) as $\mathbb{E}(Y) = p$

Generalized linear model for binary-valued data

- Recall again the generalized linear modeling (GLM) framework
- ► Consider data D = {(x₁, y₁),..., (x_n, y_n)), where x_i = (x_{i1},..., x_{ik})^T denotes the explanatory variables and the response variable y_i can have only two possible value: {0,1}
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- \blacktriangleright In GLM framework we want to model p using a linear model via a link function
- For binary data the following link function is useful

$$\log\left(\frac{p_i}{1-p_i}\right) = \mathbf{x}_i^T \boldsymbol{\beta} + \beta_0 \quad \iff p_i = \frac{1}{1+\exp(-(\mathbf{x}_i^T \boldsymbol{\beta} + \beta_0))} = \operatorname{sigm}(\mathbf{x}_i^T \boldsymbol{\beta} + \beta_0),$$

where ${\rm sigm}:\mathbb{R}\to[0,1]$ is the sigmoidal function that maps the real line to the interval between 0 and 1

• Therefore, our model is $Ber(y_i | sigm(\mathbf{x}_i^T \boldsymbol{\beta} + \beta_0))$

Logistic regression

- Machine learning terminology:
 - The sigmoidal function is called the activation function and denoted here as $\phi(\cdot)$
 - The GLM model for binary data is called the logistic regression model or linear classifier and is denoted as y = f(x) = φ(x^Tβ + β₀) = φ(β^Tx + β₀)
- Illustrations of the sigmoidal activation function (left) and linear classifiers for two covariates (right)



Neural networks: Perceptron

- Some machine learning / neural network models are inspired by neuroscience and can be seen as models which try to mimic information processing in brain
- ▶ The first neural network model by Rosenblatt was called perceptron
- Perceptron is essentially the logistic regression model where the activation function is the step function

$$y = f(\mathbf{x}) = \operatorname{sign}(\boldsymbol{\beta}^T \mathbf{x} + \beta_0),$$

where sign(x) = 0 if x < 0, and sign(x) = 1 if $x \ge 0$

- Term perceptron is nowadays used to denote the logistic regression model with an activation function that is typically something else than the step function
- Linear classifier and perceptron are limited in that they can only solve linear classification problems (where two classes are linearly separable)

Multilayer perceptron

- Multilayer perceptron (MLP) is the most basic type of deep neural network model
- MLP combines several linear classifiers (perceptrons) such that outputs of the perceptrons in the previous layer are used as the inputs to the linear classifier in the next layer
- Each node in the network implements the function

 $y = f(\mathbf{x}) = \phi(\mathbf{w}^T \mathbf{x} + b),$

where \mathbf{w} and b are the linear model weights that are different for each node



Figure from (Murphy, 2020)

Multilayer perceptron

- The nodes at the bottom correspond to the input x
- ▶ \mathbf{h}_1 denotes the outputs of the perceptrons in the first layer $(\mathbf{W}_1 = (\mathbf{w}_{11}, \dots, \mathbf{w}_{1m}))$

$$\begin{split} \mathbf{h}_1 &= (\phi(\mathbf{w}_{11}^{\mathsf{T}}\mathbf{x} + b_{11}), \dots, \phi(\mathbf{w}_{1m}^{\mathsf{T}}\mathbf{x} + b_{1m}))^{\mathsf{T}} \\ &= \phi(\mathbf{W}_1^{\mathsf{T}}\mathbf{x} + \mathbf{b}_1), \end{split}$$

▶ \mathbf{h}_2 denotes the outputs of the perceptrons in the 2nd layer ($\mathbf{W}_2 = (\mathbf{w}_{21}, \dots, \mathbf{w}_{2m})$)

$$\begin{aligned} \mathbf{h}_2 &= (\phi(\mathbf{w}_{21}^T \mathbf{h}_1 + b_{21}), \dots, \phi(\mathbf{w}_{2m}^T \mathbf{h}_1 + b_{2m}))^T \\ &= \phi(\mathbf{W}_2^T \mathbf{h}_1 + \mathbf{b}_2) \end{aligned}$$

• Output is
$$y = \psi(\mathbf{w}_3^T \mathbf{h}_2 + b_3)$$



Figure from (Murphy, 2020)

Multilayer perceptron

 The MLP model can be written more compactly as

 $y = f_3(\mathbf{f}_2(\mathbf{f}_1(\mathbf{x}; \boldsymbol{\theta}_1); \boldsymbol{\theta}_2); \boldsymbol{\theta}_3),$

where $\mathbf{f}_1(\cdot; \boldsymbol{\theta}_1)$, $\mathbf{f}_2(\cdot; \boldsymbol{\theta}_2)$ and $f_3(\cdot; \boldsymbol{\theta}_3)$ are the perceptrons from the previous page (that have outputs \mathbf{h}_1 , \mathbf{h}_2 , y) and $\boldsymbol{\theta}_i = (\mathbf{W}_i, \mathbf{b}_i)$, $\boldsymbol{\theta}_i = (\mathbf{W}_i, \mathbf{b}_i)$, and $\boldsymbol{\theta}_3 = (\mathbf{w}_3, b_3)$

Alternatively write

$$y = f(\mathbf{x}; \boldsymbol{ heta})$$

where $\boldsymbol{ heta}=(\boldsymbol{ heta}_1, \boldsymbol{ heta}_2, \boldsymbol{ heta}_3)$



Figure from (Murphy, 2020)

Multilayer perceptron:

- Although our example MLP has two hidden layers, in general MLPs can have any number of layers
- Each layer can have an arbitrary number of nodes (=width)
- MLPs can use different types of activation functions



Figure from (Murphy, 2020)

Multilayer perceptron: likelihood and inference

▶ Binary classification: choose the activation function for the last layer to be e.g. the sigmoidal function $\psi(\cdot) = \text{sigm}(\cdot)$ and use the Bernoulli likelihood for the data D

$$\mathcal{L}(\boldsymbol{ heta}) = p(D \mid \boldsymbol{ heta}) = \prod_{i=1}^{n} \mathrm{Ber}(y_i \mid f(\mathbf{x}_i; \boldsymbol{ heta}))$$

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Regression assuming additive Gaussian noise: the activation function for the last layer can be the identity function and use the Gaussian likelihood

$$\mathcal{L}(\boldsymbol{ heta}) = p(D \mid \boldsymbol{ heta}) = \prod_{i=1}^{n} \mathcal{N}(y_i \mid f(\mathbf{x}_i; \boldsymbol{ heta}), \sigma^2)$$

Multilayer perceptron: likelihood and inference

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$$\mathcal{L}(oldsymbol{ heta}) = p(D \mid oldsymbol{ heta}) = \prod_{i=1}^n \mathrm{Ber}(y_i \mid f(\mathbf{x}_i; oldsymbol{ heta}))$$

Regression assuming additive Gaussian noise: the activation function for the last layer can be the identity function and use the Gaussian likelihood

$$\mathcal{L}(\boldsymbol{\theta}) = p(D \mid \boldsymbol{\theta}) = \prod_{i=1}^{n} \mathcal{N}(y_i \mid f(\mathbf{x}_i; \boldsymbol{\theta}), \sigma^2)$$

- ▶ The large number of parameters of the model can be chosen by maximizing the likelihood
- No closed form solution but the parameters can be optimized iteratively using numerical optimization methods

$$oldsymbol{ heta}^{(s+1)} := oldsymbol{ heta}^{(s)} + \Delta rac{\partial \log \mathcal{L}}{\partial oldsymbol{ heta}} igg|_{oldsymbol{ heta}^{(s)}}$$

Multilayer perceptron: illustration

► Deep MLP can learn complex functions



Figure by Cagatay Yildiz

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Cell type identification with neural networks

- ► Computational cell type identification is an important step in scRNA-seq analysis
- Several cell type annotation methods rely on unsupervised clustering
- We we will look at a supervised cell type annotation method that uses deep learning method which is trained on a data set of labelled single cells

Automated cell type identification using neural networks (ACTINN)

- ACTINN (Ma and Pellegrini, 2019) is one of the many recently proposed supervised methods for cell type identification that use deep learning methods
- ▶ Labelled single-cell gene expression profiles $D = ((\mathbf{x}_1, y_1), \dots, (\mathbf{x}_n, y_n))$ are collected from different databases
- ▶ The number of cell types is denoted by $k, y_i \in \{1, ..., k\}$
- The method uses only those genes that appear in all the databases
- Genes with the highest 1% and lowest 1% mean expression are ignored
- \blacktriangleright Genes with the highest 1% and lowest 1% standard deviation are ignored
- ▶ The normalized gene expression vector $\mathbf{x}_i \in \mathbb{R}^d$ for d genes in cell i is obtained as

$$\mathbf{x}_i = \log_2 rac{10^4 \cdot (\mathbf{ ilde{x}}_i + \mathbf{1})}{N_i},$$

where $\tilde{\mathbf{x}}_i$ denotes the raw gene expression counts for all *d* genes and N_i is the total gene expression (count) measured for cell *i*

Automated cell type identification using neural networks (ACTINN)

► ACTINN uses the MLP with three hidden layers that have widths 100, 50 and 25

 $y = f_4(\mathbf{f}_3(\mathbf{f}_2(\mathbf{f}_1(\mathbf{x}; \boldsymbol{\theta}_1); \boldsymbol{\theta}_2); \boldsymbol{\theta}_3); \boldsymbol{\theta}_4)$

with Relu activation function $\operatorname{Relu}(h) = \max(0, h)$ for the hidden layers and the softmax activation (or link function) for f_4

softmax(
$$\mathbf{h}$$
) = $\left(\frac{\exp(\mathbf{h}(1))}{\sum_{j=1}^{k} \exp(\mathbf{h}(j))}, \cdots, \frac{\exp(\mathbf{h}(k))}{\sum_{j=1}^{k} \exp(\mathbf{h}(j))}\right)$

that maps the neural network outputs to k probabilities that sum up to one and thus allows modeling k cell types

The model is trained using the multi-class extension of the Bernoulli likelihood (i.e., likelihood for categorical random variable) based loss function

Automated cell type identification using neural networks (ACTINN)



Comparison of cell type Identification methods



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Dimension reduction

- In the previous lecture we discussed about PCA and how it can be used for dimension reduction and to analyze high-dimensional scRNA-seq data
- In this lecture we will look at some more advanced dimension reduction methods that can be presented as generative models

Probabilistic factor analysis

> Probabilistic factor analysis model can be described as the following generative model

$$\begin{split} p(\mathbf{z}) &= \mathcal{N}(\mathbf{z} \mid \mathbf{0}, \mathbf{I}) \\ p(\mathbf{x} \mid \mathbf{z}) &= \mathcal{N}(\mathbf{x} \mid \mathbf{W}\mathbf{z} + \boldsymbol{\mu}, \boldsymbol{\Psi}), \end{split}$$

where $\mathbf{z} \in \mathbb{R}^{L}$ is a low-dimensional latent variable and $\mathbf{x} \in \mathbb{R}^{D}$ denotes observed data

From data generation point of view, the model first samples a latent variable z, and it then samples data vector x given a value for latent z

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• We can marginalize out the latent variable z from p(x, z) = p(x | z)p(z) to get

$$\begin{split} p(\mathbf{x}) &= \int p(\mathbf{x} \mid \mathbf{z}) p(\mathbf{z}) d\mathbf{z} \\ &= \int \mathcal{N}(\mathbf{x} \mid \mathbf{W}\mathbf{z} + \boldsymbol{\mu}, \mathbf{\Psi}) \mathcal{N}(\mathbf{z} \mid \mathbf{0}, \mathbf{I}) d\mathbf{z} \\ &= \mathcal{N}(\mathbf{x} \mid \boldsymbol{\mu}, \mathbf{W}\mathbf{W}^T + \mathbf{\Psi}) \end{split}$$

Probabilistic factor analysis illustration



Figure 20.9: Illustration of the FA generative process, where we have L = 1 latent dimension generating D = 2 observed dimensions; we assume $\Psi = \sigma^2 \mathbf{I}$. The latent factor has value $z \in \mathbb{R}$, sampled from p(z); this gets mapped to a 2d offset $\delta = z\mathbf{w}$, where $\mathbf{w} \in \mathbb{R}^2$, which gets added to $\boldsymbol{\mu}$ to define a Gaussian $p(\mathbf{x}|z) = \mathcal{N}(\mathbf{x}|\boldsymbol{\mu} + \boldsymbol{\delta}, \sigma^2 \mathbf{I})$. By integrating over z, we "slide" this circular Gaussian "spray can" along the principal component axis \mathbf{w} , which induces elliptical Gaussian contours in \mathbf{x} space centered on $\boldsymbol{\mu}$. Adapted from Figure 12.9 of [Bis06].

Figure from (Murphy, 2020)

Probabilistic principle component analysis

A special case of the FA model where columns of **W** are orthonormal, $\Psi = \sigma^2 \mathbf{I}$ and $\mu = \mathbf{0}$ is called probabilistic principle component analysis (PPCA)

$$p(\mathbf{x}) = \int \mathcal{N}(\mathbf{x} \mid \mathbf{W}\mathbf{z}, \sigma^2 \mathbf{I}) \mathcal{N}(\mathbf{z} \mid \mathbf{0}, \mathbf{I}) d\mathbf{z} = \mathcal{N}(\mathbf{x} \mid \mathbf{0}, \mathbf{W}\mathbf{W}^T + \sigma^2 \mathbf{I})$$

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- Given an observed data set D = (x₁,..., x_n) we can estimate the model parameters W and σ²
- Compute the sample covariance matrix $\mathbf{S} = \frac{1}{n} \sum_{i=1}^{n} (\mathbf{x}_i \bar{\mathbf{x}}) (\mathbf{x}_i \bar{\mathbf{x}})^T$
- **•** Rewrite **S** using the eigenvector-eigenvalue decomposition $\mathbf{S} = \mathbf{U} \mathbf{\Lambda} \mathbf{U}^T$
- Optimal parameters are $\mathbf{W} = \mathbf{U}_L (\mathbf{\Lambda}_L \sigma^2 \mathbf{I})^{\frac{1}{2}}$ (upto arbitrary rotation) and $\sigma^2 = \frac{1}{D-L} \sum_{i=L+1}^{D} \lambda_i$

Probabilistic principle component analysis: posterior

- **>** Given **W** and σ^2 , we can use the PPCA and reduce the dimension of observed data **x**
- ▶ The embedding of **x** can be shown to have normal distribution

$$p(\mathbf{z} \mid \mathbf{x}) = \mathcal{N}(\mathbf{z} \mid \mathbf{M}^{-1} \mathbf{W}^{\mathsf{T}}(\mathbf{x} - \boldsymbol{\mu}), \sigma^2 \mathbf{M}^{-1}),$$

where $\mathbf{M} = \mathbf{W}^T \mathbf{W} + \sigma^2 \mathbf{I}$

▶ In the noise-free case of $\sigma^2 = 0$ the PPCA and PCA are directly comparable

Variational autoencoder

- ► The PPCA model is still a linear latent variable model
- ▶ We can extend the PPCA by defining the generative model to be nonlinear

$$p(\mathbf{z}) = \mathcal{N}(\mathbf{z} \mid \mathbf{0}, \mathbf{I})$$
$$p(\mathbf{x} \mid \mathbf{z}) = \mathcal{N}(\mathbf{x} \mid f(\mathbf{z}; \theta), \sigma^2 \mathbf{I}),$$

where $f(\cdot; \theta)$ is a nonlinear function which is typically parameterized by a deep neural network, such as the MLP

For nonlinear models we can compute neither the marginal likelihood p(x) nor the posterior of the latent representation p(z | x) analytically

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- For nonlinear models we can compute neither the marginal likelihood p(x) nor the posterior of the latent representation p(z | x) analytically
- Another key idea of the variational autoencoder (VAE) model is to use so-called inference network q(z | x, \u03c6) to approximate the intractable true posterior p(z | x)
- ▶ If we assume that the variationally approximated posterior has normal distribution, then

$$q(\mathbf{z} \mid \mathbf{x}, \phi) = \mathcal{N}(\mathbf{z} \mid f_{\mu}(\mathbf{x}, \phi), f_{\sigma^2}(\mathbf{x}, \phi)),$$

where $f_{\mu}(\cdot, \phi)$ and $f_{\sigma^2}(\cdot, \phi)$ are parametrized by deep neural network(s)

Variational autoencoder illustration



Figure from (Kingma and Welling, 2019)

Variational autoencoder: training

▶ It can be shown that maximizing the marginal likelihood of the data $p(\mathbf{x}) = \int p(\mathbf{x} | \mathbf{z})p(\mathbf{z})d\mathbf{z}$ corresponds to minimizing the Kullback-Leibler divergence from $q(\mathbf{z} | \mathbf{x}, \phi)$ to $p(\mathbf{z} | \mathbf{x})$

$$\mathrm{KL}(q(\mathsf{z} \mid \mathsf{x}, \phi) || p(\mathsf{z} \mid \mathsf{x})) = \int q(\mathsf{z} \mid \mathsf{x}, \phi) \log \frac{q(\mathsf{z} \mid \mathsf{x}, \phi)}{p(\mathsf{z} \mid \mathsf{x})} d\mathsf{z}$$

▶ This leads to so called evidence lower bound objective

$$\text{ELBO} = \underbrace{\mathbb{E}_{q(\mathbf{z}|\mathbf{x},\phi)} \log p(\mathbf{x} \mid \mathbf{z},\theta)}_{\text{reconstruction term}} - \underbrace{\text{KL}(q(\mathbf{z} \mid \mathbf{x},\phi) || p(\mathbf{z}))}_{\text{regularization term}},$$

which can be maximized using a reparametrization trick and stochastic gradient methods

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Deep generative models for single cell data

- scRNA-seq profiles contain both biological (mostly unknown) and technical (still poorly characterized) uncertainties
- Challenging to specify a well-motivated probabilistic model for the data
- ▶ VAE provides one principled modeling framework for complex scRNA-seq data

A variational autoencoder for scRNA-seq data

Variational autoencoder architecture with deep neural networks



Figure from (Lopez et al, 2019)

A variational autoencoder for scRNA-seq data

A probabilistic model for scRNA-seq data

$$\begin{aligned} z_n &\sim \operatorname{Normal}(0, I) \\ \ell_n &\sim \operatorname{LogNormal}(\ell_\mu, \ell_\sigma^2) \\ \rho_n &= f_w(z_n, s_n) \\ w_{ng} &\sim \operatorname{Gamma}(\rho_n^g, \theta) \\ y_{ng} &\sim \operatorname{Poisson}(\ell_n w_{ng}) \\ h_{ng} &\sim \operatorname{Bernoulli}(f_h^g(z_n, s_n)) \\ x_{ng} &= \begin{cases} y_{ng} & \text{if } h_{ng} = 0, \\ 0 & \text{otherwise.} \end{cases} \end{aligned}$$



Figure from (Lopez et al, 2019)

A variational autoencoder for scRNA-seq data illustration



Figure from (Lopez et al, 2019)

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